

**First implication of *STRA6* mutations in isolated
anophthalmia, microphthalmia and coloboma:
a new dimension to the *STRA6* phenotype**

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Abstract

Microphthalmia, anophthalmia and coloboma (MAC) are structural congenital eye malformations that cause a significant proportion of childhood visual impairments. Several disease genes have been identified but do not account for all MAC cases, suggesting that additional risk loci exist. We used SNP homozygosity mapping (HM) and targeted next-generation sequencing to identify the causative mutation for autosomal recessive isolated colobomatous micro-anophthalmia (MCOPCB) in a consanguineous Irish Traveller family. We identified a double nucleotide polymorphism (g.1157G>A and g.1156G>A; p.G304K) in *STRA6* (MIM*610745) that was homozygous in all of the MCOPCB patients. The *STRA6* p.G304K mutation was subsequently detected in additional MCOPCB patients, including one individual with Matthew-Wood syndrome (MWS; MCOPS9). *STRA6* encodes a transmembrane receptor involved in vitamin A uptake, a process essential to eye development and growth. We have shown that the G304K mutant *STRA6* protein is mislocalised and has severely reduced vitamin A uptake activity. Furthermore, we reproduced the MCOPCB phenotype in a zebrafish disease model by inhibiting retinoic acid synthesis, suggesting that diminished retinoic acid levels account for the eye malformations in *STRA6* p.G304K patients. The current study demonstrates that *STRA6* mutations can cause isolated eye malformations in addition to the congenital anomalies observed in MWS.

Key Words: autosomal recessive colobomatous micro-anophthalmia; *STRA6*; homozygosity mapping; targeted next-generation sequencing; mutation analysis; Matthew-Wood syndrome

Introduction

Microphthalmia, anophthalmia and coloboma (MAC) are related structural, congenital eye malformations which display a spectrum of severity and can occur in isolation or as part of a syndrome. Microphthalmia refers to a small eye, defined by axial length, while anophthalmia denotes the complete absence of an eye (Morrison, et al., 2002). Both conditions can be present in uni- or bi-lateral form. The third type of structural malformation, coloboma, is a segmental ocular defect resembling a “keyhole” deficiency in the iris. Although the reported prevalence at birth varies greatly, microphthalmia and anophthalmia are estimated to occur in 14 and 3 per 100,000 births respectively (Morrison, et al., 2002), and estimates of the combined prevalence reach 30 per 100,000 births (Verma and Fitzpatrick, 2007). The estimated prevalence of coloboma is 1 in 10,000 (Stoll, et al., 1997). Although individually rare, MAC is reported to cause 14.3% of cases of childhood severe visual loss (Hornby, et al., 2000).

The aetiology of MAC is not well understood. Environmental factors such as maternal vitamin A deficiency and exposure to infections, viruses or toxins during pregnancy have been linked to eye malformations, however, the extent of their contribution to MAC remains to be clarified (Busby, et al., 2005; Hornby, et al., 2002; Jana and Sharma, 2010). Familial clustering of the conditions has implicated a significant genetic component whereby microphthalmia, anophthalmia or coloboma are occasionally found to be present in different eyes of the same individual or within different individuals of the same family (Francois and Haustrate-Gosset, 1976). Inheritance of MAC may be autosomal dominant (Morle, et al., 2000), autosomal recessive (Bessant, et al., 1998) or X-linked (Graham, et al., 1991) and linkage analysis in a small number of families where these disorders segregate in a Mendelian pattern has led to the mapping of loci at 2q37.1, 14q32, 15q12-q15 and Xq27 (Bessant, et al.,

1998; Graham, et al., 1991; Hmani-Aifa, et al., 2009; Morle, et al., 2000). Heritable causes of MAC, including chromosomal anomalies and single gene mutations, have also been identified although a high degree of genetic heterogeneity is evident. Several genes that are known to play a role in ocular development have been implicated in MAC including *SOX2* (MIM*184429), *CHX10* (MIM*142993), *PAX6* (MIM*607108), *OTX2* (MIM*600037), *PAX2* (MIM*167409), *RX* (MIM*601881), *SHH* (MIM*600725) and *SIX6* (MIM*606326) (Amiel, et al., 2000; Fantes, et al., 2003; Ferda Percin, et al., 2000; Gallardo, et al., 2004; Glaser, et al., 1994; Ragge, et al., 2005a; Schimmenti, et al., 2003; Voronina, et al., 2004). However causative mutations in these genes do not account for all MAC cases (Ragge, et al., 2005b), suggesting that additional MAC loci are yet to be identified.

The current study involves a consanguineous Irish Traveller family with nine members who are affected by one or more of microphthalmia, anophthalmia and ocular coloboma of varying severity (pedigree 1; Figure 1A). Three patients have bilateral micro/anophthalmos, two patients have unilateral anophthalmos and contralateral microphthalmos with iris coloboma, two patients have unilateral anophthalmos and contralateral iris coloboma, one patient has unilateral colobomatous microphthalmos and one patient has unilateral microphthalmus (Table 1). Clinical assessment of the affected individuals confirmed that the ocular malformations are non-syndromic. A diagnosis was made of an autosomal recessive non-syndromic colobomatous micro-anophthalmia, referred to as MCOPCB. This is the first report of MCOPCB in the Irish Traveller population, an endogamous group of nomads within the Irish population. Linkage analysis at 14q32, 18q21 and 20p11 had previously excluded *OTX2*, *CHX10*, *RX* and *VSX1* (MIM+605020) as possible candidate genes in this MCOPCB pedigree. We undertook a genome-wide SNP homozygosity mapping analysis to identify the locus responsible for MCOPCB in this family.

Materials and Methods

Patients

The initial family (pedigree 1) was referred to the clinical genetic service for genetic assessment by at least one of the authors. They are members of the Irish Traveller population with significant consanguinity (average $F = 0.063$, range 0.032-0.118). Nine family members in two generations are affected by varying degrees of microphthalmia, anophthalmia and coloboma. All patients had normal cranial and abdominal ultrasounds and no extra-ocular defects were observed. The patients had normal intellectual development. A diagnosis was made of an autosomal recessive non-syndromic colobomatous micro-anophthalmia (MCOPCB). Subsequently, another child was seen in the genetics clinic from an apparently unconnected Irish Traveller family (pedigree 2). The child presented with severe bilateral microphthalmia. The optic nerves could not be identified on a CT brain scan but the bony orbits were well developed (Supp. Figure S1A). No significant intracranial pathology was found. The only other congenital anomaly was a dysplastic right kidney. A child from a third unconnected Irish Traveller family later presented at clinic (pedigree 3). The patient had bilateral anophthalmia, absent pulmonary valves, polysplenia, absent uterus and hydronephrosis with ureterocoeles. Brain MRI at age 4 months showed that structure and myelination were normal for the child's age (Supp. Figure S1B-D). The ventricular system was also normal. Her clinical features were consistent with a diagnosis of Matthew-Wood syndrome (MWS; MIM#601186). The G banded karyotype of the eight patients was normal at 550 band resolution. Ethical approval for the study was obtained from the ethics committee of the National Centre for Medical Genetics (Our Lady's Children's Hospital Crumlin, Ireland).

SNP genotyping and identification of runs of homozygosity

DNA samples from six patients (IV:1, IV:2, IV:4, III:7, IV:9 and IV:11) and seven unaffected relatives (III:2, IV:3, IV:5, III:8, IV:10, III:9 and III:10) in pedigree 1 were available for molecular analysis. Genomic DNA was extracted from peripheral lymphocytes and genotyped for 1 million single nucleotide polymorphisms (SNPs) on an Illumina BeadStation 500GX platform. Runs of homozygosity (ROH) containing a minimum of 50 SNPs were identified using Illumina BeadStudio software. ROH that were common to all six affected individuals but were not homozygous in any of the seven unaffected relatives were identified. Log R ratios and B allele frequencies in ROH of interest were examined to exclude hemizygous deletions (Supp. Figure S2).

Targeted sequence capture

A customised NimbleGen sequence capture array was designed to specifically isolate the four candidate homozygous regions at 15q23-24.1. Whole genome amplified DNA from five individuals, 3 affected (IV:1, IV:2 and IV:9) and 2 unaffected (IV:5 and IV:10), was selected for sequence capture which was performed at Roche NimbleGen.

