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## Specialized Pro-resolving Lipid **Mediators: Modulation of Diabetes-Associated Cardio-, Reno-,** and Retino-Vascular Complications

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Diabetes and its associated chronic complications present a healthcare challenge on a global scale. Despite improvements in the management of chronic complications of the micro-/macro-vasculature, their growing prevalence and incidence highlights the scale of the problem. It is currently estimated that diabetes affects 425 million people globally and it is anticipated that this figure will rise by 2025 to 700 million people.

The vascular complications of diabetes including diabetes-associated atherosclerosis and kidney disease present a particular challenge. Diabetes is the leading cause of end stage renal disease, reflecting fibrosis leading to organ failure. Moreover, diabetes associated states of inflammation, neo-vascularization, apoptosis and hypercoagulability contribute to also exacerbate atherosclerosis, from the metabolic syndrome to advanced disease, plaque rupture and coronary thrombosis. Current therapeutic interventions focus on regulating blood glucose, glomerular and peripheral hypertension and can at best slow the progression of diabetes complications. 

Recently advanced knowledge of the pathogenesis underlying diabetes and associated complications revealed common mechanisms, including the inflammatory response, insulin resistance and hyperglycemia. The major role that inflammation plays in many chronic diseases has led to the development of new strategies aiming to promote the restoration of homeostasis through the "resolution of inflammation." These strategies aim to mimic the spontaneous activities of the 'specialized pro-resolving mediators' (SPMs), including endogenous molecules and their synthetic mimetics. This review aims to discuss the effect of SPMs [with particular attention to lipoxins (LXs) and resolvins (Rvs)] on inflammatory responses in a series of experimental models, as well as evidence from human studies, in the context of cardio- and reno-vascular diabetic complications, with a brief mention to diabetic retinopathy (DR). 

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INTRODUCTION 125

> Diabetes and its associated complications pose a challenge to human health on a global scale. It is estimated that 425 m people are currently living with diabetes and this is predicted to rise to 700 m by 2025 The Lancet, 2018. The rapid rise in diabetes and its associated complications over the past three decades reflects numerous factors including aging, obesity, urbanization and greater longevity amongst patients (Zimmet et al., 2001). Among NCCDs, diabetes is one of the major global causes of premature mortality. It is frequently underestimated because very often persons with diabetes die from causes related to co-morbidities (Lee et al., 2012).

> The prevalence of diabetes in adults worldwide is predicted to be higher in developed than in developing countries, while, the incidence of diabetes is predicted to be higher in developing

These data collectively support the hypothesis that endogenously generated SPMs or 172 173 synthetic mimetics of their activities may represent lead molecules in a new discipline. 174 namely the 'resolution pharmacology,' offering hope for new therapeutic strategies to 175 prevent and treat, specifically, diabetes-associated atherosclerosis, nephropathy and 176 retinopathy. 177

Keywords: diabetic kidney disease, diabetes-associated atherosclerosis, diabetic retinopathy, lipoxins, resolvins

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countries (Wild et al., 2004). Thus, by the year 2030, the countries 182 with the largest number of people with diabetes are predicted 183 to be India, China and the United States; the countries with 184 increased prevalence of overweight and obese inhabitants, the 185 main drivers of T2D (Yach et al., 2006). The burden on already 186 challenged health care systems is unprecedented. 187

Diabetes is essentially a disorder of glucose homeostasis. 188 Conventionally, diabetes has been classified as Type-1, Type-189 2 and gestational diabetes. In T1D, autoimmune destruction 190 of the  $\beta$ -cells of the pancreas creates an insulin-deficient state 191 where patients are dependent on exogenous insulin for survival. 192 The precise mechanisms underlying the pathogenesis remain 193 elusive, but it is likely that genetic and environmental factors 194 converge to drive an autoimmune response. The observed 195 increasing incidence of T1D in developed nations is thought to 196 reflect responses to environmental triggers. Historically, T1D is 197 considered to represent 10% of total number of persons with 198 diabetes (You and Henneberg, 2016). In T2D, peripheral insulin 199 resistance in target tissues (including skeletal muscle, adipose 200 tissue and liver coupled with hypersecretion of insulin) typically 201 precedes eventual β-cell loss. The diabetes epidemic is commonly 202 attributed to T2D (Tuomi, 2005). Gestational diabetes describes 203 insulin resistance observed during pregnancy, which generally 204 resolves in the postnatal period. However, these mothers are at 205 increased risk of T2D in later life (Buchanan and Xiang, 2005). 206

It is now clear that the above classifications are an over-207 simplification. It has recently been proposed to re-define 208 diabetes based on six clinical parameters [BMI, age at diagnosis, 209 hemoglobin A1c, glutamate decarboxylase autoantibodies 210 (GADAs) (evidence of autoimmunity);  $\beta$ -cell function and 211 insulin sensitivity]. This has led to the identification of five 212 distinct pathologies associated with different disease progression 213 and risks of complications. Further characterization of the genetic 214 architecture of these subgroups may facilitate identification of 215 patients most at risk of specific complications (Rossing, 2018). 216 According to the proposed classification, the sub-groups can be 217 defined as: (1) SAID: This is the least common subtype (6.4%) 218 and traditionally classified as T1D. These patients had an early 219 onset of disease and were positive for GADAs, had low BMI and 220 were dependent on exogenous insulin (Abegunde et al., 2007). (2) 221 SIDD: these defined as a group of patients who showed insulin 222 deficiency and were GADAs negative. This group was at greatest 223 risk of DR (Jingi et al., 2017). (3) SIRD: This group represented 224 15.3% of the whole cohort of participants. These patients had a 225 high degree of insulin resistance and were likely to be overweight 226 or obese and showed kidney damage more frequently than other 227 groups. They also had a higher risk of non-alcoholic fatty liver 228

<sup>142</sup> Abbreviations: 18R-/17S-HEPE, 18R-/17S-hydroeicosapentaenoic acid; ABC-A/G, ATP-binding cassette-subfamily A/G; ACs, apoptotic cells; ACE, 143 angiotensin-converting enzyme; AGE, advanced glycation end-product; 144 AKI, acute kidney injury; ALX/FPR2, lipoxin/N-formyl peptide receptor-145 2; Apo, apolipoprotein; ATL, aspirin-triggered LX; ATLa, aspirin-triggered 146 LX analog; BM, basement membrane; BMI, body mass index; BMP, bone morphogenetic protein; BRB, blood-retinal barrier; CAD, coronary artery 147 disease; CANTOS, Canakinumab Anti-inflammatory Thrombosis Outcome 148 Study; CARDS, Collaborative Atorvastatin Diabetes Study; CCR, CC-chemokine 149 receptor; CD, cluster of differentiation; ChemR23, chemerin-like-1; CTGF, connective tissue growth factor; CVD, cardiovascular disease; DAA, diabetes-150 associated atherosclerosis; DAMPs, damage-associated molecular patterns; DHA, 151 docosahexaenoic acid; DKD, diabetic kidney disease; DPP-4, di-peptidyl peptidase-152 4; ECs, endothelial cells; ECM, extracellular matrix; eGRF, estimated glomerular 153 filtration rate; EMT, epithelial-mesenchymal transition; EPA, eicosapentaenoic acid; ESRD, end stage renal disease; GADA, GAD autoantibody; GLP-1-RA, 154 glucagon-like peptide-1 receptor agonist; GPCRs, G-protein coupled receptors; 155 HDL, high density lipoprotein; HK-2, human proximal tubular epithelial; HO, 156 heme-oxygenase; IHD, ischemic heart disease; IKK- $\beta$ , inhibitor of nuclear factor kappa-ß; IRI, ischemia reperfusion injury; JAK/STAT, janus kinase/signal 157 transducers and activators of transcription; LDL, low density lipoprotein; LDL-R, 158 LDL-receptor; LO, lipoxygenase; LX, lipoxin; MaR, maresin; MARDs, mild age-159 related diabetes; M-CSF, macrophage colony-stimulating factor; MI, myocardial 160 infarction; MMP, matrix metalloproteinase; MOD, mild obesity-related diabetes; NCCD, non-communicable chronic disease; NF-kB, nuclear factor kappa beta; 161 NLRP, NACHT, LRR and PYD domains-containing protein; Nrf2, nuclear factor 162 (erythroid-derived 2)-like 2; ox-/ac-LDL, oxidized-/acetylated-LDL; PD, protectin; 163 PDGF, platelet-derived growth factor; PMNs, polymorphonuclear neutrophils; 164 PUFA, polyunsaturated fatty acid; RAAS, renin-angiotensin-aldosterone system; RAGE, receptor for advanced glycation end-product; RCT, reverse cholesterol 165 transport; RF, renal fibrosis; ROS, reactive oxygen species; Rv, resolvin; SAID, 166 severe autoimmune diabetes; SFA, saturated fatty acid; SGLT-2, sodium glucose 167 cotransporter-2; SIDD, severe insulin-deficient diabetes; SIRD, severe insulinresistant diabetes; SMC, smooth muscle cell; SPM, specialized pro-resolving 168 mediator; SR-A, class A-macrophage scavenger receptor; STZ, streptozotocin; 169 T1D/T2D, type-1/type-2 diabetes; TF, tissue factor; TGF-\u00b31, transforming growth 170 factor-β1; Th, T helper; THBS-1, thrombospondin-1; TIF, tubulointerstitial fibrosis; TLR, toll-like receptor; VEGF, vascular endothelial growth factor. 171

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disease (Dutta and Mukhopadhyay, 2018). (4) *MOD:* Around a fifth of all participants were classified in cluster 4. These patients typically had high BMIs but they did not show insulin resistance (Kahn et al., 2006). (5) *Mild age-related diabetes (MARD):* Most of the patients (nearly 40%) in the cohort belonged to cluster 5. They were usually older adults with healthier metabolic profiles (including lower BMIs) than the other clusters (Ma et al., 2018).

