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<td><strong>Publication date</strong></td>
<td>2012</td>
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<td><strong>Publication information</strong></td>
<td>Frontiers in Physiology, 3 :</td>
</tr>
<tr>
<td><strong>Publisher</strong></td>
<td>Frontiers Media SA</td>
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<tr>
<td><strong>Item record/more information</strong></td>
<td><a href="http://hdl.handle.net/10197/5071">http://hdl.handle.net/10197/5071</a></td>
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<tr>
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<tr>
<td><strong>Publisher's version (DOI)</strong></td>
<td>10.3389/fphys.2012.00355</td>
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Crosstalk and signalling switches in mitogen-activated protein kinase cascades

Dirk Fey\textsuperscript{1}, David R Croucher\textsuperscript{1}, Walter Kolch\textsuperscript{1}, Boris N Kholodenko\textsuperscript{1,*}

Abstract

Mitogen-activated protein kinase (MAPK) cascades control cell fate decisions, such as proliferation, differentiation and apoptosis by integrating and processing intra- and extracellular cues. However, similar MAPK kinetic profiles can be associated with opposing cellular decisions depending on cell type, signal strength and dynamics. This implies that signalling by each individual MAPK cascade has to be considered in the context of the entire MAPK network. Here, we develop a dynamic model of feedback and crosstalk for the three major MAPK cascades; extracellular signal-regulated kinase (ERK), p38 mitogen-activated protein kinase (p38), c-Jun N-terminal kinase (JNK), and also include input from protein kinase B (AKT) signalling. Focusing on the bistable activation characteristics of the JNK pathway, this model explains how pathway crosstalk harmonises different MAPK responses resulting in pivotal cell fate decisions. We show that JNK can switch from a transient to sustained activity due to multiple positive feedback loops. Once activated, positive feedback locks JNK in a highly active state and promotes cell death. The switch is modulated by the ERK, p38 and AKT pathways. ERK activation enhances the dual specificity phosphatase (DUSP) mediated dephosphorylation of JNK and shifts the threshold of the apoptotic switch to higher inputs. Activation of p38 restores the threshold by inhibiting ERK activity via the PP1 or PP2A phosphatases. Finally, AKT activation inhibits the JNK positive feedback, thus abrogating the apoptotic switch and allowing only proliferative signalling. Our model facilitates understanding of how cancerous deregulations disturb MAPK signal processing and provides explanations for certain drug resistances. We highlight a critical role of DUSP1 and DUSP2 expression patterns in facilitating the switching of JNK activity and show how oncogene induced ERK hyperactivity prevents the normal apoptotic switch explaining the failure of certain drugs to induce apoptosis.

Manuscript length:
11 pages (not counting figures, tables, abstract and references),
7263 words + 105 inline equations + 14 display equations (by http://app.uio.no/ifi/texcount/),
9 figures,
5 tables.

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1 Introduction

A hallmark of cancer is dysregulation of pivotal cell fate decisions leading to aberrant proliferation and reduced apoptosis (Hanahan and Weinberg, 2011). Healthy cell fate decisions depend on a proper sensing of the cell’s intra- and extracellular environment in a process called signal transduction (Kholodenko et al., 2010). The signals are sensed by receptors that bind their cognate extracellular ligands, resulting in conformational changes that trigger the formation of multi-protein complexes and subsequent activation of GTPases and kinases (Lemmon and Schlessinger, 2010). Hereby, one receptor usually activates several downstream pathways. Main transducers are MAPK cascades, which consist of a linear array of three kinases where a GTPase activates a MAPK kinase kinase (MAPKKK; MAP3K), which phosphorolyses and activates a MAPK kinase (MAPKK; MAP2K), which in turn activates a MAPK that delivers the main pathway output by phosphorylating multiple substrates (Dhillon et al., 2007; Kolch, 2005). MAPKs and MAP2Ks are activated by dual phosphorylation, which can confer switch-like properties to the activation kinetics (Kholodenko, 2000). Sometimes a MAP4K is intercalated between the GTPase and the MAP3K. A particular cell fate cannot be attributed to the activity of a single protein in isolation, but rather depends on the context, including the temporal patterns of activation and the regulatory feedback structures within the signalling network (Kholodenko et al., 2010; Nakakuki et al., 2010; Kholodenko, 2006). Because of this complexity, the function of cellular signalling often eludes a naive intuitive understanding, thus calling for the use of mathematical modelling and analysis (Kitano, 2010; Ireton et al., 2009; Kitano, 2002). Whereas others approach the problem from a less mechanistic viewpoint using regression (Miller-Jensen et al., 2007) or Boolean and semi-logic models (Saez-Rodriguez et al., 2011, 2009), we focus on dynamic models using ordinary differential equations.

Dynamic modelling has played a key role in understanding how signalling via the ERK cascade regulates cell fate (Kholodenko et al., 2010; Sturm et al., 2010). A classic example is growth factor signalling in Rat Pheochromocytoma (PC12) cells, where treatment with epidermal growth factor (EGF) or nerve growth factor (NGF) activates the same signalling cascade (the RAF/MEK/ERK cascade) but has different effects on cell fate. EGF causes transient activation of ERK and proliferation due to negative feedback, whereas NGF causes sustained ERK activation and differentiation due to positive feedback (von Kleischheim et al., 2009; Santos et al., 2007). Similarly, the stress activated MAPKs JNK and p38 mediate diverse cellular responses. For example, growth factor induced, transient activation of JNK promotes cell survival and proliferation, whereas stress induced, prolonged JNK activity promotes growth arrest and cell death (Ventura et al., 2006). However, the mechanistic details of how this switch is generated and the factors determining the shift from proliferative to apoptotic JNK signalling are poorly understood, and mathematical modelling and analysis is largely lacking for stress activated kinases (Wagner and Nebreda, 2009; Bagowski and Ferrell, 2001).

Here, we provide a dynamic model of feedback and crosstalk for the three major MAPKs (ERK, p38, JNK) and protein kinase B (AKT) signalling. The model incorporates mechanistic details of positive feedback from JNK to its own MAP3Ks and negative crosstalk from and to other pathways. Using mathematical analysis, the model is used to decipher how JNK switches from proliferative to apoptotic signalling and how that switch is regulated by pathway crosstalk.

2 Results

We present a dynamic model of multiple MAPK cascade interactions featuring a JNK positive feedback loop that generates a proliferative-apoptotic switch. Further, we present a detailed analysis of factors controlling the dynamic properties of the JNK switch, with a particular focus on feedback loops and crosstalk.

2.1 Nominal model of MAPK interactions

Although MAPK signalling cascades have been studied extensively, the connectivity of MAPK systems is not completely understood. MAPKs feature several isoforms, a high number of inputs in the form of different GTPases and protein kinases, several scaffolding proteins that channel incoming signals into different pathways and a variety of phosphatases that modulate MAPK activation dynamics. Thus, depending on the expression and activity states of these proteins, MAPK connections change between cell types and in response to pathological aberrations. In order to analyse the kinetic behaviour and regulation of MAPK cascades we constructed a model which represents a core network of MAPK interactions based on the available literature. The topology of this model is depicted in Fig. 1.

Generally, MAPK systems are arranged in three tiered cascades consisting of MAPKs (lowest tier), MAPK kinases (MAP2Ks, second tier) and MAP kinases kinases (MAP3Ks, top tier). Activation of the kinases in each tier is modelled with double phosphorylation cycles as described in Material and Methods (Sec. 5), in which the upstream kinase acts as the enzyme catalysing the phosphorylation and therefore activation of the downstream kinase. Complementing the classical cascades, the model features several crosstalks and feedbacks (Table 1). First, JNK phosphorolyses and activates its own MAP3K (Furuhata et al., 2009; Schachter et al., 2006), generating a positive feedback loop. Second, p38 inhibits ERK activity by enhancing MEK dephosphorylation either via transcriptional upregulation or phosphorylation of protein phosphatase 2 (PP2A) (Junttila et al., 2008; Grethe and Forrn-Ares, 2006; Liu and Hofmann, 2004; Li et al., 2003; Westermann et al., 2001). Third, ERK inhibits JNK via induction of dual specificity phosphatases (DUSPs) catalysing the dephosphorylation of JNK (Monick et al., 2006; Paumelle et al., 2000). Finally, AKT inhibits JNK...
Figure 1: Scheme of the nominal MAPK interaction model. For simplicity of illustration, the double phosphorylation of MAP(K) kinases are depicted in single steps and the three inactive forms of ASK/MLK and MKK4/7 are lumped into one component. $U_i$ denote inputs that are modelled as time-dependent functions (not modelled with differential equations). Black: nominal cascades; Blue: positive feedback from JNK to its own MAP3Ks; Red: negative crosstalk from AKT to JNK signalling; Purple: negative crosstalk from p38 to ERK signalling only occurring in non-transformed cells. Lower- and upper-case letter indicate mRNAs and proteins, respectively. Single and double asterix indicate single- and double-phosphorylated active forms, respectively.