Single read sequencing and data analysis

Libraries were prepared from the captured DNA according to the Illumina protocol (Illumina Part # 1003806 Rev. B) and sequenced on an Illumina GAII sequencer (Trinity Genome Sequencing Laboratory). The single sequence reads were mapped to the reference human genome (hg18) using BWA version 0.5.7 (Li and Durbin, 2009). Reads of inadequate sequence quality and potential PCR duplicates (those with identical sequences, start and end sites) were discarded. The quality scores for the aligned reads were recalibrated using GATK (McKenna, et al., 2010). Variants and indels were identified using SAMtools (Li, et al.,

2009). Loci with inadequate sequence coverage (<3x) were excluded. Homozygous variants that were shared by the 3 patients that were sequenced, but differed to the 2 unaffected relatives, were retained for further analysis. Variants with a population (European) frequency >1%, as reported in dbSNP129, were considered unlikely to be disease-causing and were excluded. Of the remaining novel homozygous variants, non-synonymous substitutions were prioritised. Mutations of interest were validated by Sanger sequencing.

Validation of STRA6 mutations

STRA6 variants were confirmed by PCR in all available family members from pedigrees 1-3. A 248 base-pair region surrounding the NM_001142617.1:g.1157G>A and NM_001142617.1:g.1156G>A mutations in exon 11 of *STRA6* was amplified (primers 5'-ctgctggcccttttctg-3' and 3'-cctggagagctgggttg-5' and an annealing temperature of 62°C.) The PCR products were analysed by Sanger sequencing. Amino acid residues are numbered from the first methionine residue (reference sequence NM_001142617.1(STRA6_v001)), according to journal guidelines (www.hgvs.org/mutnomen). The double nucleotide polymorphism is submitted to dbSNP under accession NM_001142617.1:c.910_911delinsAA. A control panel, comprising 100 chromosomes from healthy members of the Irish Traveller population, was also tested for the *STRA6* g.1157G>A and g.1156G>A mutations by direct sequencing. The remaining exons of *STRA6* (exons 2-10 and 12-19) were amplified and sequenced in the patient diagnosed with MWS (pedigree 3). Primer sequences and reaction conditions are available on request.

Assay for retinol uptake activity using ³H-retinol/RBP

A human *STRA6* p.G304K mutant was produced using PCR and confirmed by DNA sequencing. The production of ³H-retinol/RBP was performed as previously described

(Kawaguchi, et al., 2007). STRA6 was cotransfected with lecithin retinol acyltransferase (LRAT) for the retinol uptake assay in COS-1 cells. To determine cellular ^3H -retinol uptake from ^3H -retinol/RBP, cells were washed with Hank's Buffered Salt Solution (HBSS) before incubation with ^3H -retinol/RBP diluted in serum free medium for 1 hour at 37°C. The reactions were stopped by removing the medium, washing the cells with HBSS, and solubilising the cells in 1% Triton X-100 in PBS. Radioactivity was measured with a scintillation counter.

Live and permeabilised cell staining of wild type and mutant STRA6

For live cell staining, a Myc epitope was inserted into an extracellular loop of human STRA6 between residues 132 and 133. Epitope insertion at this position does not affect vitamin A uptake activity or RBP binding activity of STRA6 (Kawaguchi, et al., 2007). STRA6-Myc constructs were transfected into COS-1 cells growing on gelatin-coated coverslips. At 24 hours after transfection, anti-Myc monoclonal antibody was added to the media. After 1 hour incubation at 37°C, the cells were washed with HBSS 3 times and fixed in 4% paraformaldehyde in PBS for 10 minutes at room temperature. After 3 washes with PBS, the fixed cells were incubated with the blocking buffer (5% normal goat serum and 0.3% Triton X-100 in PBS) for 1 hour at room temperature. The cells were then incubated with Alexa Fluor 488-conjugated goat anti-mouse antibody (Molecular Probes) diluted in the blocking buffer for another hour at room temperature. After 3 washes with PBS, the coverslips containing cells were mounted onto slides using VectaShield mounting medium. The cell surface expression of STRA6-Myc proteins was examined by fluorescence microscopy. Permeabilised staining was performed similarly to live cell staining procedures except that cells were incubated with the anti-Myc antibody after they have been fixed in 4% paraformaldehyde in PBS for 10 min.

Pharmacological inhibition of retinoid synthesis in zebrafish

Zebrafish embryos were used to model different levels of retinoic acid using citral dimethyl acetal, a precursor to the retinoic acid synthesis inhibitor citral (Marsh-Armstrong, et al., 1994). Embryos in which the primitive optic primordium had formed were treated with 0.05, 0.5 and 5 μ M citral dimethyl acetal at the 6- and 11-somite stage. The drug was added to the embryo media for one hour and then exchanged with embryo media multiple times to wash out the drug. The ratio of eye area to head size was measured in the treated zebrafish at 3 days post fertilisation (dpf). The morphology and eye structure of the treated zebrafish were examined and images taken at 3 and 5 dpf.

Results

SNP Homozygosity mapping

We undertook a genome-wide SNP homozygosity mapping analysis to identify the genetic locus involved in MCOPCB in pedigree 1. The 6 patients affected by MCOPCB shared 73 ROH (17.9 Mb), ranging in size from 12.8 kb to 3.6 Mb. ROH that were exclusively shared by the MCOPCB patients, and were not homozygous in any of the 7 unaffected relatives, were identified and comprised of four consecutive homozygous regions at 15q23-24.1. The candidate loci totalled 0.9 Mb in size and contained 15 genes (Figure 2 and Table 2). All affected individuals share a common 3.6 Mb homozygous haplotype between markers rs4777352 and rs3743487 (chr15:69342218-72909909), a finding that supports a common ancestral origin for this disease locus in pedigree 1.

Mutation analysis

A custom NimbleGen sequence capture array was designed to target the genomic loci of interest at 15q23-24.1 and the captured DNA was sequenced on an Illumina GAI sequencer. We identified 10 homozygous variants that segregated with the MCOPCB phenotype (Supp. Table S1). . Four of the variants were reported in dbSNP129 with a population frequency $\geq 1\%$ and were considered unlikely to be disease-causing (Supp. Table S2). Of the remaining 6 novel mutations, only two resulted in amino acid substitutions (Table 3). Both missense mutations occur in the gene encoding the stimulated by retinoic acid 6 protein (*STRA6*; MIM*610745) and are within the same amino acid residue. The MCOPCB patients are homozygous for a double nucleotide polymorphism (NM_001142617.1:c.910_911delinsAA) that induces a non-synonymous change (p.G304K) from glycine (non-polar) to lysine (polar) at a highly conserved residue of *STRA6* (Supp. Figure S3). Additional family members were then analysed for G304K by Sanger sequencing. We show complete segregation of the G304K mutation with the disease phenotype in all investigated family members (Figure 3 and Supp. Figure S4). The putative disease mutation was not present in 100 chromosomes of healthy, ethnically matched control samples from the Irish Traveller population.

Identification of *STRA6* p.G304K in additional patients

Subsequently, another child from an apparently unconnected consanguineous Irish Traveller family presented at clinic with eye malformations (pedigree 2; Figure 1B). The child had severe bilateral anophthalmia and a dysplastic right kidney. The *STRA6* locus was analysed by Sanger sequencing and the proband was also found to be homozygous for the G304K mutation, adding further support for its role in MCOPCB (Figure 3B). A child from a third unconnected consanguineous Irish Traveller family (pedigree 3) later presented at clinic with eye defects and congenital abnormalities (Figure 1C). Her clinical features were consistent with a diagnosis of Matthew-Wood syndrome (MWS; MIM#601186). Of note, her elder

brother was reported to have bilateral anophthalmia with no other malformations but was unavailable for clinical assessment or blood sampling. Sequence analysis confirmed that the patient with MWS was homozygous for the STRA6 p.G304K mutation (Figure 3C). To investigate the possibility of additional *STRA6* mutations in the more severely affected MWS patient, the remainder of the *STRA6* gene was sequenced. However, no other mutation suspected to be deleterious (missense, nonsense, frameshift or splice site) was identified (Supp. Table S3). SNP genotype analysis showed that the affected individuals from pedigrees 2 and 3 have the same 3.6 Mb haplotype surrounding the STRA6 p.G304K mutation as the patients from pedigree 1, supporting a common ancestral origin for the disease mutation in all three families.