Diabetes is associated with serious life-threatening and 236 life limiting complications. Acute complications include 237 hyperglycemia-induced ketoacidosis and hypoglycemia. The 238 chronic vascular complications of diabetes have a massive 239 impact on morbidity and mortality. These are classically 240 defined as microvascular and macrovascular complications and 241 reflect responses of susceptible individuals to hyperglycemia, 242 dyslipidemia and hypertension associated with diabetes 243 244 (Orban et al., 2018). The macrovascular complications include accelerated-CVD and accelerated-atherosclerosis, as discussed 245 below (Duncan et al., 2003). 246

Complications of the microvasculature include retinopathy, 247 neuropathy and nephropathy. DR is major cause of blindness in 248 the working class (Duh et al., 2017). Diabetic neuropathy 249 develops in almost half of all individuals with diabetes 250 and the lifetime risk of lower limb amputation as much 251 15% in certain populations. Diabetic neuropathy is as 252 a syndrome encompassing both somatic and autonomic 253 branches of the peripheral nervous system, and, furthermore, 254 contributes to the pathology of other diabetic complications, 255 such as impaired wound healing and erectile dysfunction 256 (Russell and Zilliox, 2014). As will be discussed in more 257 detail below, DKD is the leading cause of ESRD (Piccoli 258 et al., 2015). DKD typically develops over a long period 259 260 (decades) and, importantly, it is a major risk factor for the 261 development of macrovascular complications, including MI and stroke. 262

With best medical care the risk of major chronic complications 263 for T1D are cited as 47% for retinopathy, 17% for nephropathy 264 and 14% for CVD. These figures represent a lifetime risk. Figures 265 for T2D are more complex. Although death rates are higher for 266 people with diabetes, relative to age and sex matched cohorts, 267 a recent study has shown that in the United States, whereas 268 death rates for people with and without diabetes have fallen, 269 270 the greatest decline in mortality was actually seen in those with diabetes, presumably reflecting improved management of 271 glycemia, lipids and hypertension (Gregg et al., 2018). Moreover, 272 in a United Kingdom study, patients with T2D initiated on 273 metformin monotherapy had longer survival than did matched, 274 non-diabetic controls (Bannister et al., 2014). However, the 275 overall mortality in T2D is 60% higher than non-diabetic age and 276 sex matched controls. One consideration on these data is that 277 the lower rates reflect the relatively recent increase in incidence. 278 Mortality is typically associated with chronic complications, such 279 280 as DKD which develops over decades. The increased incidence may represent a timebomb of diabetes-associated mortality. 281 Indeed, among adults with diabetes, in the United States the 282 283 prevalence of ESRD has shown the smallest decrease as compared to other diabetic complications (Gregg et al., 2014). To an 284 extent this may reflect the efficacy of preventing atherosclerosis, 285

resulting in increased survival and increased opportunity to develop complications as a consequence of chronic exposure to hyperglycemia. As discussed below, it also reflects the need for therapeutic interventions to specifically target DKD and associated-RF. 290

This review will focus on describing recent advances in the understanding and elucidation of the underlying mechanisms and in exploring the potential of novel therapeutic approaches for treating diabetes-accelerated atherosclerosis, kidney disease and retinopathy, by using animal and human studies. For more comprehensive reviews of diabetic complications, readers are referred to several excellent recent reviews (Forbes and Cooper, 2013; Papatheodorou et al., 2016; Lotfy et al., 2017). 298

### DIABETES-ASSOCIATED ATHEROSCLEROSIS

### **Definition of DAA**

Atherosclerosis is a leading cause of vascular disease worldwide305and accounts for about 50% of all deaths in westernized societies306and 30% in developing countries (Fuster and Kelly, 2011). Its307major clinical manifestations include IHD and ischemic stroke308(Lusis, 2000), being, respectively, the world's first and third causes309of death (Barquera et al., 2015).310

The strong association between diabetes, low-grade 311 inflammation and atherosclerosis, accounts for one of the 312 major diabetes complications worldwide: DAA (Duncan et al., 313 2003). Approximately 50% of patients with T2D die prematurely 314 of a cardiovascular cause, and a further 10% die of renal failure 315 (van Dieren S et al., 2010). 316

Since Ross and Libby redefined atherosclerosis as a 317 progressive, chronic, dyslipidemic and also "inflammatory" 318 disease, advances in basic knowledge of this multifactorial 319 disease defined a key for inflammation in mediating all 320 the phases of athero-progression (Ross, 1999; Libby et al., 321 2002). Among the numerous markers of high- and low-322 grade inflammation, C-reactive protein predicts the risk of 323 atherosclerotic complications (see below) (Ross, 1999; Libby 324 et al., 2002). In the recent trial of anti-IL-1 $\beta$  antibody in a large 325 population of high risk atherosclerosis patients (CANTOS), the 326 intervention reduced inflammation and cardiovascular events. 327 Greatest impact was seen in those with highest baseline markers 328 of systemic inflammation. However, its efficacy was similar 329 in those with and without diabetes and, despite decreasing 330 inflammatory markers, did not reduce the incidence of diabetes 331 (Weber and von Hundelshausen, 2017; Everett et al., 2018). 332

## **Risk Factors for DAA**

The main modifiable risk factors for atherosclerosis have been 335 identified, and they include, but are not limited to, smoking, 336 adiposity, blood pressure, high levels of BMI, high level of LDL, 337 low level of HDL and diabetes (Herrington et al., 2016). T2D 338 is associated with an increased risk of CVD. A role for the 339 lipid-lowering therapy with statins for the primary prevention 340 of CVD in diabetes was demonstrated in CARDS, the first large 341 primary prevention study determining the action of statins in 342

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T2D patients, e.g., the efficacy of atorvastatin in preventing 343 disease irrespective of LDL levels (Colhoun et al., 2004). Over the 344 past two decades, developed countries have been able to reduce 345 the contribution of the above mentioned risk factors to mortality, 346 whereas developing countries show an increasing trend due to 347 high BMI and glucose (Barquera et al., 2015). 348

More recently, the prevalence of coronary atherosclerosis 349 was found to be higher in diabetic than in non-diabetic 350 patients and to be similar for diabetic individuals without 351 clinical CAD and non-diabetics with clinical CAD, implying 352 that prevention measures for asymptomatic diabetic individuals 353 should be similar to secondary preventive approaches among 354 355 non-diabetic population, as an aggressive prevention measure 356 for atherosclerosis in all diabetic patients, independently of their 357 CAD symptoms (Goraya et al., 2002).

#### 359 **Cellular Pathogenetic Mechanism of** 360 DAA 361

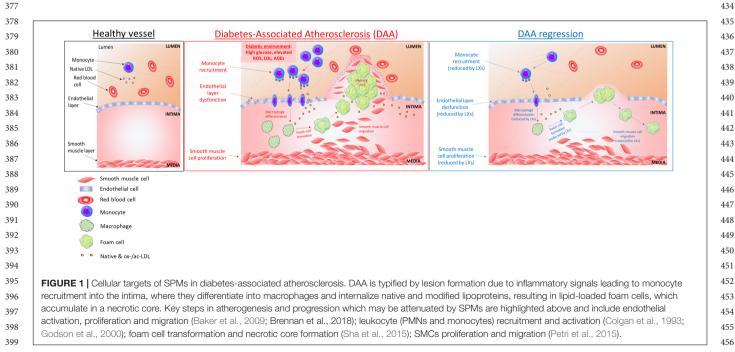
The pathogenesis of atherosclerosis shares several features 362 with other inflammatory diseases, including the infiltration 363 of monocytes and subsequent differentiation to macrophages 364 in response to locally generated signals (Scrivo et al., 2011). 365 At cellular and subcellular levels, inflammatory stimuli 366 or a disturbed blood flow induce endothelial dysfunction 367 (Cunningham and Gotlieb, 2005), altering the homeostatic 368 equilibrium depicted in Figure 1 (right). This vasoreactivity 369 allows lipoproteins apo-B to enter the intima and bind to 370 371 proteoglycans which trap the LDL particles and increase 372 their susceptibility to oxidation, acetylation and hydrolysis by secretory phospholipases thus amplifying the inflammatory 373 374 response, characterized by chemokine secretion and adhesion 375 molecules expression on ECs surface. These modifications of lipoprotein induce their aggregation in complexes and 376

subsequent retention, and, additionally, induce monocytes 400 recruitment, a crucial step in early phases of atherogenesis (Lusis, 401 2000). 402