2.1.1 JNK positive feedback

Several studies support the idea of a JNK positive feedback loop on the systems level. For example, JNK positive feedback was critical for a proper stress response of Xenopus oocytes (Bagowski and Ferrell, 2001). In mammalian cells, JNK exhibited all-or-none responses on the single cell level after treatment with anisomycin or sorbitol (Bagowski et al., 2003), and a positive feedback loop was suggested (Bagowski et al., 2003; Xiong and Ferrell, 2003). On the population level, these all-or-none responses manifest highly ultrasensitive behaviour with apparent Hill coefficients as high as 9 or 10 (Table 2), which is consistent with the presence of a positive feedback loop, which increases the degree of ultrasensitivity (Bagowski et al., 2003).

The literature contains considerable evidence supporting the existence of positive feedback from JNK to its own MAP3Ks, in particular to mixed image kinases (MLK) and apoptosis regulated kinases (ASK) (Furuhata et al., 2009; Schachter et al., 2006; Ventura et al., 2004; Phelan et al., 2001; Xu and Cobb, 1997). For example, in HEK 293, Hela and MCF-7 cells, JNK phosphorylated MLK3 directly at sites in the COOH-terminal region, which resulted in the redistribution of MLK3 to triton-insoluble membrane microdomains, increased phosphorylation of the activation loop and increased MLK3 activity (Schachter et al., 2006). Similarly in COS7 cells, JNK phosphorylated the C-terminal domain of MLK2, which was required for MLK2-induced apoptosis (Phelan et al., 2001). Further, JNK phosphorylated a MEKK1 fragment in vitro and coimmunoprecipitated with MEKK1 in HEK 293 cells Xu and Cobb (1997). MEKK1 is a MAP3K for the JNK pathway, which depending on its phosphorylation status, also can act as a scaffold for the MEKK1-MKK4-JNK pathway (Gallagher et al., 2002).

Another, more indirect route of JNK feeding back to its own MAP3Ks involves the production of reactive oxygen species (ROS). In fibroblasts, JNK produced ROS after TNF treatment, in a process that did not involve gene transcription and was inhibited by NF-κB (Ventura et al., 2004). Interestingly, several signalling pathways connect ROS to JNK activation, suggesting a JNK-ROS positive feedback loop (Shen and Liu, 2006). ASK1 in particular is readily activated by ROS, whereby ROS induces the disso-

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<th>Stimulus</th>
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<tr>
<td>Sorbitol</td>
<td>HeLa 9</td>
</tr>
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<td>Anisomycin</td>
<td>HEK293 8</td>
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<td>Jurkat 4</td>
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Table 2: Ultrasensitivity of the JNK response to stress in mammalian cell populations (Bagowski et al., 2003).
which, in turn, activated ASK1 (Furuhata et al., 2009). Where JNK activity produced hydrogen peroxide (H₂O₂) been reported in WEHI-231 mouse B lymphoma cells, in fact, such a ROS dependent positive feedback loop has

<table>
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<th>Interaction</th>
<th>Mechanism</th>
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<tr>
<td>JNK → ASK1</td>
<td>oligomerization and auto-phosphorylation of its activation loop (Shen and Liu, 2006; Goldman et al., 2004; Saitoh et al., 1998). In fact, such a ROS dependent positive feedback loop has been reported in WEHI-231 mouse B lymphoma cells, where JNK activity produced hydrogen peroxide (H₂O₂) which, in turn, activated ASK1 (Furuhata et al., 2009).</td>
<td>via JNK induced ROS production (WEHI-231) direct JNK mediated phosphorylation (HEK 293, Hela, MCF-7)</td>
<td>Furuhata et al. (2009)</td>
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<td>→ MLK3</td>
<td>phosphorylation</td>
<td>only in non-immortalised, non-transformed cells</td>
<td>Schachter et al. (2006)</td>
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<tr>
<td>p38 ↓ ERK</td>
<td>upregulation of PP2A</td>
<td>H2O2-sensitive</td>
<td>Junittila et al. (2008); Grethe and Pörn-Ares (2006); Li et al. (2003); Westermarck et al. (2001); Liu and Hofmann (2004)</td>
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<tr>
<td>ERK ↓ JNK</td>
<td>induction of DUSP4 &amp; DUSP16</td>
<td>MDCK epithelial cells, human alveolar macrophages</td>
<td>Paunelle et al. (2000); Monick et al. (2006)</td>
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<td>AKT ↑ ASK1</td>
<td>phosphorylation at S83</td>
<td>HEK293, L929</td>
<td>Kim et al. (2001)</td>
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<tr>
<td>↑ MLK3</td>
<td>phosphorylation at S674</td>
<td>HepG2</td>
<td>Barthwal et al. (2003)</td>
</tr>
<tr>
<td>↑ MKK4</td>
<td>phosphorylation at S78</td>
<td>HEK293T</td>
<td>Park et al. (2002)</td>
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<tr>
<td>JNK ↑ ERK, p38</td>
<td>induction of DUSPs via Jun gene transcription, DUSP2 is the speculative isoform assumed in the model.</td>
<td>what DUSP isoforms are involved is unclear; DUSP1,4,6 were not detected in COS7</td>
<td>Peng et al. (2009); Junittila et al. (2008); Stepmiak et al. (2006); Shen et al. (2003); Black et al. (2002); Chu et al. (1996)</td>
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Table 1: Crosstalks and feedbacks in the nominal MAPK interaction model. The list comprises core-interactions which are supposedly conserved between cell lines, but with the p38-ERK crosstalk being restricted to non-transformed cells. The comments column indicates the experimental system (cell lines) used to identify the links.

The molecular mechanisms of MAP3K activation are quite complex. For example, MLK3 activation involves GTPases binding, translocation to the membrane, dimerisation and oligomerisation and activation loop phosphorylation of MLK3 at Thr2277 and Ser281 (Schachter et al., 2006). Neglecting this complexity, and in concordance with earlier models in the literature, we model the activation of MAP3Ks as a phosphorylation process catalysed by its inputs (Kholodenko et al., 2010; Kholodenko, 2000). In the model, different JNK-MAP3Ks are lumped into one ASK/MLK component. The activation of this component is modelled as a double phosphorylation cycle with two inputs, representing the activity of upstream GTPases and active JNK (see Fig. 1). Although our model simplifies the involved molecular events, it captures the main feature of MAP3K activation, namely the phosphorylation of two conserved residues in the activation loop.

### 2.1.2 p38 inhibits ERK signalling in non-transformed cells

Generally, ERK activity promotes survival. The suppression of this activity by p38 is critical for induction of apoptosis in nontransformed cells and PP2A mediates this effect (Junittila et al., 2008). In particular, p38 mediated dephosphorylation of MEK was necessary for arsenite induced apoptosis in human skin fibroblasts (HSF) and rat primary neurons (CGN), but not in transformed and tumourigenic cell lines (HeLa, Jurkat, K562, HT-1080, WM266-4, A2058) (Li et al., 2003). Further, PP2A mediated this p38-MEK negative crosstalk and was required for both cytokine and stress induced apoptosis in human endothelial cells and rat cardiac ventricular myocytes, respectively (Grethe and Pörn-Ares, 2006; Liu and Hofmann, 2004).

How p38 regulates PP2A is uncertain. PP2A is a heterotrimer composed of a scaffold, a catalytic subunit and different regulatory subunits. Its catalytic activity can be regulated on several levels, including assembly of the heterotrimer with different regulatory subunits, and both phosphorylation or methylation of the catalytic subunit (Nguyen et al., 2012; Jaassens and Goris, 2001). Because the mechanism by which p38 upregulates PP2A activity is uncertain, and because the system dynamics depend on this mechanism, our model implements two possibilities, each on opposite ends of the dynamic spectrum: slow upregulation via gene transcription of the regulatory subunit, and fast activation via phosphorylation of the catalytic subunit.