Activity and localisation of mutant STRA6

The functional consequences of the STRA6 p.G304K mutation identified in the MCOPCB patients were investigated *in vitro* by introducing the mutation into COS-1 cells and examining protein activity and localisation. Compared to the wild-type STRA6 protein, we found that the G304K mutation almost completely abolished vitamin A uptake activity (6.4% of wild type activity level) of STRA6 from holo-RBP (Figure 4B). Live cell staining showed that the G304K mutation led to loss of cell surface expression of STRA6 (Figure 4C). Permeabilised immunostaining demonstrated that the G304K mutant was still expressed (Figure 4C) suggesting that the missense mutation either causes misfolding of STRA6 or interferes with its targeting mechanism to prevent cell surface expression, which may be responsible for the loss of vitamin A uptake activity from holo-RBP.

Pharmacological inhibition of retinoic acid synthesis in zebrafish

In vitro analyses demonstrated that the G304K STRA6 mutant has a severely diminished ability to transport retinoids into cells. Therefore, it is likely that retinoic acid levels, synthesised intracellularly, will also be diminished in MCOPCB patients homozygous for the missense mutation. Differences in retinoic acid levels within individual patients could account for the varying severity of MAC malformations. . We chose a pharmacological approach to reduce retinoic acid levels during zebrafish eye development. Zebrafish embryos were used to model different levels of retinoic acid using citral dimethyl acetal, a precursor to the retinoic acid synthesis inhibitor citral (Marsh-Armstrong, et al., 1994). Inhibition of retinoic acid synthesis produced a dose-dependent microphthalmia at 3 dpf, with an average of 40% reduction in eye size with 5 μ M inhibitor (Figure 5A and 5E-G). At 5 dpf, the effect of the retinoic acid inhibitor on eye size was even more pronounced. The inhibitor produced developmental eye defects ranging from mild to severe microphthalmia (Figure 5B-D). Furthermore, retinal pigment epithelium coloboma was also apparent and again with a range of severity (Figure 5H-J). Other developmental defects were visible, including defects in heart morphogenesis, consistent with the role of retinoic acid in multiple developmental processes.

Discussion

We report a study to determine the causative mutation for autosomal recessive non-syndromic colobomatous micro-anophthalmia (MCOPCB) in a large consanguineous pedigree from an ethnic minority Irish population. Using homozygosity mapping and targeted next-generation sequencing, we identified a novel mutation (G304K) in *STRA6* which encodes a transmembrane receptor involved in vitamin A uptake. The STRA6 p.G304K mutation was also found in a patient with Matthew-Wood syndrome, a condition

with congenital malformations which includes eye defects. The findings from cellular work and a zebrafish disease model, confirm the importance of vitamin A in normal eye development. The data presented here also shows that the same genetic mutation may be responsible for a single disorder (isolated eye defect) in one individual, but may give rise to a syndromic disorder (MWS) in another.

STRA6 and eye development

STRA6 encodes a transmembrane receptor for the retinol-binding protein (RBP) and is responsible for mediating vitamin A uptake from circulation to target organs. The protein is expressed in a number of organs that require vitamin A for normal development, including the eye, brain, placenta and testis (Kawaguchi, et al., 2007). The activity of *STRA6* may be especially important for tissues or cell types requiring a large amount of vitamin A, which is supported by the variation in tissue expression of the vitamin A transport system (RBP-*STRA6*). Retinoic acid (RA), a derivative of vitamin A, has long been known to play a critical role in vision and eye development (Hyatt and Dowling, 1997). Once transported across the membrane by *STRA6*, retinol is converted to a number of derivatives, termed retinoids, which in turn regulate the expression of vision-related genes. Retinoids can both enhance and suppress gene expression and excessive or deficient retinoid levels can result in severe birth defects (Collins and Mao, 1999). Therefore, tight regulation of retinol transport and maintenance of the correct level of endogenous retinoids is crucial for maintaining normal embryonic development (Zile, 2001). Different tissues vary in their vulnerability to vitamin A deficiency and excess. It is notable that the eye is the organ most dependent on vitamin A (Wilson, et al., 1953) and hence, most sensitive to *STRA6* mutations. Therefore it is not surprising that, while there is considerable variability in the clinical presentation of individuals with MWS, eye anomalies are a consistent feature.

Several *STRA6* missense mutations resulting in phenotypic effects have been reported to date (Chassaing, et al., 2009). The site of the missense mutation identified in the MCOPCB patients (G304) is located in the initial segment of the sixth transmembrane domain of *STRA6* (Kawaguchi, et al., 2008b) (Figure 4A). The insertion of a positively charged polar lysine side chain into the hydrophobic environment of the cell membrane is predicted to have a negative effect on the stability of the protein (White and Wimley, 1999) and the efficiency of protein anchoring (Tsuzuki, et al., 2003). This effect is very likely to cause local unfolding or significant structural rearrangements. Furthermore, due to their small size and backbone flexibility, glycine residues are often found in parts of proteins where a specific geometry is required for the polypeptide backbone (Claessens, et al., 1989). Insertion of the lysine residue, which is more restricted in conformation, is likely to further disrupt the functionally active conformation of *STRA6*.

Analysis of the G304K mutant *STRA6* protein in COS-1 cells

We propose that the G304K mutation may cause misfolding of *STRA6* or interfere with its targeting mechanism, leading to loss of vitamin A uptake activity and reduced intracellular retinoid levels. The *STRA6* G304K mutation (as identified in the MCOPCB patients) was investigated *in vitro*. We found that the *STRA6* mutation causes a severe reduction in vitamin A uptake activity from holo-RBP (6.4% of wild-type protein activity). Additional expression analysis found that the G304K mutant protein was well expressed but localised to the endoplasmic reticulum instead of the cell surface. Taken together, the cellular work suggested that patients homozygous for *STRA6* p.G304K would have severely depleted intracellular retinoid levels. The effect of low intracellular retinoid levels on embryonic development was

investigated in a zebrafish model. Mild to severe microphthalmia was observed following inhibition of retinoic acid synthesis in zebrafish embryos which supports the view that diminished retinoic acid levels account for the eye malformation phenotypes in STRA6 p.G304K patients. The functional analyses provide insight into the pathogenesis of the disorder and suggest that the ocular malformations observed in the MCOPCB patients are a result of the inability of the mutant STRA6 to mediate sufficient vitamin A uptake from circulation into the cell, leading to reduced retinoid levels during embryonic development.

Intra-familial phenotypic variability

The affected family members in pedigree 1 are homozygous for the same missense mutation but there is considerable heterogeneity in the phenotype of their eye malformations. The intra-familial phenotype heterogeneity may be attributed to the variability in vitamin A intake of the affected individuals (environmental factor) and/or differences in the degree of loss of STRA6 function (genetic factor). RBP/STRA6 independent mechanisms of retinoid delivery, including random diffusion and transport of retinyl esters on plasma lipoproteins, can provide cellular retinoids in the absence of RBP or STRA6. However, these random mechanisms have many limitations that made it necessary to evolve the RBP/STRA6 system (Smith and Goodman, 1976). Firstly, random retinoid distribution is associated with mild to severe toxic side effects (Adams, 1993; Nau, 2001; Nau, et al., 1994). The RBP/STRA6 system is designed to achieve specific delivery of vitamin A to target cells. The specificity is achieved through the high affinity and low off-rate of RBPs binding to retinol and the specific binding of RBP to its high-affinity receptor STRA6 on specific target cells that can store vitamin A. Secondly, random retinoid distribution requires constant retinoid intake to supply target tissues and cannot provide a stable supply during times of insufficiency. RBP plays the essential role of mobilizing vitamin A stored in the liver and serving as a buffer to maintain

stable vitamin A concentration in the blood (Blomhoff, et al., 1990; Quadro, et al., 2005). The buffering function is important given the adverse effect of both low and high retinoid levels on the growth and function of diverse organs. Thirdly, random retinoid distribution may not satisfy tissues that need a large amount of retinoid for proper function such as the eye. The combined actions of these RBP/STRA6-independent mechanisms of vitamin A uptake mean that the intracellular retinol levels will differ between individuals and may contribute to the variation in phenotypic severity.