Once chemoattracted to the inflammatory injury area, the 403 monocyte undergoes a series of processes that allow cell 404 locomotion (i.e., rolling, adhesion, polarization, crawling) to 405 reach the endothelial transmigration sites, in proximity of 406 low shear stressed athero-prone regions, where blood flow is 407 disturbed, such as bifurcations of arteries (Cunningham and 408 Gotlieb, 2005). Once transmigrated and eventually infiltrated into 409 the intima layer, monocytes differentiate into macrophages in 410 response to locally produced factors, such as M-CSF (Moore 411 and Tabas, 2011). This program of differentiation includes 412 upregulation of class A- macrophage scavenger receptor (SR-413 A), CD 36 (CD-36) and other cell surface receptors, to facilitate 414 ox-LDL or ac-LDL uptake: in physiologic conditions, this 415 process allows an efficient removal of excessive lipids from the 416 blood circulation ("scavenging" action of macrophage). When 417 homeostasis fails, dysregulation of this phenomenon leads to 418 uncontrolled accumulation of lipids and cholesterol derivatives 419 in macrophages, and their transformation into foam cells in the 420 subintima endothelial layer (Kunjathoor et al., 2002). 421

Lipid-loaded foam cells, cellular debris, calcium deposits 422 and connective tissue contribute to generate the so-called *fatty* 423 streak (a hallmark and first sub-clinical sign of atherosclerosis), 424 increasing inflammation and inducing necrosis and foam cell 425 death. As the lesion grows invade both the luminal space and the 426 intima. The necrotic area is confined within a fibrous cap made of 427 connective tissue, composed of SMCs and collagen. Fibrous cap 428 atheromas are the first clinically detectable atherosclerotic lesions 429 (Li and Glass, 2002). 430

As the lipid core increases in size, the fibrotic cap is invaded 431 by macrophages and lymphocytes, inducing the thinning of 432 the cap. The mechanism by which a sustained macrophage 433



invasion weakens the fibrous cap involves phagocytosis of the
ECM and the release of proteolytic enzymes (i.e., plasminogen
activators and matrix metalloproteinases, MMPs). The thinned
fibrous cap is prone to rupture, exposing the inflammatory and
thrombogenic molecules (TF, collagen) of the lipid core, highly
increasing the risk of thrombosis (Pepine, 1998).

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## Molecular Pathogenetic Mechanisms of DAA

As above stated, diabetes accelerates atherosclerosis, contributing 467 to higher rates of mortality and morbidity among diabetic 468 patients. The molecular mechanisms behind this likely reflects 469 increased inflammation and decreased blood flow (Abe and Berk, 470 471 2013) but are not fully understood. Several possible triggers have 472 been thus far hypothesized, including hyperglycemia, insulin resistance, increased activation of PDGF-dependent pathways, 473 increased level of TF or decreased level of HDL, and AGEs 474 and their receptors (RAGE) signaling activation (Beckman 475 et al., 2002). Insulin and hyperglycemia play key roles in 476 distinct phases of disease progression, via different mechanisms 477 and differentially affecting the three major cell types: SMCs, 478 macrophages and ECs (Bornfeldt and Tabas, 2011). 479

It has been hypothesized that, in advanced plaques, insulin 480 resistance may promote apoptosis of PMNs, SMCs, and 481 macrophages. In particular, death of SMCs can lead to the 482 thinning of the fibrotic cap, whereas death of macrophages is 483 associated with a defective phagocytic clearance of the cells 484 (efferocytosis), promoting plaque necrosis (Greenlee-Wacker, 485 2016). These processes converge to precipitate plaque rupture 486 and acute thrombotic vascular occlusion (Brophy et al., 2017). 487 488 A relation between diabetes obese-induced adiposity and 489 atherosclerosis in young adults have been observed (McGill et al., 1995). Elevated SFAs has been associated with obesity and insulin 490 resistance (Funaki, 2009) and causes defective efferocytosis of 491 apoptotic macrophage (Li et al., 2009), subsequently causing 492 a secondary cellular necrosis and inflammation amplifying the 493 plaque necrosis (Thorp and Tabas, 2009). The combined pro-494 apoptotic effect of macrophage insulin resistance and the anti-495 efferocytic effect of SFAs may create a "perfect storm" for plaque 496 necrosis, as proposed by Bornfeldt and Tabas (2011). 497

Hyperglycemia accelerates formation of early/mid stage 498 lesions of atherosclerosis by promoting an inflammatory 499 phenotype of which adhesion molecule expression in ECs 500 is a hallmark. Increased flux through the aldose reductase 501 pathway accelerates glucose metabolism and generates ROS. 502 Increased adhesion molecule expression leads to increased 503 monocyte/macrophage accumulation and atherogenesis. In 504 505 SMCs, a principal effect of increased glucose uptake appears to 506 be increased secretion of the monocyte chemoattractant protein-1, a chemokine which acts in concert with ECs. This leads 507 508 to an increased production of endothelium-derived contracting factors, which oppose the protective activity of nitric oxide 509 (Meininger et al., 2000; vanDam et al., 2000). Ultimately, this 510 leads to greater recruitment of monocytes into the growing 511 lesion (Bornfeldt and Tabas, 2011), thereby further contributing 512 to an enhanced inflammatory response. Those events have 513

been shown to promote adventitial inflammation and vasa 514 vasorum neovascularization in experimental models of diabetic 515 atherosclerosis. In particular, over the past two decades, the 516 work from Cosentino and Luscher (1998) has established the 517 strong relationship between hyperglycemia, oxidative stress and 518 inflammation, together with an increased risk of CVD in T2D 519 (Beckman et al., 2013; Paneni et al., 2013). Very recently, their 520 studies demonstrated epigenetic regulation of immune-metabolic 521 pathways to increased inflammation, neovascularization and 522 intraplaque hemorrhage in human diabetic atherosclerosis 523 (Guzik and Cosentino, 2018). 524

Insulin and hyperglycemia are not the only possible factors so far correlated to the underlying pathogenetic mechanism of DAA. Hyperglycemia enhances shear stress-induced platelet activation (Gresele et al., 2003). PDGF has been shown to play a major role in the pathology of vascular diseases. Inhibition of PDGF receptor activation attenuates DAA in experimental models (Lassila et al., 2004). 531

The inflammatory component of microangiopathic processes 532 is independently associated with plaque rupture, leading to 533 coronary thrombosis. TF, the most potent trigger of the 534 coagulation cascade, is increased in diabetic patients with 535 poor glycemic control. Circulating TF microparticles are also 536 associated with apoptosis of plaque macrophages, closing the link 537 among inflammation, plaque rupture and blood thrombogenicity 538 (Fallon et al., 1997; Singh et al., 2012). 539

AGE/RAGE signaling has been a well-studied cascade in 540 many different disease states, particularly diabetes. It heavily 541 influences both cellular and systemic responses to increase bone 542 matrix proteins through activation of PKC, p38 MAPK, TGFβ, 543 NF $\kappa\beta$  and ERK1/2 signaling pathways in both hyperglycemic 544 and calcification conditions. AGE/RAGE signaling has been 545 shown to increase oxidative stress and to promote diabetes-546 mediated vascular calcification through activation of NADPH 547 oxidase-1 and decreased expression of superoxide dismutase-1. 548 AGE/RAGE signaling in diabetes-mediated vascular calcification 549 is also attributed to increased oxidative stress resulting in the 550 phenotypic switch of SMCs to osteoblast-like cells in AGEs-551 induced calcification (Kay et al., 2016). HDL, responsible for 552 free cholesterol removal, are reduced in patients with insulin 553 resistance and diabetes, conditions for which the role of obesity 554 is highly detrimental (Rashid and Genest, 2007; Barter, 2011). In 555 addition to their role as lipid lowering agents, via inhibition of 556 3-hydroxy-3-methylglutaryl coenzyme A reductase, pleiotropic 557 responses to statins may include reduction of SMCs proliferation, 558 as observed in *in vitro* and *ex vivo* models (Fleg et al., 2008). 559

## **Current Therapies in DAA**

There are currently no available therapies for the regression 562 of atherosclerosis (Fuster et al., 1998). Therefore, new 563 therapeutic targets are needed in order to offer an alternative 564 type of intervention to invasive surgery, such as stenting 565 or endarterectomy. Current therapies in DAA adopted 566 antiplatelet/anticoagulant therapy, stabilizing the plaque 567 (Engelberg et al., 1956; Colwell, 1997), including the use of 568 low-dose aspirin (75-162 mg/day) for secondary prevention 569 of cerebrovascular and cardiovascular events in all diabetic 570