#### 2.1.3 ERK inhibits JNK

ERK signalling strongly induces several DUSPs, some of which negatively regulate JNK activity. For example, DUSP4 is readily induced in response to several growth factors (Cagnol and Rivard, 2012; Legewie et al., 2008) and stabilisation of DUSP16 by ERK mediated phosphorylation at Ser-446 was observed in both COS-7 (fibroblasts)
Although the exact mechanism is poorly understood, and elevated expression of DUSP1, DUSP4 and DUSP6 could not be detected in COS-7 cells expressing active MLK3, DUSPs were suggested as potential mediators of the JNK-ERK crosstalk (Junttila et al., 2008; Shen et al., 2003). JNK can also inhibit p38, as JNK activity inhibited both ERK and p38 signalling in mouse cardiomyocytes (Peng et al., 2009) and c-Jun deficient hepatocytes showed increased phosphorylation of p38 (Stepniak et al., 2006). The JNK – ERK/p38 crosstalk may involve a p53-DUSP2 dependent pathway, as the c-Jun mediated inhibition of p38 observed in hepatocytes was p53 dependent (Stepniak et al., 2006) and DUSP2 was identified as a transcriptional target of p53 in mouse embryonic fibroblast and breast cancer cell lines (Yin et al., 2003). Further, DUSP2 was shown to dephosphorylate ERK and p38 in NIH3T3 and HeLa cells (Chu et al., 1996) and was implicated in the inactivation of ERK2 during p53-dependent apoptosis in breast and colon cancer cell lines (Yin et al., 2003; Dickinson and Keyse, 2006). Based on these data and neglecting p53 as possible intermediate, the dynamic model features JNK induced expression of DUSP2 mRNA and protein and DUSP2-catalysed dephosphorylation of ERK and p38.

2.2 Dynamics of the core network

Based on the model structure in Fig. 1 a dynamic model of MAPK interactions can be derived (Kholodenko et al., 2010; Kholodenko, 2006). For detailed introductions to dynamic modelling of cellular systems we refer to Iglejas and Ingalls (2009); Aldridge et al. (2006) and, in particular with regards to ERK/MAPK signalling, Orton et al. (2005); Kolch et al. (2005); Kholodenko (2006). A successful modelling strategy keeps the model simple, yet biologically relevant and capable of meaningful predictions. To that end, the developed model contains several biologically reasonable assumptions, simplifications and generalisations as explained in Material and Methods (Sec 5).

In particular, the model lumps isoforms and kinases that share the same upstream activators and downstream substrates into a single component wherever possible (Fig. 1). Crucially, the adopted simplifications preserve the network’s feedback and crosstalk structures, reduce the risk of over-parameterisation and facilitate the mathematical analysis of the model.

2.2.1 MAPK dynamics in response to growth factors or stress

The developed model reflects the current understanding of how the p38 and JNK systems respond to stress (Junttila et al., 2008), and is consistent with earlier MAPK models in the literature, which, albeit not concerned with p38 and JNK, featured growth factor induced ERK signalling (Kholodenko et al., 2010; Nakakuki et al., 2010; von Kriegsheim et al., 2009). Fig. 2 presents an overview of the system dynamics, illustrating how our model responds to growth factor and stress signals. Generally speaking, growth factors predominantly activate ERK and

2.1.4 AKT inhibits JNK signalling

In response to several growth factors and insulin AKT mediates survival signalling, in part, by phosphorylation and inhibition of apoptotic proteins (Hers et al., 2011). Active AKT phosphorylates inhibitory sites of JNK upstream kinases at both the MAP2K and MAP3K level (see Table 1). On the MAP3K level, phosphorylation of ASK1 at Ser 83 by AKT reduced JNK activity in response to oxidative stress and serum starvation, and decreased ASK1 dependent apoptosis in HEK 293 and L929 cells (Kim et al., 2001). Similar results were obtained in HepG2 cells, where insulin induced AKT activity led to phosphorylation of MLK3 at Ser 674 (Barthwal et al., 2003). On the MAP2K level, AKT phosphorylated MKK4 at Ser 78 in response to insulin or constitutively active AKT, which reduced JNK activity and anisomycin induced apoptosis in HEK 293T cells (Park et al., 2002).

The dynamic model does not distinguish different MAP3Ks and MAP2Ks in the JNK pathway, but features combined ASK/MLK and MKK4/7 components, as MAP3Ks and MAP2Ks respectively. We model both components taking a domain oriented approach (Conzelmann et al., 2008; Kiyatkin et al., 2006; Borisov et al., 2006, 2005) and assuming that the phosphorylation processes at the activation loop and the inhibitory site are independent, as described in detail in Material and Methods.

2.1.5 JNK inhibits ERK and p38

JNK can inhibit ERK on several levels, involving both indirect upstream mechanisms and direct dephosphorylation upon the transcriptional induction of DUSP expression (Junttila et al., 2008). The model of direct ERK dephosphorylation via transcriptional DUSP induction is supported by two studies showing that the JNK-ERK crosstalk is at least partially independent of the ERK upstream kinases MEK and Raf. First, v-Jun transcriptional activity reduced both basal and growth factor induced ERK phosphorylation at least partially independent of Raf (Black et al., 2002). Second, JNK activity induced by ceramide and TNF-α blocked growth factor stimulated ERK phosphorylation, and this inhibition required c-Jun transcriptional activity but did not involve MEK (Shen et al., 2003).
JNK, and also AKT, albeit to different extents. The activation dynamics may be sustained or transient, depending on type and context of the stimulation (Nakakuki et al., 2010; von Kriegsheim et al., 2009). For example, PC12 cells exhibit sustained ERK activation in response to NGF, whereas EGF causes transient ERK dynamics due to the activation of several negative feedback loops (von Kriegsheim et al., 2009; Douville and Downward, 1997; Marshall, 1995). These negative feedbacks act upstream of the ERK cascade, at the level of growth factor receptors and their adaptors, and result in a transient input signal for the MAPK system. We can model these transient effects taking a modular approach in which the inputs are modelled by time dependent functions (Nakakuki et al., 2010). Hereby, a step input corresponds to a sustained signal, whereas a pulse input, which drops back to the low basal level after a certain, relatively short time period, corresponds to a transient signal. Fig. 2 panels A and B show that in response to growth factors, the ERK dynamics qualitatively follow the input signal, whereas JNK responds transiently, and only to growth factors that do not activate AKT. Stress signals predominantly activate p38 and JNK and sometimes ERK, but to a much weaker extent. Figure 2C shows that the JNK response to stress is sustained for both transient and sustained stress inputs.

2.2.2 Dynamics of stress induced apoptosis in the presence of growth factors

The core model reflects the current understanding of JNK dependent apoptosis induction. In Junttila et al. (2008) a conceptual model was proposed, in which PP2A mediated suppression of ERK by p38 is critical for JNK mediated apoptosis. The idea is that stress induced activation of p38 suppresses the normal ERK activity of proliferating and differentiating cells and, subsequently, this loss of ERK activity sensitises the cells to JNK mediated apoptosis. Our dynamic model is a mathematical representation of this idea amenable to theoretical analysis. Indeed, simulating the dynamic model with a step input of stress signals

\[
u_{p38}(t) = u_{JNK}(t) = \begin{cases} 
1 & \text{for } t > 0 \\
0 & \text{otherwise}
\end{cases}
\]

in the presence of a constant mitotic signal \(u_{ERK}(t) = 1\) mimics the data and sequence of events described in Junttila et al. (2008). Hereby, the qualitative behaviour is largely independent of the exact mechanism of PP2A activation. For both mechanisms, either transcriptional up-regulation of PP2A or its activation by p38-induced phosphorylation, the JNK switch occurs at a 3-6 h delay following the apoptotic stimulus (Fig. 3). The delay is largely determined by the strength of the p38-PP2A crosstalk. Decreasing the expression rate of PP2A in the transcriptional upregulation model or decreasing the catalytic activity of p38 towards PP2A in the model of phosphorylation induced PP2A activation increases the time of JNK activation (Fig. 3, dashed lines). In the following, we dissect the MAPK interaction network generating these com-

Figure 2: Dynamics of the core network in response to different stimuli. (A) Form of input signals. Solid: sustained input, dashed: transient input. (B-D) System responses for different stimuli. (B) Growth factor strongly stimulating the ERK and JNK inputs, but only weakly the AKT and p38 inputs: (C) Growth factor strongly stimulating the ERK and JNK and AKT inputs, but not the p38 input. (D) Stress signal strongly stimulating the p38 and JNK inputs, weakly stimulating the ERK input and not stimulating the AKT input.