Matthew Wood syndrome or isolated eye defects

STRA6 has previously been implicated in MWS, a multisystem disorder that includes microphthalmia and anophthalmia (Chassaing, et al., 2009; Golzio, et al., 2007; Pasutto, et al., 2007; West, et al., 2009; White, et al., 2008). MWS patients typically present with various combinations of eye, heart, lung and diaphragmatic defects. Of note, all of the 20 MWS patients reported in the literature to date had eye anomalies in the form of anophthalmia or microphthalmia, with heart malformations being the second most consistent feature. Currently, seventeen different *STRA6* mutations (8 missense, 3 nonsense and 6 frameshift) have been found in patients with MWS (Chassaing, et al., 2009). The clinical spectrum associated with *STRA6* mutations is extremely variable and no correlations between the nature of a *STRA6* mutation and phenotypic severity have been found (Chassaing, et al., 2009). The current study has shown that patients homozygous for the *STRA6* p.G304K mutation can develop an isolated eye phenotype (pedigrees 1 and 2) or the more severe MWS (pedigree 3), raising the question of how the G304K mutation produces an isolated eye malformation of variable severity in some patients but a multi-system disorder in others? Several mechanisms are proposed.

Firstly, one of the simplest explanations for the observed phenotypic differences may relate to the nature and location of the pathogenic *STRA6* mutation and its impact on *STRA6* function. It is plausible that MWS patients have more severe *STRA6* mutations which completely abolish vitamin A uptake activity, while the *STRA6* mutation observed in the MCOPCB patients may result in only a partial reduction in vitamin A uptake activity. *STRA6* is highly enriched in the retinal pigment epithelium in the adult eye (Bouillet, et al., 1997) suggesting a high requirement of tightly regulated vitamin A uptake in this organ. Accordingly, even small deficiencies in vitamin A are sufficient to produce an ocular malformation. Therefore, although the *STRA6* mutation observed in the MCOPCB patients results in an eye defect, there may still be sufficient uptake of vitamin A for proper functioning of the non-ocular tissues, such as the heart and brain, which do not have as high a dependency on vitamin A. In contrast, the more severe MWS mutations may result in complete loss of *STRA6* activity in all tissues, leading to the multi-system malformations which extend beyond eye defects. However, the G304K mutation identified in the MCOPCB patients reduces *STRA6* activity to a level similar to that observed for MWS mutations (~6%) (Kawaguchi, et al., 2008a) suggesting that the variation in phenotype arises through an alternate mechanism.

Secondly, differences in the timing and tissue expression of the mutation may explain why some patients with *STRA6* mutations have a phenotype restricted to the eye, but others present with multi-organ syndromes. Thirdly, a second as yet unidentified disease gene may contribute to the broad spectrum of symptoms associated with MWS, similar to the tri-allelic inheritance model proposed in Bardet-Biedl syndrome (BBS) (Katsanis, et al., 2001). However, Isken and colleagues demonstrated that knock-down of *stra6* in zebrafish produced a MWS phenotype whereby *STRA6* deficiency resulted in craniofacial and cardiac defects

and microphthalmia (Isken, et al., 2008). Although these observations need to be confirmed in germline mutants, they indicate that loss of *STRA6* alone is sufficient to produce the MWS phenotype. Finally, the *STRA6* p.G304K mutation may exhibit variable expressivity, which is not uncommon for MAC disease genes (Zenteno, et al., 2006). Similar to BBS, the inter- and intra-familial clinical variability in MCOPCB could be explained, in part, by the presence of second-site mutations that serve as modifiers of expressivity (Beales, et al., 2003; Leitch, et al., 2008; Stoetzel, et al., 2006). Recent work by Yeyati and colleagues has demonstrated that functional reduction of a second gene, *HSP90*, in two zebrafish eye mutants significantly influenced the expressivity of the eye defect (Yeyati, et al., 2007). Therefore it is possible that unidentified modifier loci may contribute to the variable expression of *STRA6* mutations.

It is also possible that while genetic factors predispose to eye malformations, the interaction of environmental risk factors may contribute to the development of multiple congenital anomalies, such as those observed in MWS. It is well established that vitamin A intake alone (either insufficient or excessive) can lead to severe developmental defects without any genetic contribution (Wilson, et al., 1953). The evidence from teratogen- and dietary-induced animal models of congenital diaphragmatic hernia (a feature of MWS) is suggestive of environmental and/or nutritional factors also being involved and more research in this area is warranted (Clugston, et al., 2006; Holder, et al., 2007).

We identified a missense mutation (*STRA6* p.G304K) that segregates with recessive MCOPCB and MWS in families from the Irish Traveller population. We have shown that substituting a highly conserved glycine at residue 304 with a lysine essentially abolishes vitamin A uptake activity of *STRA6*, most likely due to mislocalisation of the mutant protein. Inhibition of retinoid synthesis in a zebrafish disease model reproduced the MCOPCB phenotype suggesting that individuals homozygous for the G304K allele are unable to attain

the intracellular levels of vitamin A and retinoids required to support normal eye development during embryogenesis. To our knowledge, this is the first report of *STRA6* mutations associated with isolated eye malformations. Currently, diagnostic genetic testing of the *STRA6* gene is standard for patients presenting with MWS but is not typically recommended for patients with non-syndromic eye anomalies. However, our study shows that *STRA6* mutations can be responsible for non-syndromic eye malformations and we suggest that patients with isolated eye defects such as anophthalmia/microphthalmia should be considered for *STRA6* screening.

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References

- Adams J. 1993. Structure-activity and dose-response relationships in the neural and behavioral teratogenesis of retinoids. *Neurotoxicol Teratol* 15(3):193-202.
- Amiel J, Audollent S, Joly D, Dureau P, Salomon R, Tellier AL, Auge J, Bouissou F, Antignac C, Gubler MC and others. 2000. PAX2 mutations in renal-coloboma