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patients (Angiolillo, 2009). Evidence that LDL causes CVDs is 571 overwhelming. It has also been proven beyond all doubt that 572 lowering the level of LDL using statins reduces cardiovascular 573 risk. However, many people remain at high risk even when 574 their level of LDL has been reduced by aggressive treatment 575 with statins. One reason for this residual risk can be a low 576 level of HDL, an independent, inverse predictor for CAD. It 577 has therefore been suggested that raising the level of HDL 578 should be considered as a therapeutic strategy for reducing the 579 residual cardiovascular risk that persists in some people, despite 580 aggressive LDL-cholesterol lowering with statins. HDL particles 581 have several functions with the potential to protect against 582 583 arterial disease, the best known of which relates to their ability to 584 promote cholesterol efflux from macrophages in the artery wall. 585 However, HDLs have several additional protective properties that 586 are independent of their involvement in cholesterol metabolism. For example, they have properties that reduce oxidation, vascular 587 inflammation and thrombosis, improve endothelial function, 588 promote endothelial repair, enhance insulin sensitivity and 589 promote insulin secretion by pancreatic  $\beta$  islet cells (Barter, 590 591 2011). These beneficial effects may be responsible for coronary plaque stabilization in patients treated with those molecules 592 which can up regulate HDL expression including Apo-A1 or 593 peroxisomal proliferator-activated receptors agonists, holding 594 great promise in the treatment of diabetic atherosclerosis. 595

### 597 The Regression of DAA

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The regression of existing lesions is the holy-grail in management of atherosclerosis. Over the past two decades major advances have been made to this end. Fisher's lab and his collaborators Young, Hazen, Smith and Moore have firmly established the principle that regression of atherosclerosis is a possible therapeutic goal (Fisher, 2016).

Although monocytes are recruited into the plaque during 604 its growth, they also have the capacity to emigrate from 605 atherosclerotic lesion. Using murine models of regression, 606 including the "transplantation mouse," a transplant model in 607 which plaque-bearing aortic segments are transferred into 608 normolipidemic mice (Reis et al., 2001); the "reversa mouse," 609 a genetic "switch" model in which LDL production can be 610 conditionally reduced in LDL- $R^{-/-}$  mice (Feig et al., 2011); and 611 acute treatment models, in which Apo- $E^{-/-}$  mice are injected 612 either with Apo-A1 (Hewing et al., 2014), with a microsomal 613 triglyceride transfer protein inhibitor or with an anti-microRNA 614 (miR) (anti-miR-33) (Rayner et al., 2011; Moore et al., 2013; 615 Distel et al., 2014) a decrease in plaque size and, consequently, 616 regression of pre-existing atherosclerosis was demonstrated 617 (Llodra et al., 2004; Randolph, 2008; Feig et al., 2009). A possible 618 explanation of the reduction in CD68<sup>+</sup> macrophage cell content 619 620 is that monocytes can enter the lymphatic system, reaching the lymph nodes, or they can migrate across the arterial 621 622 endothelium toward the artery lumen to directly enter the circulating bloodstream (Llodra et al., 2004; Randolph, 2008). 623 The main processes involved in atherogenesis are also the main 624 target for regression, namely, the retention of apo-B-containing 625 lipoproteins in the arterial wall and the reaction of macrophages 626 to these particles (Williams and Tabas, 2005). The resulting foam 627

cells secrete pro-inflammatory cytokines and chemokines, as 628 well as retention factors that amplify the inflammatory response 629 and promote macrophage *chemostasis*. These accumulating 630 macrophages experience endoplasmic reticulum stress, which, if 631 prolonged, results in apoptosis. This cell death, coupled with 632 defective efferocytosis, due to an uncontrolled lipid accumulation, 633 in which essentially SFAs decrease the fluidity of the plasma 634 membrane, leads to the formation of the necrotic core that is 635 characteristic of advanced plaques (Funaki, 2009; Thorp and 636 Tabas, 2009; Bornfeldt and Tabas, 2011). 637

The key mechanisms that promote regression are: lipid unload 638 of the foam cell and promotion of RCT, via upregulation of the 639 efflux protein ABCA-1 expression on plaque macrophages and 640 the subsequent cholesterol efflux toward exogenous acceptors 641 (i.e., Apo-E-containing HDL) (Chinetti-Gbaguidi et al., 2011); 642 a decrease in the expression of retention factors (Brodsky 643 and Fisher, 2008); a reduced monocyte recruitment via 644 their transformation in monocyte-derived dendritic cells and 645 subsequent upregulation of CC-chemokine receptor (CC-R)-646 7 on their surface, which allow emigration to the lymphatic 647 system, restoring permeabilization and reducing lymphatic vessel 648 fibrosis (Ivanov et al., 2016). Finally, the retention/migration 649 factors contributing to macrophage loss from the plaque, through 650 reverse transmigration to the lumen or through trafficking to the 651 adventitial lymphatic (Potteaux et al., 2011). 652

In the context of diabetes, as depicted in Figure 1 (central 653 panel), regression of atherosclerosis is impaired. High glucose 654 levels modulate LXR-dependent gene expression, by inhibiting 655 the LXR-dependent expression of ABCA1, but not ABCG1 656 (Hussein et al., 2015) and by inducing miR-33, a key negative 657 regulator of the RCT factors, ABCA1 and HDL (Wijesekara 658 et al., 2012). In mouse models of insulin-deficient diabetes, it 659 has been shown that leukocytosis (monocytosis and neutrophilia) 660 is hyperglycemia-dependent. The myelopoiesis is driven by 661 increased expression of certain DAMPs, specifically, signaling 662 through the pattern recognition AGE/RAGE. The relevance 663 to human health and disease is suggested by the correlation 664 between serum S100A8/S100A9 (the associated DAMPs) and the 665 incidence of CAD in a subset of T1D patients from the Pittsburgh 666 EDC study, highlighting the potential importance of glucose 667 control and lipid-lowering therapy as strategies to promote 668 regression of atherosclerosis in diabetics and also suggesting 669 a number of therapeutic targets, including disruption of the S100A8/S100A9-RAGE signaling axis (Nagareddy et al., 2013).

## DIABETIC KIDNEY DISEASE

### **Definition of DKD**

DKD typically develops over many decades. It is characterized 677 by progressive proteinuria (microalbuminuria 30-299 mg/24 h 678 to macroalbuminuria > 300 mg/24 h) with a subsequent decline 679 in glomerular filtration reflected by increased serum creatinine 680 (National Kidney Foundation, 2002). The pathophysiology 681 of DKD typically reflects the convergence of hemodynamic, 682 metabolic and inflammatory insults in susceptible individuals 683 (Harjutsalo and Groop, 2014). Current interventions focus on 684

tight glycemic control and RAAS blockade by ACE inhibition or angiotensin receptor antagonism to dilate the efferent arteriole and reduce glomerular hypertension. At best, these interventions slow the progress of disease (Forbes and Cooper, 2013). There is a growing appreciation that oxidative stress and inflammation are key drivers of DKD and may be appropriate targets for therapeutic intervention. Circulating inflammatory cytokine levels correlate with albuminuria and elevated levels of soluble TNF-receptor-1 is an independent predictor of decline in renal function (Krolewski et al., 2014). 

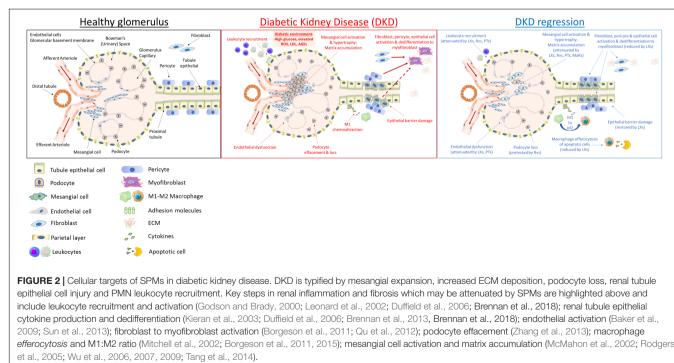
#### Cellular Pathogenetic Mechanisms of DKD

The physiological functionality of a healthy glomerulus is outlined in Figure 2 (left). Changes in renal hemodynamics, reflecting glomerular and systemic hypertension, arise early in DKD and lead to glomerular hyperfiltration. RAAS activation leads to increased angiotensin II and endothelin-1 causing efferent arteriolar vasoconstriction and hyperfiltration. Glomerular damage is characterized by podocyte effacement resulting in proteinuria. Renal hypertrophy is also observed in DKD reflecting accumulation of mesangial matrix, glomerular BM thickening and tubular hypertrophy. As matrix expands, it accumulates to form Kimmelstiel-Wilson nodules, a pathological feature of DKD. TIF is considered the major determinant of progression of DKD (Duffield, 2014). The mechanisms underlying TIF have been exhaustively investigated in the context of chronic kidney disease, including DKD (Leaf and Duffield, 2017). At a cellular level, several mechanisms have been proposed including activation of resident fibroblasts to matrix producing myofibroblasts, detachment of pericytes and

matrix production, recruitment of fibrocytes from bone marrow and EMT (Kalluri and Weinberg, 2009). Whereas the role of EMT in TIF has been questioned the loss of several epithelial cell markers (de-differentiation) has been observed together with expression of pro-fibrotic mediators such as CTGF and the TGF<sup>β1</sup> activator THBS-1 (Thiery et al., 2009). Experimental evidence suggests that partial EMT and chronic inflammation converge to create a profibrotic *milieu* facilitating collagen production by fibroblasts and recruited hematopoietic cells in the kidney (Zeisberg and Duffield, 2010; Buchtler et al., 2018). Glomerulosclerosis and TIF lead eventually to organ failure and a requirement for renal replacement therapy (hemodialysis or transplantation). Efforts to directly target inflammation in DKD have included manipulating chemokine and cytokine signals in T2D, such as antagonism of CCR2/CCR5 (Huh et al., 2018). 