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<th>(\hat{u}_{ERK})</th>
<th>(\hat{u}_{p38})</th>
<th>(\hat{u}_{JNK})</th>
<th>(\hat{u}_{AKT})</th>
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<tr>
<td>(B)</td>
<td>1.00</td>
<td>0.15</td>
<td>1.00</td>
<td>0.20</td>
</tr>
<tr>
<td>(C)</td>
<td>1.00</td>
<td>0.15</td>
<td>1.00</td>
<td>0.50</td>
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<tr>
<td>(D)</td>
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<td>1.00</td>
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where \(\hat{u}_i\) (\(i = \text{ERK, p38, JNK}\)) denotes the maximal value of the ERK, p38 and JNK input (see panel A).
plex dynamics, by providing a systems level analysis of these interactions.

2.3 Analysing feedback structures

MAPK systems exhibit complex dynamic behaviour, depending on the topology of feedbacks and kinetic parameters. Although the parameters are important for the responses observed, the network topology in terms of feedback loops determines what qualitative behaviours are possible (Kholodenko, 2006). Generally speaking, negative feedback can generate (sustained) oscillations, whereas positive feedback can generate bistability. Bistability is thought to be important in cell fate decisions, as it is characterised by hysteresis and can generate irreversible switches (Xiong and Ferrell, 2003; Novak and Tyson, 1993). An example is the caspase system, where positive feedback generates an irreversible switch between two stable steady states; an off-state corresponding to survival and an on-state corresponding to apoptosis (Eissing et al., 2004). As the model features a JNK positive feedback loop, we sought to determine under which conditions the system exhibits bistability.

2.3.1 Positive feedback and bistability of the JNK module

A convenient tool for analysing bistability is the loop breaking approach (Angeli et al., 2004). Loop breaking is a graphical analysis tool consisting of two steps. First, break the feedback loop and plot the input/output (i/o) relationship in steady state for the open loop system. The resulting curve is called the steady state characteristic of the open loop. Second, close the loop graphically by plotting a straight line through the origin, whereby the slope of the line represents the inverse strength of the feedback. For example, unitary feedback \( u = y \) is represented by a straight line of slope one. The intersection points of the two lines represent the steady states of the closed loop system. In order to assess the stability of the steady states (similarly to nullclines in classical phase plane analysis), two technical prerequisites have to be satisfied; existence of a well-defined i/o characteristic and monotonicity, both of which can be satisfied for simplified MAPK cascades (not exhibiting negative feedback). For details we refer to the original literature (Angeli et al., 2004).

Zooming into the JNK module of the nominal model, the loop breaking approach reveals that the JNK system is indeed bistable for a wide range of feedback strengths (Fig. 4). Note that this result does not depend on the exact parameters, but rather the sigmoidal input-output characteristic of the JNK cascade. In this analysis, the feedback strength corresponds to the catalytic activity of JNK to phosphorylate ASK/MLK. More precisely, let \( x_0, x_1 \) and \( x_2 \) denote the concentrations of non-, single- and double-phosphorylated ASK/MLK, accordingly, and let further, \( k_b \) be the catalytic activity of the upstream ASK/MLK input \( u \) and \( k_f \) the catalytic activity of active JNK \( y \), then

\[
e_{\text{phos}, i} = \frac{(k_f u + k_b y) x_i}{K_d + x_0 + x_1}, \quad i = 0, 1
\]  

(1)

describes the rate of ASK/MLK phosphorylation. Hereby, a feedback strength of 100% corresponds to \( k_b = k_f \), i.e. equal catalytic activities of input and JNK. Figure 4 shows that for typical values of MAPK phosphorylation and dephosphorylation parameters (Kholodenko et al., 2010; Nakakuki et al., 2010; Kholodenko, 2000; Huang and Ferrell, 1996) the strength of the positive feedback can be reduced to less than 40% before bistability is lost.

The point of transition from monostable to bistable behaviour is called pitchfork bifurcation and depends not only on the feedback strength, but also the upstream input. Recall that ASK/MLK are not only phosphorylated by JNK feedback, but also upstream inputs (such as GTPase recruited kinases or MAP4Ks). For the graphical analysis, assuming a constant input corresponds to a right-shift of the feedback line, whereby the value of the right-shift indicates the strength of the input (Fig. 4). Applying a feedforward input to a feedback system that was originally not bistable (due to a low feedback gain), can push it into a bistable regime and beyond. Hereby the system moves from a monostable-off regime through a bistable regime to a monostable-on regime (Fig. 4). Further, a combined analysis of feedback and feedforward input shows that even for appropriate inputs, bistability is lost if the feedback strength is too low. In fact, in order for a bistable regime to exist, the inverse of the feedback strength has to be smaller than the maximal slope of the sigmoidal i/o characteristic (Fig. 4).

2.3.2 Negative feedback via dual specificity kinases

The core model depicted in Fig 1 does not contain negative feedback within the JNK module. However, negative feedback is not uncommon in MAPK cascades and is often context dependent. For instance, ERK possesses several negative feedback loops that are activated in a stimulation dependent manner in response to EGF, but not NGF or HRG (Nakakuki et al., 2010; von Kriegsheim et al., 2009; Santos et al., 2007). With regard to JNK signalling, several DUSPs exhibit catalytic activity towards JNK and may be induced by active JNK (Boutros et al., 2008; Dickinson and Keyse, 2006). One such example is DUSP1 (Bokemeyer et al., 1996). Therefore, we explored the possibility of DUSP1 mediated negative feedback in the JNK module. Note that the system is not monotonic because of the negative feedback. Consequently, graphical analysis using loop breaking cannot assess the stability of the steady states but only their existence and should be complemented by local stability analysis or simulations.

Negative feedback to upstream components of JNK can decrease ultrasensitivity and lead to oscillations (data not shown, see for example Kholodenko et al. (2010) for a general treatment). In contrast, DUSP1 mediated, slow negative feedback can disable the bistable switch generated by
Figure 3: Trajectories of the core model mimic the sequence of events Juntila et al. (2008) that occur in response to a stress stimulus $u_{p38}(t) = u_{JNK}(t) = 1$ for $t > 0$ and the presence of a constant mitotic signal $u_{ERK}(t) = 1$ for all $t$. D2 and D4/16 denote DUSP2 and DUSP4/16, which mediate the JNK $\leftrightarrow$ ERK, p38 and ERK $\leftrightarrow$ JNK crosstalks, respectively (see also Table 1). The qualitative behaviour is independent of the mechanistic details implementing p38-PP2A interaction. Two mechanism are shown: (A,B) p38 induces PP2A gene expression, whereby the red line in panel B represents the total level of PP2A protein, (C,D) p38 phosphorylates PP2A, whereby the red line in panel D represents phosphorylated PP2A. (A-D) The timing of JNK activation depends on the strength of PP2A upregulation: Solid lines indicate PP2A levels comparable to those of the other phosphatases. Dashed lines indicate reduced levels of PP2A expression, which delays JNK activation.

Figure 4: Analysis of JNK positive feedback using the loop breaking approach. Here, $g$ denotes the feedback strength, i.e. the ratio $g = k_b/k_f$ in (1). (A-D) Solid, blue lines represent the steady-state characteristic of i/o-system. Dashed lines indicate different feedback configurations, whereby the slope represents the feedback strength and the right-shift measured from the origin the feedforward stimulus. (A) Illustration of the loop breaking approach (for a detailed explanation see main text) (B) Depending on the feedback strength, the JNK system exhibits monostable or bistable behaviour ($u=0$). (C) Simultaneous feedback and feedforward stimulation can push the system from a monostable-off (white), through a bistable (light green), to a monostable-on (light red) regime. (D) No bistable behaviour is possible for feedback gains lower than the inverse of the maximal slope of the i/o characteristic.
the fast positive feedback loop (Fig. 5). Depending on the
relative feedback strength, a transiently bistable regime
exists, in which the JNK system responds with prolonged
activity in response to a short lived stimulus. Hereby, the
positive feedback maintains the on-state after the input
subsides, but only until the slow negative feedback takes
effect, diminishing the (initial) i/o characteristic of the
system, at which point JNK switches off (Fig. 5).