- syndrome: mutational hotspot and germline mosaicism. *Eur J Hum Genet* 8(11):820-6.
- Beales PL, Badano JL, Ross AJ, Ansley SJ, Hoskins BE, Kirsten B, Mein CA, Froguel P, Scambler PJ, Lewis RA and others. 2003. Genetic interaction of BBS1 mutations with alleles at other BBS loci can result in non-Mendelian Bardet-Biedl syndrome. *Am J Hum Genet* 72(5):1187-99.
- Bessant DA, Khaliq S, Hameed A, Anwar K, Mehdi SQ, Payne AM, Bhattacharya SS. 1998. A locus for autosomal recessive congenital microphthalmia maps to chromosome 14q32. *Am J Hum Genet* 62(5):1113-6.
- Blomhoff R, Green MH, Berg T, Norum KR. 1990. Transport and storage of vitamin A. *Science* 250(4979):399-404.
- Bouillet P, Sapin V, Chazaud C, Messaddeq N, Decimo D, Dolle P, Chambon P. 1997. Developmental expression pattern of *Stra6*, a retinoic acid-responsive gene encoding a new type of membrane protein. *Mech Dev* 63(2):173-86.
- Busby A, Dolk H, Armstrong B. 2005. Eye anomalies: seasonal variation and maternal viral infections. *Epidemiology* 16(3):317-22.
- Chassaing N, Golzio C, Odent S, Lequeux L, Vigouroux A, Martinovic-Bouriel J, Tiziano FD, Masini L, Piro F, Maragliano G and others. 2009. Phenotypic spectrum of *STRA6* mutations: from Matthew-Wood syndrome to non-lethal anophthalmia. *Hum Mutat* 30(5):E673-81.
- Claessens M, Van Cutsem E, Lasters I, Wodak S. 1989. Modelling the polypeptide backbone with 'spare parts' from known protein structures. *Protein Eng* 2(5):335-45.
- Clugston RD, Klattig J, Englert C, Clagett-Dame M, Martinovic J, Benachi A, Greer JJ. 2006. Teratogen-induced, dietary and genetic models of congenital diaphragmatic hernia share a common mechanism of pathogenesis. *Am J Pathol* 169(5):1541-9.
- Collins MD, Mao GE. 1999. Teratology of retinoids. *Annu Rev Pharmacol Toxicol* 39:399-430.
- Fantes J, Ragge NK, Lynch SA, McGill NI, Collin JR, Howard-Peebles PN, Hayward C, Vivian AJ, Williamson K, van Heyningen V and others. 2003. Mutations in *SOX2* cause anophthalmia. *Nat Genet* 33(4):461-3.
- Ferda Percin E, Ploder LA, Yu JJ, Arici K, Horsford DJ, Rutherford A, Bapat B, Cox DW, Duncan AM, Kalnins VI and others. 2000. Human microphthalmia associated with mutations in the retinal homeobox gene *CHX10*. *Nat Genet* 25(4):397-401.
- Francois J, Haustrate-Gosset MF. 1976. [Genetic counseling in cases of microphthalmos and anophthalmos]. *J Genet Hum* 24 Suppl:35-41.
- Gallardo ME, Rodriguez De Cordoba S, Schneider AS, Dwyer MA, Ayuso C, Bovolenta P. 2004. Analysis of the developmental *SIX6* homeobox gene in patients with anophthalmia/microphthalmia. *Am J Med Genet A* 129A(1):92-4.
- Glaser T, Jepeal L, Edwards JG, Young SR, Favor J, Maas RL. 1994. *PAX6* gene dosage effect in a family with congenital cataracts, aniridia, anophthalmia and central nervous system defects. *Nat Genet* 7(4):463-71.
- Golzio C, Martinovic-Bouriel J, Thomas S, Mougou-Zrelli S, Grattagliano-Bessieres B, Bonniere M, Delahaye S, Munnich A, Encha-Razavi F, Lyonnet S and others. 2007. Matthew-Wood syndrome is caused by truncating mutations in the retinol-binding protein receptor gene *STRA6*. *Am J Hum Genet* 80(6):1179-87.
- Graham CA, Redmond RM, Nevin NC. 1991. X-linked clinical anophthalmos. Localization of the gene to Xq27-Xq28. *Ophthalmic Paediatr Genet* 12(1):43-8.
- Hmani-Aifa M, Ben Salem S, Benzina Z, Bouassida W, Messaoud R, Turki K, Khairallah M, Rebai A, Fakhfekh F, Soderkvist P and others. 2009. A genome-wide linkage scan in

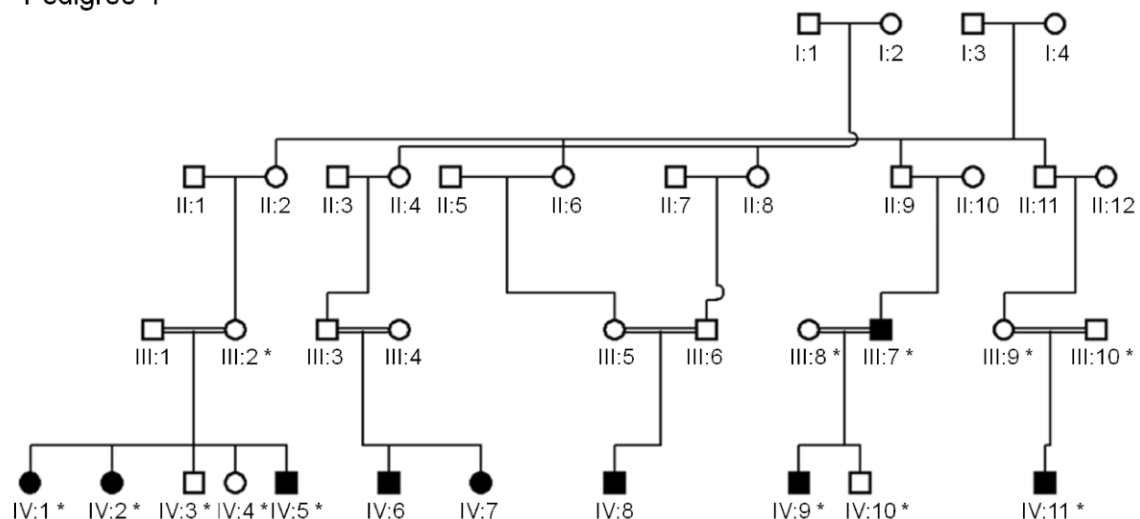
- Tunisian families identifies a novel locus for non-syndromic posterior microphthalmia to chromosome 2q37.1. *Hum Genet* 126(4):575-87.
- Holder AM, Klaassens M, Tibboel D, de Klein A, Lee B, Scott DA. 2007. Genetic factors in congenital diaphragmatic hernia. *Am J Hum Genet* 80(5):825-45.
- Hornby SJ, Gilbert CE, Rahi JK, Sil AK, Xiao Y, Dandona L, Foster A. 2000. Regional variation in blindness in children due to microphthalmos, anophthalmos and coloboma. *Ophthalmic Epidemiol* 7(2):127-38.
- Hornby SJ, Ward SJ, Gilbert CE, Dandona L, Foster A, Jones RB. 2002. Environmental risk factors in congenital malformations of the eye. *Ann Trop Paediatr* 22(1):67-77.
- Hyatt GA, Dowling JE. 1997. Retinoic acid. A key molecule for eye and photoreceptor development. *Invest Ophthalmol Vis Sci* 38(8):1471-5.
- Isken A, Golczak M, Oberhauser V, Hunzelmann S, Driever W, Imanishi Y, Palczewski K, von Lintig J. 2008. RBP4 disrupts vitamin A uptake homeostasis in a STRA6-deficient animal model for Matthew-Wood syndrome. *Cell Metab* 7(3):258-68.
- Jana M, Sharma S. 2010. Bilateral anophthalmia with septo-optic dysplasia. *Oman J Ophthalmol* 3(2):86-8.
- Katsanis N, Ansley SJ, Badano JL, Eichers ER, Lewis RA, Hoskins BE, Scambler PJ, Davidson WS, Beales PL, Lupski JR. 2001. Triallelic inheritance in Bardet-Biedl syndrome, a Mendelian recessive disorder. *Science* 293(5538):2256-9.
- Kawaguchi R, Yu J, Honda J, Hu J, Whitelegge J, Ping P, Wiita P, Bok D, Sun H. 2007. A membrane receptor for retinol binding protein mediates cellular uptake of vitamin A. *Science* 315(5813):820-5.
- Kawaguchi R, Yu J, Wiita P, Honda J, Sun H. 2008a. An essential ligand-binding domain in the membrane receptor for retinol-binding protein revealed by large-scale mutagenesis and a human polymorphism. *J Biol Chem* 283(22):15160-8.
- Kawaguchi R, Yu J, Wiita P, Ter-Stepanian M, Sun H. 2008b. Mapping the membrane topology and extracellular ligand binding domains of the retinol binding protein receptor. *Biochemistry* 47(19):5387-95.
- Leitch CC, Zaghloul NA, Davis EE, Stoetzel C, Diaz-Font A, Rix S, Alfadhel M, Lewis RA, Eyaïd W, Banin E and others. 2008. Hypomorphic mutations in syndromic encephalocele genes are associated with Bardet-Biedl syndrome. *Nat Genet* 40(4):443-8.
- Li H, Durbin R. 2009. Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics* 25(14):1754-60.
- Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R. 2009. The Sequence Alignment/Map format and SAMtools. *Bioinformatics* 25(16):2078-9.
- Marsh-Armstrong N, McCaffery P, Gilbert W, Dowling JE, Drager UC. 1994. Retinoic acid is necessary for development of the ventral retina in zebrafish. *Proc Natl Acad Sci U S A* 91(15):7286-90.
- McKenna A, Hanna M, Banks E, Sivachenko A, Cibulskis K, Kernytsky A, Garimella K, Altshuler D, Gabriel S, Daly M and others. 2010. The Genome Analysis Toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data. *Genome Res* 20(9):1297-303.
- Morle L, Bozon M, Zech JC, Alloisio N, Raas-Rothschild A, Philippe C, Lambert JC, Godet J, Plauchu H, Edery P. 2000. A locus for autosomal dominant colobomatous microphthalmia maps to chromosome 15q12-q15. *Am J Hum Genet* 67(6):1592-7.
- Morrison D, FitzPatrick D, Hanson I, Williamson K, van Heyningen V, Fleck B, Jones I, Chalmers J, Campbell H. 2002. National study of microphthalmia, anophthalmia, and