## Molecular Pathogenetic Mechanisms of DKD

As depicted in Figure 2 (central panel), high glucose exerts specific toxic effects on the resident cells of the kidney, including specialized parietal epithelial cells (podocytes), mesangial cells, endothelia, fibroblasts and epithelia driving cellular dedifferentiation (Eddy and Neilson, 2006; Liu, 2011). Many of these responses are driven by autocrine and paracrine mediators released by target cells and infiltrating monocytes/macrophages, as typified by responses to TGF- $\beta$ 1 and its downstream targets, including CTGF (Murphy et al., 1999; Strutz et al., 2000; Boor and Floege, 2011). Hyperglycemia leads to ROS production and activation of inflammatory responses including NF-kB and janus kinases and signal transducer and activator of transcription 



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#### proteins (JAK-STAT) activation and subsequent downstream 799 cytokine production (Sifuentes-Franco et al., 2018). 801

#### Current Therapies in DKD 802

Numerous large scale genome-wide association studies have 803 been carried out in DKD over recent years (Sandholm et al., 804 2014; Ahlqvist et al., 2015; Teumer et al., 2015; Wuttke and 805 Köttgen, 2016; van Zuydam et al., 2018). These studies have 806 frequently implicated inflammatory pathways in the pathogenesis 807 of DKD. Such genetic validation of therapeutic targets includes 808 the JAK-STAT pathway. STAT-1,3 activation is observed in 809 renal biopsies from people with DKD (Berthier et al., 2008). 810 811 Baricitinib, a small molecule JAK-STAT inhibitor, has shown 812 efficacy in a small scale clinical trial. Treatment with baricitinib 813 was associated with decreased inflammatory biomarkers (e.g., 814 urinary chemokine CCL-2, plasma soluble tumor necrosis factor receptor-1, intracellular adhesion molecule-1 and serum amyloid 815 A). Baricitinib decreased albuminuria in participants with T2D 816 and DKD (Tuttle et al., 2018). 817

Glucose stimulates inflammasome assembly, caspase-1 818 activation and IL-1\beta release (Schroder et al., 2010). IL-1\beta 819 activation by the NACHT, LRR and PYD domains-containing 820 protein (NLRP) inflammasome is an important component of 821 CKD (Vilaysane et al., 2010). Blockade of IL-1β activity in post 822 MI patients with CKD reduced the risk of adverse cardiovascular 823 events among those with CKD (Ridker et al., 2017). As described 824 above, the CANTOS trial targeted IL-1ß in atherosclerosis 825 patients and the intervention reduced inflammation and 826 cardiovascular events (Ridker et al., 2011). Comparable effects 827 were observed among those with baseline albuminuria or 828 diabetes. Canakinumab, however, was without effect on serial 829 830 measures of eGFR, creatinine, the urinary albumin:creatinine 831 ratio or reported adverse renal events during trial follow-up (Ridker et al., 2018). 832

Despite the identification of numerous drivers of fibrosis, 833 such as the TGF superfamily, thus far efforts to target RF 834 per se have been unsuccessful. A recent double blind phase 835 II study assessed whether modulating TGF-\u00b31 activity with a 836 TGF-β1-specific, humanized, neutralizing monoclonal antibody 837 was effective in slowing renal function loss in patients with 838 diabetic nephropathy on RAAS inhibition treatment over a 12-839 month period. No significant impact on disease progression 840 was observed (Voelker et al., 2017). Other approaches have 841 focused on the balance between BMP family agonist-antagonist 842 activities and promoted BMP-7 and or small peptide mimetics, 843 such as THR-123 (Ali and Brazil, 2014; Tampe and Zeisberg, 844 845 2014; Brazil et al., 2015). However, some of these data have been controversial (Sugimoto et al., 2012). Systemic administration 846 847 of BMP-7 protein is problematic due to the low availability in 848 the kidney, explaining the need for a huge amount of BMP-7 for its reno-protective action, which might exert adverse effects 849 elsewhere (Vukicevic et al., 1998; Yanagita, 2012). CTGF/CCN2 850 has also been proposed as a potential target (Falke et al., 851 2014, 2017). Other therapeutic approaches which have been 852 853 proposed in the context of DKD include attenuation of NFkB signaling (Lee et al., 2012), breakdown of AGEs (Rabbani 854 and Thornalley, 2018) or RAGE antagonism (Bongarzone 855

et al., 2017). In this context, bardoxolone methyl is a novel 856 synthetic triterpenoid belonging to the antioxidant inflammation 857 modulator class. Antioxidant inflammation modulators potently 858 induce the antioxidant and cytoprotective transcription factor 859 Nrf2, reduce the pro-inflammatory activity of the IKK-β/NF-860  $\kappa B$  pathway, increase the production of antioxidant and 861 reductive molecules, and decrease oxidative stress, thereby 862 restoring redox homeostasis in areas of inflammation. Activation 863 of anti-oxidant responses via Nrf2 and inhibition of NF-864 kB by the triterpenoid bardoxolone methyl reduces oxidative 865 stress, inflammation and promotes mitochondrial function in 866 numerous experimental models of CKD, including DKD (Pergola 867 et al., 2011). Unfortunately, clinical trials of bardoxolone methyl 868 in patients with stage 4 CKD and T2D were prematurely 869 terminated for safety concerns (Tayek and Kalantar-Zadeh, 870 2013). Bardoxolone methyl treatment was associated with 871 approximately double the risk of heart failure as placebo. 872 Subsequent analysis suggests that these data did not represent 873 toxicity per se and that further development of this compound 874 may be warranted with more careful patient selection (Chin et al., 875 2014, 2018). 876

## Novel Therapeutic Approaches in DKD

It is important to note some recent advances that suggest 879 renoprotection in response to newer therapeutics which 880 regulate blood glucose and reduce cardiovascular risk in T2D. 881 Intriguingly these reno-protective responses may be independent 882 of glucose lowering. Such interventions include the incretin-883 based therapeutics (GLP-1-RAs, e.g., liraglutide or DPP-4 884 inhibitors and SGLT-2 inhibitors) enhancing glycemic control 885 with a low risk of hypoglycemia. However, the use of these agents 886 is limited in those with significant renal impairment. A recent 887 trial treatment with liraglutide, a GLP-1 analog, was associated 888 with a 22% lower incidence of doubling serum creatinine, 889 persistent macroalbuminuria, development of ESRD or death 890 from renal disease relative to controls (Mann et al., 2017). Similar 891 data have been reported for other GLP-1 receptor agonists and 892 for DPP-4 inhibitors which inhibit breakdown of endogenous 893 GLP-1. SGLT-2 inhibitors suppress glucose reabsorption by 894 the proximal tubule and therefore increase glucose excretion. 895 The SGLT-2 inhibitors target reabsorption of both glucose and 896 sodium as a result there is increased sodium delivery to the 897 macula densa activating tubule-glomerular feedback afferent 898 arteriolar vaso-modulation, resulting in increased renal blood 899 flow and decreased glomerular hyperfiltration. SGLT-2 inhibition 900 is associated with lower rates of albuminuria and lowering rates 901 of eGFR decline (Tomkin, 2014). Intriguingly, bariatric surgery 902 in T2D appears to have specific reno-protective effects which 903 may relate to enhanced GLP-1 responsiveness (Docherty and 904 le Roux, 2014). Miras et al. (2015) reported that, 1-year post-905 bariatric surgery, a decrease urinary albumin/creatinine ratio was 906 observed whereas no benefit was seen on other microvascular 907 complications, i.e., retinopathy or neuropathy. SGLT-2 inhibitors 908 and GLP-1 targeting drugs attenuate inflammatory responses 909 in DKD. As we will discuss below, we propose that exploiting 910 the bioactivity of endogenous lipid modulators that promote 911 the resolution of inflammation and suppress fibrosis is a novel 912

therapeutic paradigm worthy of consideration as adjuvant 913 therapy in DKD. 914 915