2.4 Regulation of the JNK apoptotic switch by
crosstalk

Mitogenic and survival signals regulate the JNK apopto-
totic switch through crosstalk occurring on several levels
(Fig. 1). We can distinguish two mechanisms; firstly, inhib-
ition of JNK activation by phosphorylation of upstream
JNK kinases at inhibitory residues and secondly enhanced
JNK dephosphorylation by upregulation of phosphatases.
The first mechanism is mediated by AKT, a classical me-
diator of survival signalling. The second mechanism, is
mediated via ERK, a classical mediator of proliferative
and differentiation signalling.

In the following, we use the nominal model to decipher
how MAPK crosstalk integrates different mitotic, survival
and stress signals, particularly focusing on the bistable
switch. First, we model the system with constant mi-
totic and survival inputs $u_{\text{ERK}}(t) = \hat{u}_{\text{ERK}}, u_{\text{AKT}}(t) =
\hat{u}_{\text{AKT}}$ and let the trajectories relax to steady state. Then
we apply stress stimuli in the form of step inputs

$$u_{p38}(t) = \begin{cases} 0 & \text{for } t < 0 \\ \hat{u}_{p38} & \text{otherwise,} \end{cases}$$

$$u_{JNK}(t) = \begin{cases} 0 & \text{for } t < 0 \\ \hat{u}_{JNK} & \text{otherwise.} \end{cases}$$

It is useful to define the switching threshold as the value of
$\hat{u}_{JNK}$ at which JNK switches from the off to the on state.

2.4.1 How AKT controls the JNK switch

AKT signalling affects the switching threshold and reg-
ulates the JNK on-state (Fig. 6). Increasing AKT ac-
tivity decreases the value of the JNK on-state. Whereas
bistable behaviour is still possible for moderate AKT sig-
nalling, strong AKT signalling abrogates the JNK apop-
totic switch and permits only moderate, proliferative JNK
activity.

The regulation of the JNK switch by AKT does not de-
pend on the exact topology of the crosstalk, as isolated
crosstalk at the MAP2K or the MAP3K levels exhibits
similar control patterns (Fig. 6). One slight difference
is that crosstalk on the MAP3K level has slightly more
impact on the switching threshold and admits some sensi-
tivity of the proliferative regime with respect to the JNK
input, meaning that changing the JNK input changes the
level of JNK activity (orange and red curves in Fig. 6B). In
contrast, the curves resulting from the MAP2K crosstalk
are almost flat, meaning that changing the JNK input
does not affect JNK activity other than switching it on or
off (Fig. 6C). Thus, the MAP2K crosstalk model quickly
saturates for all levels of AKT activity, after which chang-
ing the input has no effect on the output. In contrast, the
MAP3K crosstalk model does not saturate when AKT ac-
tivity is high, and after crossing a certain threshold, JNK
responds linearly to changes in the input.

2.4.2 How ERK and p38 control the JNK switch

Increasing the input of ERK signalling shifts the
switching-threshold towards higher JNK inputs, but has
little effect on the value of the on-state (Fig. 7). Crucially,
no intermediate JNK activation is possible, JNK is either
off or on. Further, the strength of apoptotic JNK signalling
once activated, is independent of the ERK input.

The regulation of the JNK switch by ERK depends
on the p38-ERK crosstalk, $p38 \rightarrow PP2A \rightarrow ERK$. No-
mal, nontransformed cells can initiate the JNK apoptotic
switch depending on the level of PP2A expression and p38
signalling. Here, increasing $p38$ pathway activation and
PP2A expression increases the regime of tolerable ERK
stimuli for which JNK inputs can initiate the apoptotic
switch (Fig. 7B). In contrast, in transformed and tumouri-
genic cells, lacking p38-ERK crosstalk, even very moder-
ate stimulation of the ERK pathway prevents the JNK
apoptotic switch (Fig. 7C).

2.4.3 How different DUSP-mediated crosstalk
patterns shape JNK dynamics

DUSPs are important regulators of MAPK activities. The
main function of DUSPs is to dephosphorylate the ac-
tivation loop of MAPKs, often with overlapping sub-
strate specificity (Bermudez et al., 2010; Boutros et al.,
2008; Dickinson and Keyse, 2006). Importantly, several
DUSPs are in turn regulated by MAPKs and induced in
response to mitotic, differentiation and stress signals.
Therefore, the regulation of DUSPs can occur on several
levels including the regulation of DUSP phosphatase ac-
tivity, substrate specificity, protein stability and gene ex-
pression (Boutros et al., 2008; Dickinson and Keyse, 2006).

The resulting feedback and crosstalk structures are com-
plex, and lack a complete understanding. Owing to this
complexity, we formulated several models based on re-
ported DUSP specificities in the literature (Tab. 3, Pat-
terson et al. (2009); Boutros et al. (2008); Dickinson and
Keyse (2006)). By focusing on MAPK induced gene tran-
scription and neglecting the complexity of posttranscrip-
tional DUSP regulations, these models are used to analyse
the effects of different crosstalk structures.

DUSP1 expression can be induced by active $p38$ and
JNK depending on the cell context (Tab. 3), and it is
often upregulated in cancer. The JNK-induced DUSP1
expression and the resulting negative feedback onto JNK
was already analysed in Sec. 2.3.2, Fig. 5. In this section
we focus on the $p38$ induced DUSP1 expression and re-
sulting $p38 \rightarrow p38/JNK$ crosstalk. We have already seen
in Sec. 2.4.2, Fig. 7 that $p38 \rightarrow ERK \rightarrow JNK$ crosstalk is
a critical regulator of the JNK switch. However, the core
model also features $JNK \rightarrow ERK/p38$ crosstalk mediated
Figure 5: Modulation of the bistable switch by negative feedback. (A) Scheme of the extended JNK model, in which active JNK induces the expression of DUSP1 mRNA (mD) and protein (pD).

Figure 8 shows the responses for different crosstalk patterns to step inputs of stress signals,

\[ u_{p38}(t) = u_{JNK}(t) = \begin{cases} 1 & \text{for } t > 0 \\ 0 & \text{otherwise} \end{cases} \]

in the presence of a constant mitotic signal \( u_{ERK}(t) = 1 \) for all \( t \). We can distinguish two qualitatively different behaviours, irrespective of the presence or absence of ERK feedback (mediated by DUSP4 or DUSP5/6). The models in the first group do not feature p38 induced DUSP1 expression, and DUSP2 deletion in these models has little effect on the JNK activation dynamics and the JNK switch (Fig. 8A,B). Within this group, model A is the core model, but model B also includes DUSP4 mediated negative feedback to ERK; ERK → ERK/JNK (Tab. 3), resulting in accelerated JNK activation dynamics (Fig. 8B).

In contrast, the models in the second group feature p38 induced DUSP1 expression, and deletion of DUSP2 in these models abrogates the JNK switch, resulting in reduced, moderate JNK activity (Fig. 8C-E). In addition to the core interactions, model C includes this p38 induced DUSP1 expression, which slightly delays the JNK activation dynamics, but does not obliterate the JNK switch (Fig. 8C). However, deleting DUSP2 in model C abrogates the JNK switch and results in only moderate JNK activity (Fig. 8C). Summarising, these models predict that abrogation of JNK dependent apoptosis requires both p38 induced expression of DUSP1 and downregulation or deletion of DUSP2. Adding ERK negative feedback mediated by DUSP4 (model D) or DUSP4 and DUSP5/6 (model E) to model C does not alter the JNK dynamics or the behaviour of the DUSP2 deletion (Fig. 8D,E).

The robustness of the core model with respect to either i) gain of p38 induced DUSP1 or ii) loss of JNK induced DUSP2 in isolation, can be explained as follows. To lock JNK into the highly active state (in the presence of ERK input), ERK activity needs to be suppressed, either by p38 activity via the PP2A-ERK link or by JNK activity via the DUSP2-ERK link. In the absence of DUSP1, p38 activity is sufficiently high to suppress ERK. In presence of DUSP1, the p38 activity is reduced and ERK is not sufficiently suppressed by p38-PP2A alone. Here, the JNK-DUSP2-ERK crosstalk becomes crucial as it complements the p38-PP2A mediated ERK inhibition, which explains the fragility of the JNK switch if both elevation of p38-induced DUSP1 and loss of JNK-induced DUSP2 occur simultaneously.