- coloboma (MAC) in Scotland: investigation of genetic aetiology. *J Med Genet* 39(1):16-22.
- Nau H. 2001. Teratogenicity of isotretinoin revisited: species variation and the role of all-trans-retinoic acid. *J Am Acad Dermatol* 45(5):S183-7.
- Nau H, Chahoud I, Dencker L, Lammer EJ, Scott WJ. 1994. Teratogenicity of Vitamin A and Retinoids. *Vitamin A in Health and Disease*: Marcel Dekker, Inc.
- Pasutto F, Sticht H, Hammersen G, Gillessen-Kaesbach G, Fitzpatrick DR, Nurnberg G, Brasch F, Schirmer-Zimmermann H, Tolmie JL, Chitayat D and others. 2007. Mutations in STRA6 cause a broad spectrum of malformations including anophthalmia, congenital heart defects, diaphragmatic hernia, alveolar capillary dysplasia, lung hypoplasia, and mental retardation. *Am J Hum Genet* 80(3):550-60.
- Quadro L, Hamberger L, Gottesman ME, Wang F, Colantuoni V, Blaner WS, Mendelsohn CL. 2005. Pathways of vitamin A delivery to the embryo: insights from a new tunable model of embryonic vitamin A deficiency. *Endocrinology* 146(10):4479-90.
- Ragge NK, Brown AG, Poloschek CM, Lorenz B, Henderson RA, Clarke MP, Russell-Eggitt I, Fielder A, Gerrelli D, Martinez-Barbera JP and others. 2005a. Heterozygous mutations of OTX2 cause severe ocular malformations. *Am J Hum Genet* 76(6):1008-22.
- Ragge NK, Lorenz B, Schneider A, Bushby K, de Sanctis L, de Sanctis U, Salt A, Collin JR, Vivian AJ, Free SL and others. 2005b. SOX2 anophthalmia syndrome. *Am J Med Genet A* 135(1):1-7; discussion 8.
- Schimmenti LA, de la Cruz J, Lewis RA, Karkera JD, Manligas GS, Roessler E, Muenke M. 2003. Novel mutation in sonic hedgehog in non-syndromic colobomatous microphthalmia. *Am J Med Genet A* 116A(3):215-21.
- Smith FR, Goodman DS. 1976. Vitamin A transport in human vitamin A toxicity. *N Engl J Med* 294(15):805-8.
- Stoetzel C, Laurier V, Faivre L, Megarbane A, Perrin-Schmitt F, Verloes A, Bonneau D, Mandel JL, Cossee M, Dollfus H. 2006. BBS8 is rarely mutated in a cohort of 128 Bardet-Biedl syndrome families. *J Hum Genet* 51(1):81-4.
- Stoll C, Alembik Y, Dott B, Roth MP. 1997. Congenital eye malformations in 212,479 consecutive births. *Ann Genet* 40(2):122-8.
- Tsuzuki D, Hichiya H, Okuda Y, Yamamoto S, Tamagake K, Shinoda S, Narimatsu S. 2003. Alteration in catalytic properties of human CYP2D6 caused by substitution of glycine-42 with arginine, lysine and glutamic acid. *Drug Metab Pharmacokinet* 18(1):79-85.
- Verma AS, Fitzpatrick DR. 2007. Anophthalmia and microphthalmia. *Orphanet J Rare Dis* 2:47.
- Voronina VA, Kozhemyakina EA, O'Kernick CM, Kahn ND, Wenger SL, Linberg JV, Schneider AS, Mathers PH. 2004. Mutations in the human RAX homeobox gene in a patient with anophthalmia and sclerocornea. *Hum Mol Genet* 13(3):315-22.
- West B, Bove KE, Slavotinek AM. 2009. Two novel STRA6 mutations in a patient with anophthalmia and diaphragmatic eventration. *Am J Med Genet A* 149A(3):539-42.
- White SH, Wimley WC. 1999. Membrane protein folding and stability: physical principles. *Annu Rev Biophys Biomol Struct* 28:319-65.
- White T, Lu T, Metlapally R, Katowitz J, Kherani F, Wang TY, Tran-Viet KN, Young TL. 2008. Identification of STRA6 and SKI sequence variants in patients with anophthalmia/microphthalmia. *Mol Vis* 14:2458-65.

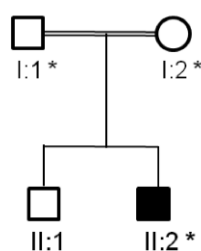
- Wilson JG, Roth CB, Warkany J. 1953. An analysis of the syndrome of malformations induced by maternal vitamin A deficiency. Effects of restoration of vitamin A at various times during gestation. *Am J Anat* 92(2):189-217.
- Yeyati PL, Bancewicz RM, Maule J, van Heyningen V. 2007. Hsp90 selectively modulates phenotype in vertebrate development. *PLoS Genet* 3(3):e43.
- Zenteno JC, Perez-Cano HJ, Aguinaga M. 2006. Anophthalmia-esophageal atresia syndrome caused by an SOX2 gene deletion in monozygotic twin brothers with markedly discordant phenotypes. *Am J Med Genet A* 140(18):1899-903.
- Zile MH. 2001. Function of vitamin A in vertebrate embryonic development. *J Nutr* 131(3):705-8.

Figure titles and legends

A Pedigree 1



B Pedigree 2



C Pedigree 3

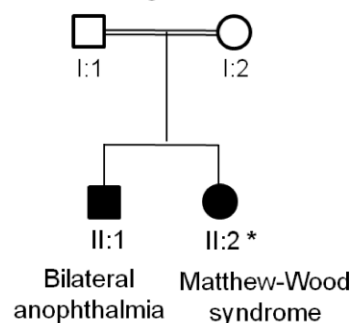


Figure 1. Pedigrees of Irish Traveller families with MCOPCB. (A-C) The inheritance pattern in each pedigree is compatible with an autosomal recessive disease gene. Pedigrees 1-3 are from the Irish Traveller population but are unrelated to each other. **A:** DNA samples were

available for 6 affected and 7 unaffected family members in pedigree 1. **B:** DNA was available for the affected proband and his parents in pedigree 2. **C:** The female proband in pedigree 3 was diagnosed with Matthew-Wood syndrome (MWS). Her elder brother had bilateral anophthalmia only and no other clinical features. DNA was available for the MWS patient.