#### 916 THE ROLE OF MACROPHAGE IN 917 DIABETES COMPLICATIONS 918 919

As described, macrophages are key players in atherosclerotic 920 lesions, regulating the local inflammatory milieu and plaque 921 stability by the secretion of many inflammatory molecules, 922 growth factors and cytokines (Wolfs et al., 2011). The 923 macrophage paradigm classically reflects the heterogeneity of 924 their monocyte progenitor: alternative crawling monocytes 925 continuously patrol the endothelium of blood vessels in 926 927 the steady state. The patrolling monocytes rarely extravasate 928 in the steady state. In contrast, during inflammation or infection, classical monocytes are the first to extravasate when 929 inflammation signaling occurs, and, within few hours they 930 differentiate in M1 "pro-inflammatory macrophages," induced 931 by  $INF\gamma$  + LPS or by TNFa, characterized by a high 932 933 phagocytic profile. At later stages of inflammation, non-classical monocytes (or non-phlogistic monocytes) trans-migrate and 934 initiate a differentiation program into 'M2'-like macrophages, 935 which play a role in resolving of inflammation and tissue repair 936 (Geissmann et al., 2008). M2 macrophages can polarize toward 937 different phenotypes according to various stimuli present in 938 their surrounding micro-environment (Mosser and Edwards, 939 2008) and to their distinct gene expression profiles (Mantovani 940 et al., 2004). In particular, "M2a" or "alternative" macrophage 941 is the product of Th2 activation (by IL4 and IL13 cytokines 942 or fungal and helminth infections) and is responsible for a 943 944 type II inflammatory response (consisting in killing parasites 945 and inducing a Th2 response to allergy). "M2b" or "type II" macrophage is elicited by IL-1 receptor ligands, immune 946 complexes and LPS, triggering the activation of Th2 system. 947 "M2c" or "deactivated" macrophage is induced by IL10, TGF-β 948 and glucocorticoids and is mainly immunoregulatory, through 949 950 matrix deposition and tissue re-modeling (Martinez and Gordon, 2014). A fourth type, "M2d," or "angiogenic" macrophage is 951 elicited by IL-6 and adenosine and is mainly involved in wound 952 healing (Ferrante and Leibovich, 2012). However, recent findings 953 provide evidence for proliferation of local macrophages or trans-954 differentiation from other vascular cells as alternative sources 955 (Nagenborg et al., 2017). In particular, it has been shown that 956 cholesterol-loading induces the trans-differentiation of SMCs to 957 macrophage-like cells (Rong et al., 2001) and more recently, it 958 has been shown that approximately 50% of foam cells might have 959 a SMC origin (Allahverdian et al., 2014). Linear tracing studies 960 961 from Randolph's lab have shown that tissue-specific factors 962 drive highly specialized macrophage functions irrespective of their ontological origin, suggesting tremendous plasticity and 963 964 redundancy in the mononuclear phagocyte system. Whether embryonic and adult macrophages possess specialized roles has 965 yet to be formally tested. However, the conceptual understanding 966 and genetic tools are now sufficiently developed to precisely 967 follow both embryonic and adult macrophage subsets in health 968 and disease, which should allow important and unanswered 969

questions in the field to be addressed. In order to develop novel 970 therapies, a critical future goal is to harness this new found 971 understanding that different macrophage lineages exist within 972 tissues and clarify whether these distinct lineages differentially 973 contribute to tissue damage and repair (Epelman et al., 2014).

## Role of Macrophage in DAA

In the context of atherosclerosis, macrophages uniquely possess a 977 dual functionality, regulating lipid accumulation and metabolism 978 and sustaining the chronic inflammatory response, two well-979 documented pathways associated with the pathogenesis of the 980 disease (Moore and Tabas, 2011). 981

Established atherosclerotic plaques from patients with 982 existing CAD undergoing carotid endarterectomy classified 983 as symptomatic (where the patient has experienced previous 984 ischemic events but without any CVD diagnosis) or asymptomatic 985 (where a patient has no history of ischemic events or 986 CVD) have been recently comprehensive histologically and 987 immunohistochemically characterized for their cellular content 988 and macrophage subsets of atherosclerotic lesion. Symptomatic 989 plaques were defined as highly hemorrhagically active and 990 the internal carotid was the most diseased segment, based on 991 the predominant prevalence of fibrotic and necrotic tissue, 992 calcifications, and hemorrhagic events. Immunohistochemical 993 analysis showed that both M1 and M2 macrophages are present 994 in human plaques. However, M2 macrophages were localized 995 to more stable locations within the lesion. Importantly, M1 996 markers and Th 1-associated cytokines were highly expressed in 997 symptomatic plaques, whereas expression of the M2 markers, 998 mannose receptor and CD163 and Th2 cytokines were inversely 999 related with disease progression (de Gaetano et al., 2016). 1000 A strong relation between macrophage, mitochondria and 1001 glucose dysregulation has recently emerged in a number of 1002 studies from Fredman and Tabas (2017). Clearance of ACs by 1003 phagocytes (efferocytosis) prevents post-apoptotic necrosis and 1004 dampens inflammation. Mitochondrial fission in response to 1005 AC uptake is a critical process that enables macrophages to 1006 clear multiple ACs and to avoid the pathologic consequences of 1007 defective efferocytosis in vivo (Yurdagul et al., 2017). 1008

## Role of Macrophage in DKD

In a renal context, macrophages constitute a major subset of the 1011 infiltrating inflammatory cells and their contribution to renal 1012 fibrogenesis is well established (Duffield, 2010). Macrophage 1013 infiltration has been found to correlate with TIF on kidney 1014 biopsies (Young et al., 1995), and to correlate negatively with 1015 outcome in CKD of diverse etiologies (Tinckam et al., 2005; 1016 Duffield, 2010). However, the role of macrophages in this context 1017 is not entirely clear-cut. M1 macrophages are recruited to the 1018 kidney at early time points in a murine IRI model, whereas at 1019 later time points, M2 macrophages predominate. Additionally, 1020 in this model, depletion of macrophages prior to IRI has been 1021 found to attenuate inflammation and TIF, whereas macrophage 1022 depletion after 3-5 days is shown to slow tubular cell proliferation 1023 and repair (Lee et al., 2011). Macrophages exhibit some plasticity, 1024 and may not remain committed to a single phenotype. As a 1025 component of the programmed resolution of inflammation, a 1026

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phenotypic change is triggered by altered cytokine and lipid 1027 mediator profiles in the microenvironment, and M1 phenotype 1028 macrophages thus 'switch' to a pro-resolving M2 phenotype 1029 (Nathan and Ding, 2010; Lee et al., 2011). In the context of 1030 chronic inflammation, or repeated injuries, the factors that 1031 determine if macrophages are predominantly reparative versus 1032 predominantly pro-inflammatory remain unclear. Directing 1033 more of the macrophage population toward a pro-resolving 1034 phenotype may provide a novel therapeutic approach in CKD. 1035 Although much of our current understanding of the ontogeny 1036 and functional plasticity of macrophages has been derived from 1037 murine models, it is important to note that, together with the 1038 above mentioned contribution from Randolph on macrophage 1039 ontology, a recent study in human heart reveals two populations 1040 1041 of macrophages with different origins and functions: CCR2 1042 expressing macrophages are recruited from bone marrow and proliferate and are functionally proinflammatory and abundant 1043 in regions of scarring, whereas macrophages lacking CCR2 are 1044 maintained by local proliferation and express genes associated 1045 with tissue repair (Bajpai et al., 2018). 1046

## DIABETIC RETINOPATHY: A BRIEF OVERVIEW

## Definition, Pathogenesis, and Current Therapies

Diabetic retinopathy (DR) is a microvascular complication of
diabetes, clinically characterized by progressive alterations
in the microvasculature that lead to retinal ischemia,
neovascularization, altered retinal permeability and macular
edema. It is currently the leading cause of blindness in the adult
working population (Congdon et al., 2004; Yau et al., 2012).

The disease can be divided into two main stages - non-1061 proliferative retinopathy and proliferative retinopathy 1062 distinguished by the absence or presence of abnormal 1063 neovascularization, respectively. The final stage "proliferative 1064 retinopathy" is characterized by neovascularization of the disk 1065 or iris or vitreous hemorrhage or retinal detachment (Wilkinson 1066 et al., 2003). Macular edema or "diabetic maculopathy" can occur 1067 at both the non-proliferative and proliferative stages as a result 1068 of fluid accumulation under the macula. 1069

The pathological and morphological alterations associated with DR were long considered to be primarily microvascular in nature, as a result of hyperglycemia and the metabolic pathways it activates. The onset of clinically detectable DR is characterized by changes in the micro-vessels of the eye which includes thickening of the BM, loss of vascular permeability, loss of pericytes, capillary occlusions and microaneurysms (Xu et al., 2014).

However, recent studies have demonstrated that retinal neurodegeneration is a critical feature associated with the progression of the disease and may in fact precede the development of clinically detectable microvascular damage (Lieth et al., 2000; Puro, 2002).