3 Discussion

The process of building a multi-pathway model is quite complex, as are the implications of its analysis for cell biology and cancer. Both are discussed in the following.

3.1 Theoretical considerations

The network depicted in Fig 1 synthesises information from different cell types in the literature. However, a complete picture of MAPK crosstalk is still lacking. The network of DUSP interactions is particularly difficult to dis-
Figure 6: Regulation of switch by AKT negative crosstalk. (A-C) Interaction schemes and simulated dose responses for crosstalk at different levels: MAP3K and MAP2K level (A); MAP3K level (B) and MAP2K level (C), whereby active AKT phosphorylates and inhibits ASK/MLK and/or MKK4/7 as indicated. Left: Interaction scheme. Right: Dose responses with respect to the JNK input $u_{\text{JNK}}$ for different AKT activation levels; $u_{\text{JNK}}(t) = \hat{u}_{\text{JNK}}$ for $t > 0$; $u_{\text{AKT}} = \hat{u}_{\text{AKT}}$ for all $t$; blue curves indicate low, red lines high AKT activity; dashed lines indicate a switch from low to high JNK activity.
Figure 8: Dynamics of the apoptotic switch for different crosstalk patterns. Centre and right columns show the trajectories of MAPKs activation after a stress stimulus ($u_{p38} = u_{JNK} = 1$ for $t > 0$) in the presence of a mitotic signal ($u_{ERK} = 1$ for all $t$) for the indicated interaction patterns. Top left: Scheme of MAPK interactions. Black arcs indicate core interactions, green arcs indicate additional DUSP mediated interactions analysed in panels (A-E). Top right: Table summarising the different DUSP interaction patterns corresponding to panels (A-E). (A) Core model (see Fig. 1 for a detailed scheme) (B) Core model and DUSP4 mediated negative feedback on ERK. (C) Core model and p38 induced DUSP1 expression mediating negative feedback to ERK and crosstalk to JNK: p38 $\dashrightarrow$ JNK,p38 ERK $\dashrightarrow$ JNK, ERK. (D) Model C and DUSP4 mediated negative feedback on ERK. (E) Model D and DUSP5/6 mediated negative feedback on ERK.
Figure 7: Regulation of the JNK switch by ERK and p38. (A) Interaction scheme. (B-C) Simulated dose responses with respect to the JNK inputs for different levels of constant ERK pathway stimulation: \( u_{\text{JNK}}(t) = \hat{u}_{\text{JNK}} \) for \( t > 0 \); \( u_{\text{p38}}(t) = 1 \) for \( t > 0 \); \( u_{\text{ERK}} = \hat{u}_{\text{ERK}} \) for all \( t \); blue curves indicate low, red lines high ERK stimulation; dashed lines indicate a switch from low to high JNK activity. (B) Dose responses for primary/nontransformed cells exhibiting p38 ⊥ ERK crosstalk. (C) Dose responses for transformed/tumourigenic cells lacking the p38 ⊥ ERK crosstalk (no PP2A upregulation in the model).

3.1.1 Modularity and neglected components

The nominal model neglects several context dependent MAPK crosstalks and feedbacks that are not necessary for implementing the JNK apoptotic switch. For example, ERK features several (negative) feedback loops (von Kriegsheim et al., 2009; Birtwistle et al., 2007). Generally, this feedback acts upstream of MEK (on the components that are not included in the model, such as growth factor receptors, their adaptors and GTPases), thereby transiently shaping Raf activation, i.e. the input of the model \( u_1 \). Thus, although the model does not account for ERK feedbacks explicitly, it can account for different Raf activation patterns by choosing \( u_1 \) accordingly as a time-dependent input function (Nakakuki et al., 2010). One advantage of this modular approach is that it facilitates further model development as the input functions can be replaced by additional sets of differential equations. Therewith, the model can be easily connected to other models describing the dynamics of different receptors and GTPases.

Future model development will concern including more mechanistic detail, in particular with regard to the well-studied ERK cascade. For instance, ERK exhibits a strong negative feedback to Raf-1 in response to EGF which alters the efficiency of MEK inhibition (Sturm et al., 2010) and might affect the p38-PP2A-MEK crosstalk. In con-
<table>
<thead>
<tr>
<th>DUSP</th>
<th>induced by</th>
<th>substrates</th>
<th>comments/references</th>
</tr>
</thead>
<tbody>
<tr>
<td>DUSP4/16</td>
<td>ERK (JNK)</td>
<td>JNK, ERK</td>
<td>Oncogenic Ras activity induces DUSP4 mRNA and protein synthesis and stabilises DUSP4 protein (Cagnol and Rivard, 2012); ERK phosphorylation stabilises DUSP16 protein, which dephosphorylate JNK&gt;&gt;p38&gt;ERK, mRNA expression was not analysed (Katagiri et al., 2005)</td>
</tr>
<tr>
<td>DUSP2</td>
<td>JNK (ERK)</td>
<td>ERK, p38</td>
<td>JNK/c-Jun activity and transformed v-JUN enhances ERK dephosphorylation (Shen et al., 2003; Black et al., 2002); DUSP2 was implicated in regulating the JNK → ERK/p38 crosstalk (Jeffrey et al., 2006); DUSP2 dephosphorylates ERK and p38 (Dickinson and Keyse, 2006)</td>
</tr>
<tr>
<td>DUSP1</td>
<td>p38 (ERK, JNK)</td>
<td>p38, JNK</td>
<td>p38 induces DUSP1, which dephosphorylates p38 and JNK (Hu et al., 2007; Small et al., 2007); DUSP1 mRNA is induced by p38 in response to heat shock (macrophages) (Wong et al., 2005), ansomycin (VSMC) (Bokemeyer et al., 1998), arsenite, UVC (C3H 10T1/2) (Li et al., 2001); and by ERK in response to serum (CCL-39) (Brondello et al., 1997), PDGF, phorbol ester, angiotensin II (VSMC) (Bokemeyer et al., 1998), heat shock, H2O2 (C3H 10T1/2) (Li et al., 2001); and by JNK after stress (NIH 3T3) (Bokemeyer et al., 1996)</td>
</tr>
<tr>
<td>DUSP5/6</td>
<td>ERK</td>
<td>ERK</td>
<td>MEK/ERK but not PI3K/AKT or p38/JNK regulates DUSP6 mRNA (stabilisation) and protein (destabilisation) levels (Bermudez et al., 2011) Ras and ERK activity induces dusp5 mRNA and regulates DUSP5 protein stability (Cagnol and Rivard, 2012; Kucharska et al., 2009)</td>
</tr>
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</table>

Table 3: Inducible DUSPs implemented in the model. Bold face indicate activities used in the nominal model. Normal face indicate activities used in Sec. 2.4.3. Round brackets indicate activities not used in this study but implemented in the model, wherewith the model is readily adaptable to cell type and cancer specific interaction patterns by setting the parameters in the model accordingly (see also Tab. 5).
3.1.2 Spatial aspects
Mathematically, the model describes the cell as well mixed compartment and does not distinguish subcellular compartments. This simplification might not be an issue for the ERK-JNK crosstalk as it features both a nuclear (DUSP4) and a cytosolic (DUSP16) component, but might overestimate the effects of the JNK-ERK/p38 and p38-JNK crosstalks, because their mediators DUSP2 and DUSP1 are exclusively localised in the nucleus. In general, the spatial regulation of DUSPs and MAPKs is complex, as DUSPs can both shuttle and sequester MAPKs in the nucleus and cytosol (Caunt and Keyse, 2012; Mandell et al., 2005; Karlsson et al., 2004; Masuda et al., 2001). In addition, many MAPKs and their MAP2Ks also shuttle between the cytosol and nucleus, and activation and deactivation can take place in both compartments (Plotnikov et al., 2011). For instance, MKK3/6 are located both in the cytosol and nucleus and can mediate p38 activation in the nucleus (Ben-Levy et al., 1998). Both ERK and MEK constitutively shuttle between the cytosol and nucleus, and ERK can be activated in both compartments (Fujisaka et al., 2006). Likewise, ERK deactivation by DUSPs can occur in the nucleus and cytosol depending on the localisation of particular DUSP isoforms. Nuclear DUSPs seem to serve as anchoring proteins that retain dephosphorylated ERK in the nucleus to prevent re-activation in the cytosol (Lenormand et al., 1998). Thus, spatial context seems to play an intricate role in modulation of MAPK activities, more work would be needed to decipher and model the spatial regulations of DUSPs and their effects on MAPK activities.