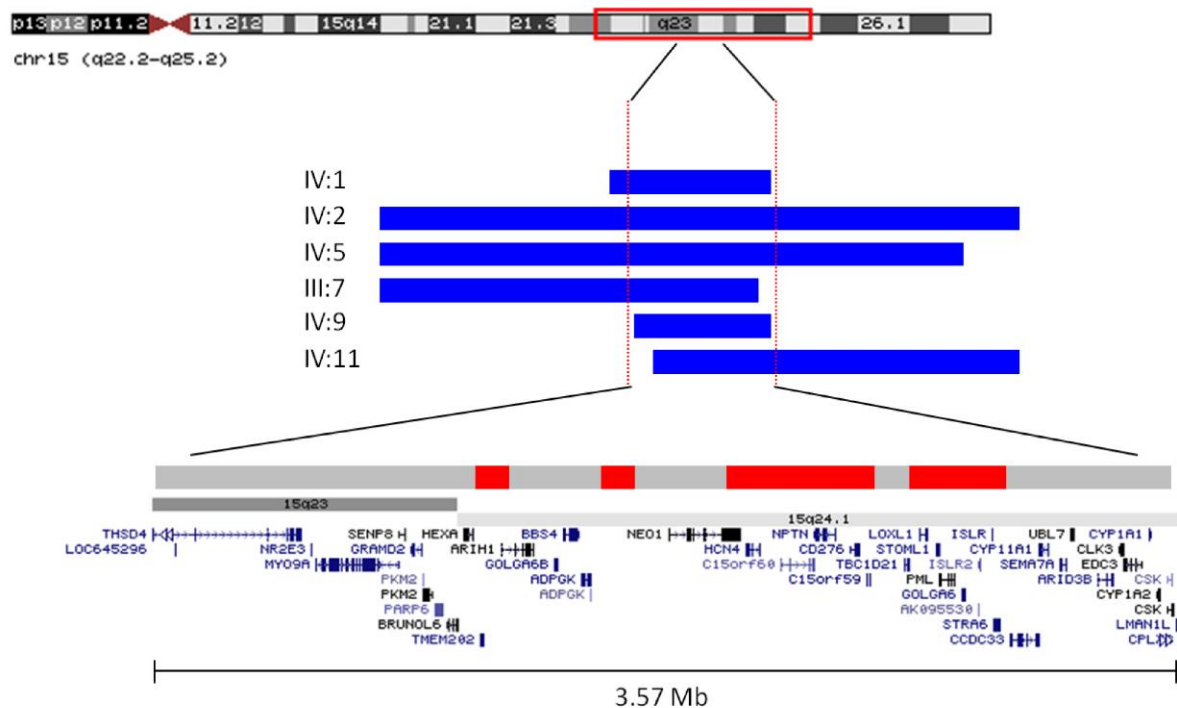
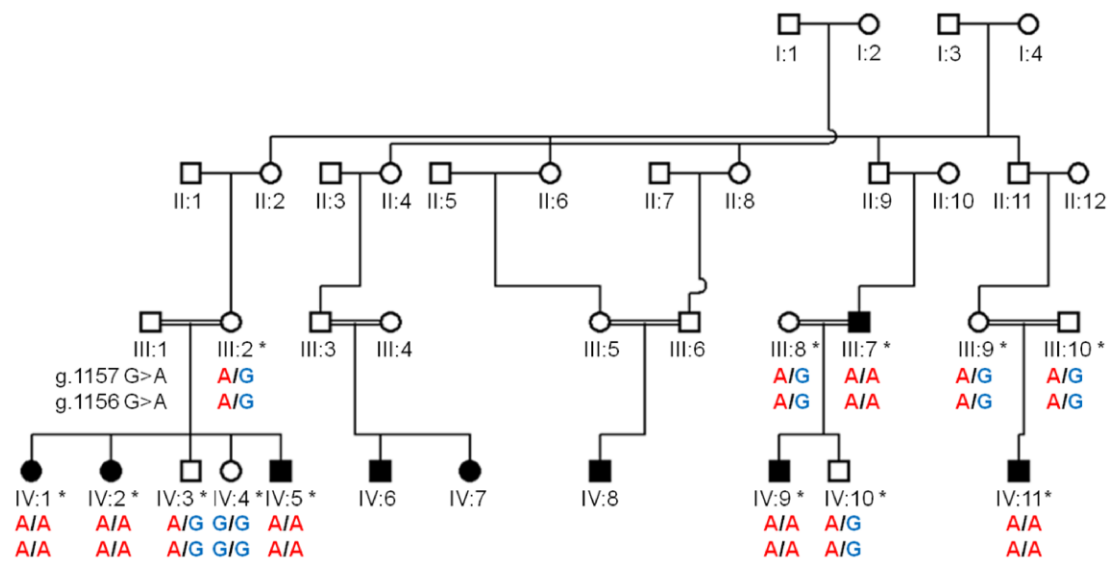
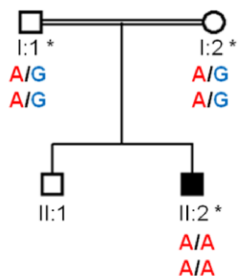


Figure 2. Putative MCOPCB disease locus at 15q23-24.1 in pedigree 1. The ROH at 15q23-24.1 in 6 members of the family affected by MCOPCB are shown as horizontal blue bars. The ROH vary in size from 4.1 Mb to 11.1 Mb. The maximal shared region is 3.57 Mb containing 43 RefSeq genes. Analysis of the homozygosity patterns in 7 unaffected family members (grey bars) reduced the ROH of interest to 0.9 Mb in size and 15 genes (red bars).

A Pedigree 1



B Pedigree 2



C Pedigree 3

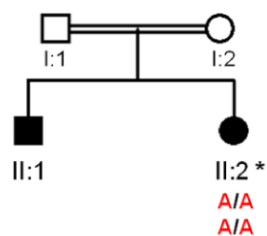


Figure 3. Segregation of *STRA6* p.G304K with MCOPCBAnalysis of Sanger sequence confirmed segregation of the G304K mutation with the eye malformation phenotype in

pedigrees 1-3 (A-C). The top and bottom genotypes denote the DNA mutations at chromosome 15 positions 72,270,250 ((NM_001142617.1:g.1157G>A) and 72,270,251 (NM_001142617.1:g.1156G>A) respectively (hg18).