Under pathological DR conditions, break-down of the BRBoccurs as a result of the presence of increased levels of vascular

permeability factors, such as VEGF in the vitreous of the eye. The resulting "leaky" vasculature leads to increased albumin flux into the retina and fluid accumulation resulting in macular edema and possible vessel hemorrhage (Klaassen et al., 2013).

Thickening of the vascular BM occurs early in the disease 1088 and represents one of the first histologically detectable structural 1089 alterations. Several biochemical alterations contribute to 1090 BM thickening in vivo. Increased expression of the matrix 1091 components of the BM, including fibronectin (Roy et al., 1996), 1092 collagen IV (Roy et al., 1994) and laminin (Ljubimov et al., 1996) 1093 can be detected long before the formation of diabetic lesions. BM 1094 turnover is tightly regulated by the delicate balance of synthesis 1095 and degradation of BM components by MMPs, urokinases and 1096 their inhibitors. This balance is disturbed during DR (Kowluru 1097 et al., 2012). 1098

Several interconnected biochemical pathways associated with 1099 hyperglycemia have been implicated in the pathogenesis of DR, 1100 including increased polyol pathway flux, increased hexosamine 1101 pathway flux and activation of protein kinase C. A crucial role 1102 is played by hyperglycemia-induced ROS production and AGEs 1103 formation (Forbes and Cooper, 2013). The retina is the most 1104 metabolically active tissue in the body, rendering it particularly 1105 susceptible to oxidative stress (Wu et al., 2014). Although all 1106 retinal cells express RAGE ubiquitously, retinal pericytes, in 1107 particular, have been shown to accumulate AGEs, contributing to 1108 BRB breakdown, which is in part accredited to pericyte loss, but 1109 also to AGE-induced leukocyte adherence to retinal ECs (Moore 1110 et al., 2003). 1111

Growing consensus is emerging in the predominant role 1112 of inflammation in the pathogenesis of DR (Rubsam et al., 1113 2018). The formation of AGEs and the activation of PKC 1114 have been implicated in the activation of pro-inflammatory 1115 mediators, such as NF-kB, connecting hyperglycemic-induced 1116 oxidative stress to inflammation. An increase in a number of pro-1117 inflammatory cytokines and chemokines has been demonstrated 1118 in both diabetic patients and models of experimental retinopathy 1119 (Doganay et al., 2002; Sato et al., 2009). Blocking the 1120 activity of pro-inflammatory cytokines (such as TNF-a, IL-1121 6, and IL-1) has shown beneficial effects in models of 1122 retinopathy. An IL-1 receptor antagonist reduces inflammatory 1123 responses in a rodent model of T2D (Vallejo et al., 2014) 1124 while breakdown of the BRB was completely ablated in 1125 a TNF- $\alpha$  knockout diabetic mouse (Huang et al., 2011). 1126 Chemokines, such as MCP-1 and IL-8, are also elevated in 1127 diabetic eye disease and contributed to neovascularization 1128 and fibrosis (Yoshida et al., 2003). However, their expression 1129 was reduced by inhibitors of VEGF, suggesting that the 1130 action of both MCP-1 and IL-8 are mediated through 1131 pathways involving VEGF. Hyperglycemic conditions also drive 1132 increased expression of a number of growth factors (including 1133 VEGF and TGF $\beta$ ) mediating the retinal damage associated 1134 with DR, such as BM thickening, vascular permeability and 1135 neovascularization. 1136

TNF-a and VEGF have received particular attention 1137 for their role in the vascular lesion and neovascularization 1138 associated with late stage retinopathy. Therefore, anti-TNFa (i.e., 1139 Infliximab) (Sfikakis et al., 2005) and anti-VEGF (i.e., Avastin) 1140 1141 (Haritoglou et al., 2006) intravitreal therapies are standard 1142 clinical therapeutic options for the treatment of DR.

Many of the agents developed to target the various biochemical pathways driven by hyperglycemia have had limited effect clinically, pointing to a need for new therapeutics targets.

# THE ROLE OF SPMs IN RESOLVING INFLAMMATION

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The inflammatory response consists of two phases: initiation 1151 and resolution. The initiation phase is characterized by the site-1152 specific accumulation and coordinated activation of a host of 1153 immune effector cells in an inflammatory cytokine and pro-1154 1155 inflammatory lipid mediator rich environment. Inflammation is 1156 critical in the host response to infection and injury, however, timely resolution is necessary for the restoration of tissue 1157 homeostasis, thereby limiting excessive tissue injury, preventing 1158 the development of a chronic inflammatory state (Serhan et al., 1159 2007). Non-resolving inflammation is a major driver of disease. 1160 Multiple mechanisms ensure physiological resolution toward 1161 tissue homeostasis. Cells like macrophages switch phenotypes 1162 by secreting molecules like reactive oxygen intermediates, lipids 1163 and proteins which impact a cell from displaying pro- or an 1164 anti-inflammatory behaviors (Buckley et al., 2014). 1165

Whereas inflammation and its effective outcome, i.e., a return 1166 to homeostasis, were typically considered a manifestation of 1167 the passive dissipation of pro-inflammatory stimuli, including 1168 lipids, such as prostaglandins and leukotrienes, it is now clear 1169 that the resolution of inflammation is an active and dynamically 1170 regulated process reflecting responses to endogenously generated 1171 1172 mediators, including cytokine and lipids (Godson and Brady, 1173 2000; Maderna and Godson, 2009; Serhan, 2014).

The specialized SPMs are a family of endogenously produced 1174 pro-resolving lipid mediators derived from the metabolism of 1175 PUFAs, which include LXs, resolvins (Rvs), protectins (PDs) 1176 1177 and MaRs. They were discovered by Serhan et al. (1984). LXs (Lipoxygenase interaction products) were firstly isolated in a 1178 human leukocyte (Serhan et al., 1984) and classified as derivatives 1179 of the  $\omega 6$  fatty acid arachidonic acid (20:4, n-6). Rvs (*Resolution* 1180 phase interaction products) were firstly identified in a resolving 1181 inflammatory exudate in 2000 (Serhan et al., 2000), PDs (termed 1182 neuroprotectin D1 if generated in neural tissue for its protection 1183 in neurons, glial cells, and brain stroke; or protectin D1 for other 1184 tissue in 2004 (Bazan, 2005) and MaRs (Macrophage mediator 1185 in Resolving Inflammation) in 2009 (Serhan et al., 2009). Rvs, 1186 PDs and MaRs are classified as derivatives of w3 fatty acids: 1187 specifically, Rvs can either form from the EPA (20:5, n-3) [RVs 1188 1189 E-series] or from the DHA (22:6, n-3) [RVs D-series]; while, PDs 1190 and MaRs only derive from DHA. As their precursors, all these derivatives are classified as PUFAs and they demonstrated potent 1191 anti-inflammatory and immunoregulatory actions (Serhan et al., 1192 2008). 1193

Within a few hours from barrier break, tissue injury or trauma, eicosanoids are crucial in initiating the cardinal signs of inflammation (redness, heat, pain and swelling). As part of the vascular response, leukocytes traffic to the site of injury. The prostaglandins PGE<sub>2</sub> and PGI<sub>2</sub> (involved in vasodilation) 1198 and the leukotriene LTB4 (involved in chemotaxis and adhesion) 1199 stimulate the migration of PMN to the tissue. In parallel 1200 to the PMN-monocyte sequence, lipid mediator composition 1201 of the inflammatory exudate switches class, from eicosanoids 1202 to SPMs, marking the beginning of the end of the acute 1203 inflammatory response. LXs are the first SPM to be locally 1204 produced, highlighting its role as "stop" signal to eicosanoid 1205 production (exudate switch), as firstly described by the work of 1206 Levy et al. (2001). LXs and Rvs also stimulate the recruitment 1207 of monocytes. The resolving macrophages then clear apoptotic 1208 PMNs, inflammatory debris by efferocytosis (stimulated by 1209 LXs, Rvs, PDs). After this has taken place, normal structure 1210 and homeostasis can be restored. "Resolution" is defined as 1211 the period between peak inflammatory cell influx and the 1212 clearance of these cells from the tissue site and the restoration 1213 of functional homeostasis. Subsequent post-resolution events 1214 involves activation of adaptive immunity B- and T-lymphocytes 1215 (Fullerton and Gilroy, 2016). 1216

Failed resolution can lead to increased levels of prostaglandinsand leukotrienes, chronic inflammation and fibrosis. UltimatelySPMs reduce the magnitude and duration of inflammation (Aritaet al., 2007), stimulate re-epithelialization (Hellmann et al., 2018),wound healing (Dalli, 2017) and tissue regeneration (Dalli, 2017).