3.1.3 Parameter dependency
The parameterisation of dynamic models is complex, usually requiring time-course measurements in several different conditions and fitting of the model using global optimisation algorithms, whereby the resulting parameter estimates might vary depending on cell type and experimental context. To obtain a nominal model, we chose the parameters values in concordance with earlier models of MAPK signalling and kinetic information in the literature, such as half-life measurements of DUSPs (see Material and Methods). For simplification, the model assumed equal parameters for different MAPKs. We do not expect this assumption to withstand experimental validation during parameter estimation as it was adopted for theoretical reasons. (We refer to Cirit et al. (2010); von Kriegsheim et al. (2009); Nakakuki et al. (2010) for compilations of kinetic parameters and Legewie et al. (2008) for turnover rates.) First, assuming equal binding constants for kinases acting on shared substrates established symmetries in the model that yielded simplified, Michaelis-Menten-like kinetic expressions in a model reduction step (see Material and Methods). Second, choosing equal catalytic activities for the different MAPKs simplifies analysing the model based on the rationale that, in this case, the system dynamics are dominated by the systems structure and not biased towards possible imbalances of particular parameter values. Nonetheless, an important feature of the model is that the bistable nature of the JNK apoptotic switch does not depend on the exact parameter values used, but relies on feedback structure and sigmoidal shape of the i/o characteristic. Nevertheless, in order to achieve a quantitatively predictive model, future work would be needed for data collection and parameterisation, particularly with regard to stress activated kinases where little kinetic information is available.

3.2 Biological implications
Although the idea of a JNK positive feedback loop is not new (Ventura et al., 2006), a detailed understanding of how JNK feedback structures and crosstalk regulate cell fate is still missing (Wagner and Nebreda, 2009). Earlier modelling studies considered MAPK systems in isolation from each other, either focusing on ERK signalling and its feedbacks (Kholodenko et al., 2010; Sturm et al., 2010; Nakakuki et al., 2010; von Kriegsheim et al., 2009), or p38-JNK crosstalk (Sundaramurthy and Gakkhar, 2010; Sundaramurthy et al., 2009). In contrast, this manuscript provides a systems level model of JNK positive feedback, its regulation by pathway crosstalk including ERK and AKT signalling, and a mathematical analysis of how this system integrates different proliferative, survival and proapoptotic stimuli, thereby determining cell fates. The model helps us to understand the experimental observations in the literature, and incorporates several ideas. First, the magnitude and temporal profile of JNK signalling is important, as the anti-apoptotic, proliferative response is associated with moderate, but rapid JNK activation, whereas the proapoptotic response is associated with later, more sustained JNK activation (Ventura et al., 2006; Lamb et al., 2003; Sakon et al., 2003). Second, both mitotic signalling via ERK and survival signalling via AKT modulate the JNK apoptotic switch (Junttila et al., 2008; Molton et al., 2003). Third, the loss of negative crosstalk from p38 to ERK dysregulates JNK dependent apoptosis, which is crucial for cell transformation (Junttila et al., 2008; Arroyo and Hahn, 2005). Overall, JNK signalling involving a positive feedback loop takes a centre stage in the proposed model, which explains how ERK and AKT mediated crosstalk modulates and switches proliferative and proapoptotic JNK signalling.

3.2.1 Differences of ERK and AKT control over the JNK apoptotic switch
In the model, the switch to apoptotic JNK signalling depends crucially on a JNK positive feedback loop, which, once activated, causes high levels of sustained JNK activity. This switch is modulated by ERK and AKT signalling in different ways. ERK activity shifts the threshold for the JNK apoptotic switch to higher values, but has no
effect on the strength of apoptotic JNK signalling. The mechanism underlying this behaviour is the enhanced dephosphorylation of JNK, whereby JNK activity is either sufficient to activate the JNK positive feedback loop and inhibit ERK signalling, or does not reach this threshold level. In contrast, AKT activity predominantly regulates the strength of JNK signalling by reducing the value of the JNK-on-state with little effect on the switching threshold. The mechanism behind this is the phosphorylation and inhibition of JNK upstream kinases, which reduces the strength of both, the feedforward loop and the feedback loop. Crucially, the reduced feedback strength yields a reduced level of the JNK-on state.

3.2.2 Transformed versus normal cells

Transformed cell differ from normal cells in that they lack PP2A mediated negative crosstalk from p38 to ERK (Junttila et al., 2008). In the model, the loss of p38-ERK negative crosstalk severely increases the JNK switching threshold, thus desensitising the cells from stress induced apoptosis. Taken together, these observations suggest the involvement of the JNK apoptotic switch in cellular senescence as follows. With each cell cycle, cells accumulate DNA damage and experience a shortening of the chromosomal telomeres. Once a certain threshold of DNA damage or telomere shortening is crossed, senescence occurs or apoptosis is induced. The loss of p38-ERK crosstalk would increase this threshold to unphysiological levels, thus rendering transformed cells biologically immortal. In this senescence model, AKT is not involved, as the AKT-JNK crosstalk does not alter the apoptotic threshold, but, instead, prevents the apoptotic switch in the presence of survival signals.

4 Conclusions

The developed model explains how pathway crosstalk harmonises MAPK responses resulting in pivotal cell fate decisions that differ markedly between transformed and non-transformed cells. In the proposed model, JNK can switch from a transient to sustained activity due to multiple positive feedback loops. Once activated, positive feedback locks JNK into a highly active state that promotes cell death. The switch is differentially regulated by the ERK, p38 and AKT pathways. ERK activation enhances the dual specify phosphatase (DUSP) mediated dephosphorylation of JNK and shifts the threshold of the apoptotic switch to higher inputs. In nontransformed cells, activation of p38 can restore the threshold by inhibiting ERK activity via the phosphatases PP1 or PP2A. Finally, AKT activation inhibits the JNK positive feedback, thus abrogating the apoptotic switch and allowing only proliferative signalling. The model is most valuable for understanding how cancerous deregulations disturb the signal-processing of internal and external cues and provides possible explanations for certain drug resistances. For instance, oncogene induced ERK hyperactivity prevents the normal apoptotic switch and provides possible explanations for the complex and tumour specific behaviour of MAPK systems (Bermudez et al., 2010; Wagner and Nebreda, 2009; Dickinson and Keyse, 2006).

In regards to interactions necessary for facilitating the switch between transient and sustained JNK activity, our model predicts a critical role for DUSP1 and DUSP2 expression patterns. In the model, both expression of DUSP1 and deletion of DUSP2 are necessary for preventing the JNK apoptotic switch (as the nominal model is robust to dysregulation of either DUSP in isolation). The result is particularly interesting in the context of a) cancer, as many cancers show increased expression of DUSP1 and reduced expression of DUSP2, and b) tumour related conditions such as hypoxia, where low oxygen levels upregulate DUSP1 and downregulate DUSP2 (Lin et al., 2011; Patterson et al., 2009). According to our model, these conditions would prevent the JNK apoptotic switch. Indeed, forced expression of DUSP2 abolished hypoxia induced chemoresistance in human cancer cell lines (Lin et al., 2011), and inhibition of DUSP1 sensitised several resistant cancer cell lines to JNK dependent apoptosis (Wang et al., 2008; Small et al., 2007; Sanchez-Prez et al., 2000; Laderoute et al., 1999).

The current model represents a core network of MAPK interactions critical for the switch from proliferative to apoptotic signalling. The model is canonical in the sense that it generalises and integrates information from different cell lines by focusing on interactions that are a) readily observed in several cell lines and b) important for controlling the JNK bistable switch. The canonical model forms a basis for experimental design and can be tailored to different experimental systems on two levels by a) parameter estimation (the epigenetic and disease background of particular cell types is reflected in different parameter values of the model) and b) extending the model to incorporate different MAPK isoforms, upstream and downstream signalling and scaffolds. Such refined and validated models possess quantitative predictive power and cannot only be used for identifying gaps in knowledge by testing the model predictions, but also for predicting the effect of drugs, thus building the theoretical basis for identifying optimal treatment strategies.

5 Material and Methods

This section describes how the different components depicted in Fig. 1 are modelled mathematically.