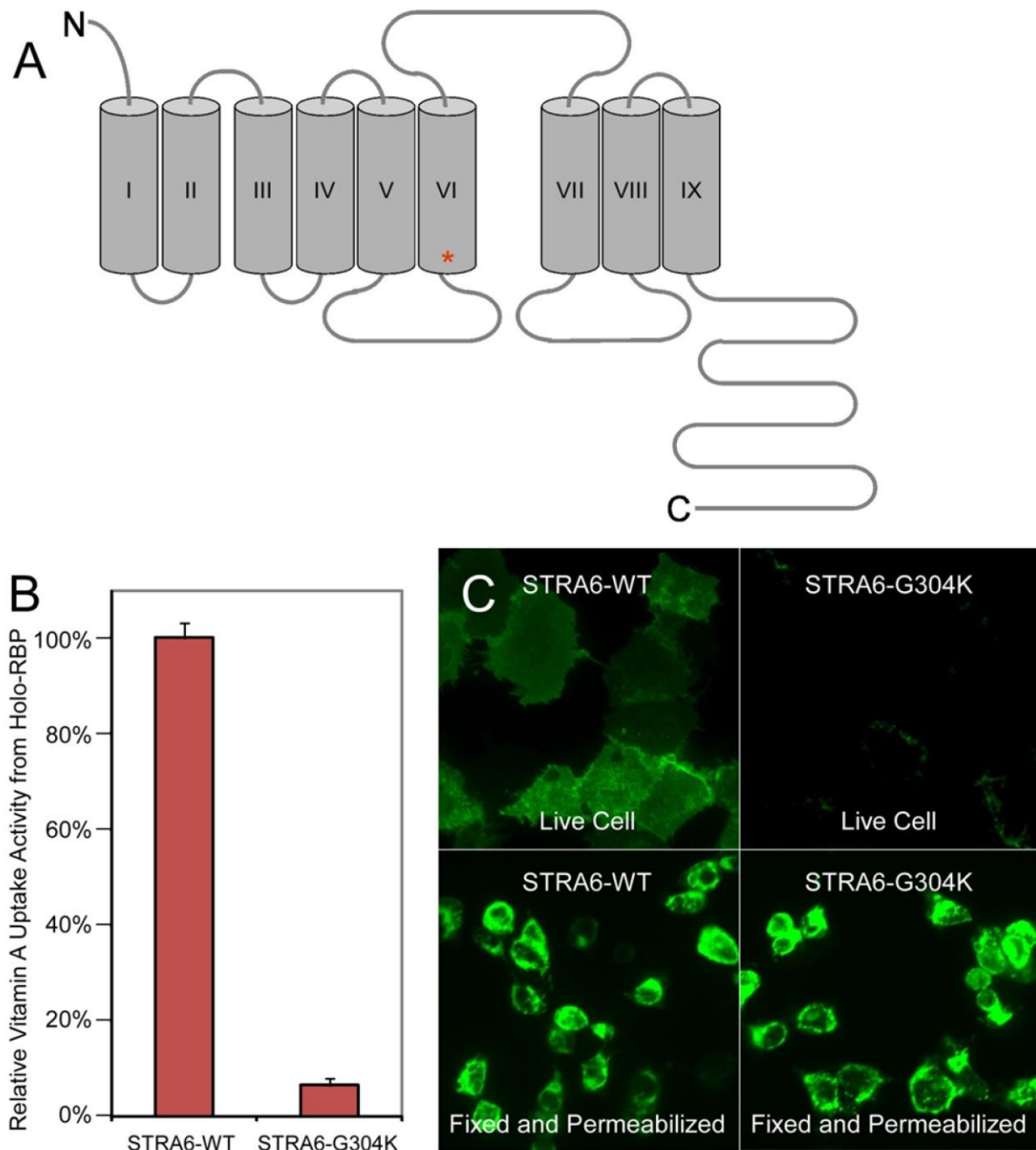


Figure 4. Examination of vitamin A uptake and localisation of the G304K STRA6 mutant protein in COS-1 cells. **A:** STRA6 transmembrane topology model based on Kawaguchi et al. 2008 (Mapping the membrane topology and extracellular ligand binding domains of the retinol binding protein receptor. *Biochemistry* 47:5387-5395). Residue G304 is located at the initial segment of transmembrane domain 6 (red asterisk). **B:** Retinol uptake assay from holo-RBP comparing wild-type human STRA6 and the G304K STRA6 mutant. Wild-type

STRA6's activity is defined as 100%. The mutant has greatly reduced retinol uptake activity (6.4%). C: The G304K STRA6 mutant is expressed similarly to the wild-type protein as shown in the permeabilised staining but is poorly expressed on the cell surface as shown in the live cell staining.

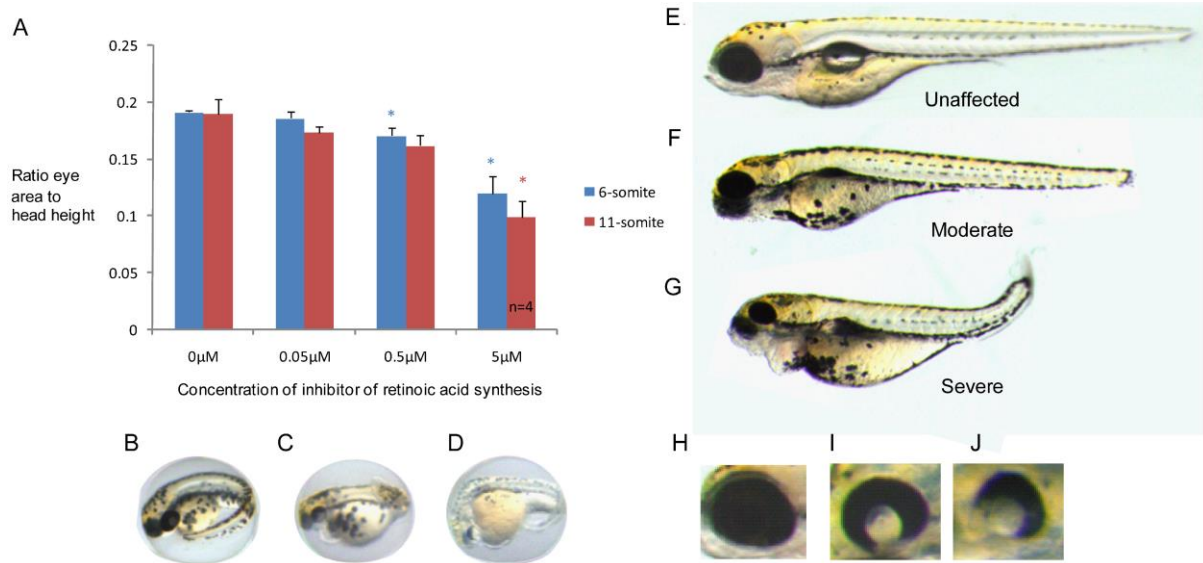


Figure 5. Pharmacological inhibition of retinoic acid synthesis in zebrafish embryos. Treatment of zebrafish embryos with a retinoic acid inhibitor, citral dimethyl acetal, has a dose-dependent effect on eye development, producing microphthalmic zebrafish. **A:** The x-axis represents the concentration of citral dimethyl acetal and the y-axis denotes the ratio of eye area to head height. Measurements were taken 3 days post fertilization. The embryos were treated during early eye development at the 6- (blue) or 11- (red) somite stage. P-values ≤ 0.05 are denoted by a *. Unless otherwise stated, $n=5$. **B, C, D:** Representative images of an untreated, moderately affected and severely affected zebrafish respectively at 3 days post fertilisation. **E, F, G:** Representative images of an unaffected, moderately affected and severely affected zebrafish respectively following treatment with citral dimethyl acetal. The images were taken at 5 days post fertilisation. **H, I, J:** Images of a wild type zebrafish eye and a moderate and severe retinal pigment epithelium coloboma respectively after inhibition of retinoic acid synthesis.

Tables

Table 1. Clinical features of patients with MCOPCB in pedigree 1

Ind	Sex	Age	Eye malformation
IV:1*	M	16	Right microphthalmia, left iris coloboma
IV:2*	F	5	Right choroidal coloboma, left anophthalmia (small cystic mass)
IV:5*	F	15	Bilateral microphthalmia
IV:6	M	4	Right microphthalmia, left microphthalmia with coloboma
IV:7	F	2	Right microphthalmia and choroidal coloboma, left anophthalmia
IV:8	M	20	Bilateral anophthalmia
III:7*	M	26	Right anophthalmia, left microphthalmia with sclerocornea
IV:9*	M	15 mo	Right microphthalmus, left anophthalmia and optic cyst
IV:11*	M	16 mo	Bilateral anophthalmus (poor fornices)

Nine members of an Irish Traveller family are affected with varying degrees of microphthalmia, anophthalmia and ocular coloboma. DNA from 6 affected individuals (denoted with a *) was available for molecular analysis.

Table 2. Candidate loci identified by SNP homozygosity mapping

Genomic position	ROH size (kb)	RefSeq genes
15:70,482,945-70,578,645	95.70	<i>TMEM202, ARIH1</i>
15:70,900,743-70,985,517	84.77	Intergenic
15 :71,350,548-71,867,728	517.18	<i>NEO1, HCN4, C15orf60, NPTN, CD276, C15orf59</i>
15:72,004,570-72,306,485	301.92	<i>LOXL1, STOML1, PML, GOLGA6, ISLR2, ISLR, STRA6</i>

SNP homozygosity mapping identified four consecutive ROH at 15q24.1 that were shared by the 6 MCOPCB patients but were not homozygous in the 7 unaffected relatives. The four regions total 0.9 Mb and contain 15 RefSeq genes. Genomic positions are based on NCBI build 36.1 (hg18).

Table 3. Novel homozygous mutations at the 15q24.1 locus that segregate with the MCOPCB phenotype

Position	Gene	Type	IV:1	IV:2	IV:9	IV:5	IV:10	Substitution
71,382,144	<i>NEO1</i>	3' utr	CC	CC	CC	CT	CT	-
71,401,251	<i>HCN4</i>	3' utr	GG	GG	GG	AA	AA	-
71,640,342	<i>NPTN</i>	Exon	AA	AA	AA	AG	GG	Synonymous
72,270,250	<i>STRA6</i>	Exon	TT	TT	TT	CT	CC	G304E*
72,270,251	<i>STRA6</i>	Exon	TT	TT	TT	CT	CC	G304R*
72,274,265	<i>STRA6</i>	3' utr	AA	AA	AA	AG	GG	-

Sequence analysis of the 15q24.1 candidate locus identified 6 novel homozygous mutations that segregated with the MCOPCB phenotype. Two of the exonic mutations are in *STRA6* and result in non-synonymous substitutions. *The mutations at positions 72,270,250 (NM_001142617.1:g.1157G>A) and 72,270,251 (NM_001142617.1:g.1156G>A) are consecutive DNA bases of the same amino acid residue (304) and combine to give a p.G304K substitution in *STRA6* (NM_001142617.1:c.910_911delinsAA). The genomic positions refer to build hg18.