5.1 Model of kinase activation

Activation of mitogen activated protein kinases requires the phosphorylation of two conserved amino acid residues, whereby several upstream kinases can act as enzymes facilitating the phosphorylation. Consider the reaction scheme in Fig. 9, where we assumed that only one enzyme can be bound to the kinase at any one time. Using the law of
mass action the reaction rates are described by

\begin{align}
\frac{dx_0}{dt} &= -r_1 - r_3, \\
\frac{dx_1}{dt} &= v_1 - r_2 + v_3 - r_4, \\
\frac{dx_2}{dt} &= v_2 + v_4, \\
\frac{dx_{0u_1}}{dt} &= r_1 - v_1, \\
\frac{dx_{1u_1}}{dt} &= r_2 - v_2, \\
\frac{dx_{0u_2}}{dt} &= r_3 - v_3, \\
\frac{dx_{1u_2}}{dt} &= r_4 - v_4.
\end{align}

Assuming equal dissociation constants is a strong assumption, in particular for different enzymes, but reduces the risk of over-parameterisation and facilitates the theoretical analysis of the model.

De-phosphorylation of kinases is catalysed by phosphatases and a mathematical model can be derived analogously, resulting in kinetic expressions resembling (5). Therewith, a complete model of a double phosphorylation cycle is given by

\[
\frac{dx_0}{dt} = - \sum_i (k_i \hat{u}_i) \frac{x_0}{K_d + x_0 + x_1} + \sum_j (k_{-j} \hat{u}_j) \frac{x_1}{K_d + x_1 + x_2},
\]

\[
\frac{dx_2}{dt} = \sum_i (k_i \hat{u}_i) \frac{x_1}{K_d + x_0 + x_1} - \sum_j (k_{-j} \hat{u}_j) \frac{x_2}{K_d + x_1 + x_2},
\]

where \( u_i \) denote the concentrations of the upstream kinases, \( v_j \) the concentrations of the phosphatases.

### 5.2 Model of kinase inhibition by phosphorylation

Some kinases can be rendered catalytically inactive by phosphorylation at inhibitory sites. Examples are ASK1 and MKK4, which can be phosphorylated by AKT at Ser 83 and Ser 78, respectively (Kim et al., 2001; Park et al., 2002). There is little, mostly conflicting information available on whether phosphorylation of the inhibitory site depends on the phosphorylation status of the activating sites, or, in turn, whether phosphorylation at the inhibitory site affects the phosphorylation/de-phosphorylation of the activating sites. Hence, we take a domain oriented approach accounting for all possible combinations of the phosphorylation status (but neglecting the possibility of a trimeric complex), resulting in a model comprising six distinct states (Conzelmann et al., 2008; Kiyatkin et al., 2006; Borisov et al., 2006, 2005).

Assume that the phosphorylation status of the activating site does not affect the binding of the inhibitor enzyme and the phosphorylation of the inhibitory site, and vice versa that the phosphorylation status of the inhibitory site does not affect the binding of of the activating enzyme and the phosphorylation of the activating site. Assume further, that only one enzyme can be bound to the kinase at any one time, i.e. a trimeric complex consisting of activating enzyme, kinase and inhibitory enzyme is not possible. Then the reaction scheme in Fig. 9 can be extended to account for kinase inhibition:

\[
X_i \overset{U_i}{\longrightarrow} X_{il}, \quad (i = 0, 1, 2),
\]

where \( X_{il} \) denotes the kinase phosphorylated at the inhibitory site and \( U_i \) the enzyme catalysing this phosphorylation.
Figure 9: Schemes of a double phosphorylation cycle with two kinases. X denotes the protein to be phosphorylated with the index indicating its phosphorylation status. U denote the kinases catalysing the phosphorylations. (A) Full mechanistic scheme modelled using mass action kinetics. For clarity of presentation, the dephosphorylation reactions are not depicted. (B) Reduced scheme modelled using Michaelis-Menten type kinetics as derived in the main text; $k_1$ and $k_2$ denote the catalytic activity of $U_1$ and $U_2$, respectively. $P$ denotes the phosphatase catalysing the dephosphorylation.

Similarly to (5) a mathematical description can be derived, yielding reaction kinetics for (de-)phosphorylation at the activating sites of the form

$$r_{\text{phos}} = \frac{\sum_x k_x \tilde{y} x}{K_d + x_0 + x_1 + x_{0f} + x_{1f}},$$

$$r_{\text{dephos}} = \frac{\sum_x k_x \tilde{z} x}{K_d + x_0 + x_1 + x_{0f} + x_{1f}},$$

where $(y, z) \in \{(x_0, x_1), (x_1, x_2), (x_{0f}, x_{1f}), (x_{0f}, x_{1f})\}$ and reaction kinetics for (de-)phosphorylation of the inhibitory site of the form

$$r_{\text{phos},1} = \frac{k_{1x} \tilde{y} x}{K_1 + x_0 + x_1 + x_2},$$

$$r_{\text{dephos},1} = \frac{k_{1x} \tilde{z} x}{K_1 + x_0 + x_1 + x_{1f} + x_{2f}},$$

where $(y, z) \in \{(x_0, x_1), (x_1, x_{1f}), (x_{2f}, x_{2f})\}$.

5.3 Model of phosphatase expression

The expression levels of inducible phosphatases depend on several parameters, including the activities of upstream MAPKs (Fig 1). We model the rate of mRNA synthesis using sigmoidal Hill functions of activator kinase concentrations (motivated by thermostatical arguments (Frank et al., 2012)) and assume first order kinetics for translation and degradation rates. Therewith a dynamic model of gene expression is described as

$$\frac{dx}{dt} = k_{\text{syn}} \frac{u^n}{K_n + u^n} - k_{\text{deg}} x,$$

$$\frac{dy}{dt} = p_{\text{syn}} x - p_{\text{deg}} y,$$

where $u$ denotes the concentration of upstream kinase activity, $x$ and $y$ denote mRNA and protein concentrations, respectively, and $k_{\text{syn}}, K_n, k_{\text{deg}}, p_{\text{syn}}$ and $p_{\text{deg}}$ are kinetic parameters. Many of these parameters can be fixed based on biologically reasonable assumptions and kinetic data available in the literature.

The half life of several unmodified DUSP proteins were reported as being between 20 and 45 min (DUSP1,4,5,6,16). However, following posttranslational regulation by phosphorylation and ubiquitination, these half lives ranged from as short as 7.5 min to as long as 4 h (Cagnol and Rivard, 2012; Kucharska et al., 2009; Katagiri et al., 2005). The half life of DUSP6 mRNA was reported as being between 20 to 40 min, which decreased to as little as 8 min following the inhibition of basal MEK activity (Bermudez et al., 2011). Neglecting this complexity, the model assumes equal half lives for protein and mRNA at a value of 30 min for all phosphatases, which fixes the degradation parameters according to $k_{\text{deg}} = p_{\text{deg}} = \log(2)/(30\text{min})$. Further, the model assumes a 10-fold amplification from the mRNA level to the protein level, fixing the protein synthesis parameter according to $p_{\text{syn}} = 10p_{\text{deg}}$. The expression level of mRNA was normalised ($x \leq 1$), which fixes the mRNA synthesis parameter at $p_{\text{syn}} = k_{\text{deg}}$. The remaining parameters were chosen such that for strong, constant activity of the upstream kinase yields reasonable expression levels of around 80% ($x \approx 0.8$) of the maximal possible value. Further assuming a reasonable degree of ultrasensitivity $n = 2$, fixes the threshold of half activation at $K = 0.5$.

Acknowledgments

The research leading to these results has received funding from the Science Foundation Ireland under grant No. 06/CE/B1129, and the European Union Seventh Framework Programme (FP7/2007-2013) ASSET project under grant agreement number FP7-HEALTH-2010-259348.

References


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Table 4: Reactions, rate expressions and parameters of the (de-)phosphorylation processes in the core model. For simplicity ASK in the table denotes ASK/MLK in the model.
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Table 5: Reactions, rate expressions and parameters of the phosphatase expression processes in the core model. Units: $^a$ unitless, $^b$ min$^{-1}$. The parameters $a$, $b$ and $\gamma$ are structural parameters $\in \{0,1\}$ that determine which MAPK induces which phosphatase. The values given in the table relate to the core model.
4 (DUSP4/MKP2) resulting in nuclear ERK1/2 inhibition. Oncogene.


pathway fails to elicit bax activation or apoptosis unless the phosphoinositide 3'-kinase (PI3K) pathway is inhibited. *Oncogene* 22(30), 4690–4701.