While most of the studies involving SPMs have been 1222 conducted on rodents models, major and recent advances have 1223 been represented by the work of Motwani et al. (2016) in 1224 humans, where a new translational model of self-resolving acute 1225 inflammatory response triggered by the intradermal injection 1226 of UV-killed Escherichia coli into the forearm of healthy 1227 volunteers was described. For the first time SPMs endogenous 1228 production have been identified in humans over the course of the 1229 inflammatory response. It has also been shown that resolution is 1230 an active process accelerated by addition of exogenous SPMs. 1231

The molecular mechanisms through which SPMs exert their 1232 responses include activation of distinct GPCRs and regulation 1233 of gene expression. The binding, and consequent activation, 1234 of the LX/N-formyl peptide receptor-2 (ALX/FPR2) GPCR by 1235 lipids, such as LXA<sub>4</sub> and RvD1 as well as Annexin-1 peptide 1236 (Krishnamoorthy et al., 2010; Maderna et al., 2010; Bena et al., 1237 2012), and the RvE1 agonism at the ChemR23 GPCR (Arita 1238 et al., 2007) are key to reduce PMN infiltration and subsequently 1239 stimulate efferocytosis by macrophages, heralding the initiation of 1240 pro-resolving cascade of events. 1241

### **SPMs IN ACUTE INJURIES**

The anti-inflammatory and pro-resolving properties of SPMs, including LXs, Rvs and their mimetics, particularly 15(R/S)-methyl-LXA<sub>4</sub> (Wu et al., 2013), benzo-LXA<sub>4</sub> (Sun et al., 2009), BDA-RvD1 (Orr et al., 2015) and have been demonstrated in several types of experimental acute renal and peritoneal injury (see below).

Moreover, SPMs have recently been shown to play a key 1252 role in dampening both sterile inflammation and infection (or 1253 non-sterile inflammation). In this context, a recent advance 1254

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is represented by the above mentioned study on the selfresolving properties of SPMs in an acute and local *E. coli*-induced
translational skin-blisters model (Motwani et al., 2016).

## Biosynthesis and Functions of LXs in Acute Injuries

Native LXs, LXA<sub>4</sub> and LXB<sub>4</sub>, are endogenous eicosanoids, 1262 transcellularly biosynthesized by 5- and 15-LO interaction 1263 of activated leukocytes with epithelium, endothelium or 1264 platelets (Serhan et al., 1984; Serhan, 1989; Serhan and 1265 Sheppard, 1990) Acetylation of cyclooxygenase-2 by aspirin 1266 1267 can trigger the biosynthesis of their 15R-carbon epimers, 15-epi-LXA<sub>4</sub> and 15-epi-LXB<sub>4</sub> [15-epi-LXs or aspirin-triggered 1268 LXs (ATLs)] (Serhan, 2005). Although native LXs have 1269 1270 demonstrated potent anti-inflammatory and pro-resolution bioactions (Claria et al., 1996; Fierro and Serhan, 2001; 1271 Chandrasekharan and Sharma-Walia, 2015), their therapeutic 1272 potential is compromised for by their chemical instability and 1273 for by their rapid metabolic inactivation by prostaglandin 1274 1275 dehydrogenase-mediated metabolic inactivation in vivo (Clish et al., 2000), with the growing need to synthesize 1276 their mimetics. 1277

First-generation synthetic LXA<sub>4</sub> analogs were designed in 1278 1995-1998 by Serhan, Petasis and colleagues to minimize 1279 metabolism of the molecule (Parkinson, 2006). These 1280 relatively stable pharmacological agents, together with 1281 myeloid-specific ALX-R-expressing transgenic mice, have 1282 provided powerful tools to explore LX functions in vivo. 1283 1284 Among those, pharmacokinetic analysis of ATLa, such as methvl (5R.6R.7E.9E.11Z.13E.15S)-16-(4-fluorophenoxy)-1285 1286 5,6,15-trihydroxy-7,9,11,13-hexadecatetraenoate, revealed 1287  $\beta$ -oxidation as a novel route for LXA<sub>4</sub> metabolism, prompting the development of second-generation 3-oxa-LXA4 analogs with 1288 improved pharmacokinetic disposition (Parkinson, 2006). 1289

1290 Second-generation *3-oxa-LXA*<sub>4</sub> *analogs*, such as 1291 (5R,6R,7E,9E,11Z,13E,15S)-16-(4-fluorophenoxy)-3-oxa-

1292 5,6,15-trihydroxy-7,9,11,13-hexadecatetraenoic acid, have shown
1293 potency and efficacy comparable to ATLa in diverse animal
1294 models after topical, intravenous or oral delivery (Guilford and
1295 Parkinson, 2005).

More recently, a new class of LX-analogs featuring a benzo-1296 fused ring system have been designed and proved to be as 1297 potent as native LXA4 in a series of in vitro and in vivo 1298 studies (O'Sullivan et al., 2007; Petasis et al., 2008). In particular, 1299 it was found to stimulate phagocytosis of apoptotic PMN by 1300 1301 macrophages, in a zymosan-induced peritonitis murine model of acute inflammation (O'Sullivan et al., 2007). Further exploration 1302 1303 of the mechanism of action through which PMN phagocytosis 1304 by bone marrow-derived macrophage was elicited revealed that expression, activation and internalization of ALX/FPR2 by LXA4 1305 and the glucocorticoid-derived Annexin A1 peptide (Ac2-26) 1306 were essential (Maderna et al., 2010). 1307

In early 2000, the work from Leonard et al. (2002) suggested a framework for understanding SPMs bioactions in renal IRI and the molecular basis for renoprotection by LXs in this setting. They firstly demonstrated, in a murine renal IRI, that the stable synthetic LXA<sub>4</sub> analog 15-epi-16-(FPhO)-LXA<sub>4</sub>-1312 Me is reno-protective, as gauged by lower serum creatinine, 1313 attenuated leukocyte infiltration and reduced morphologic 1314 tubule injury. Subsequently, they employed complementary 1315 oligonucleotide microarray and bioinformatic analyses to probe 1316 the transcriptomic events that underpin LX renoprotection 1317 and found that epi-LXA4 modified the expression of many 1318 differentially expressed pathogenic mediators, including 1319 cytokines, growth factors, adhesion molecules and proteases. 1320 Importantly, this LX-modulated transcriptomic response 1321 included many genes expressed by renal parenchymal cells (such 1322 as the Claudin family epithelial tight junctions) (Kieran et al., 1323 2003). 1324

## Biosynthesis and Functions of Rvs in Acute Injuries

Rvs are produced by 12/15-LO, p450, and/or 5-LO, in trans-1329 cellular or intracellular biosynthetic systems of leukocytes or 1330 leukocytes plus endothelia/epithelia (Serhan et al., 2000). The 1331 novel lipid mediators produced from EPA were first isolated 1332 from resolving exudates that proved to contain 18R-HEPE as 1333 well as several other related bioactive compounds and were 1334 therefore collectively named 18R-E series (Serhan et al., 2000). 1335 The first bioactive product isolated from exudates, coined 1336 RvE1, reduced inflammation and blocked human PMN trans-1337 endothelial migration. 1338

RvDs are derived from DHA. During inflammation, 1339 endogenous DHA is converted to 17S-HEPE which are 1340 then converted in 17S-hydroxyl-containing RvDs (RvD1-RvD6) 1341 and docosa-conjugated triene-containing PD1/NPD1, via 15-LO 1342 (15S-lipoxygenation)-initiated biochemical pathways (Serhan 1343 et al., 2002; Hong et al., 2003; Marcheselli et al., 2003) or to 14S 1344 hydroxyl-containing MaRs via 12-LO (12S-lipoxygenation)-1345 initiated biochemical pathways. 5-LO catalyzes sequentially with 1346 15-LO or 12/15-LO, generating RvDs (Hong et al., 2003) and 1347 some MaRs (Serhan et al., 2009). 1348

RvD1 is converted by eicosanoid oxidoreductases to 17-oxo-1349 RvD1 and 8-oxo-RvD1. The former is an inactive metabolite, 1350 while the latter is still effective in suppressing PMN infiltration 1351 (Sun et al., 2007). RvE1 is metabolized to 12-oxo-RvE, 18-1352 oxo-RvE1, 10,11-dihydroxy RvE, 19-hydroxy RvE1, 20-hydroxy 1353 RvE1 in tissue or cells, of which the first four metabolites are 1354 inactive partially or completely in inflammation resolution, and 1355 thus are representative of RvE1 metabolic deactivation (Arita 1356 et al., 2006; Hong et al., 2008). Human PMNs convert PD1 1357 to its omega-22 hydroxy product (Serhan and Petasis, 2011). 1358 The metabolic deactivation of Rvs dysregulated in pathological 1359 conditions, may result in their deficiency, or in diminishing the 1360 pharmacological efficacy of administered resolvins. Therefore, a 1361 series of stable analogs have been successfully synthesized, such 1362 as a *p*-fluorophenoxyl added to RvE1 and RvD1 ω-terminal, 1363 which blocks the critical metabolic inactivation of RvE1 or 1364 RvD1 without attenuating the anti-inflammatory pro-resolving 1365 activities (Arita et al., 2006; Hong et al., 2008; Tang et al., 2014) 1366 In particular, the RvE1 analog 19-(p-fluorophenoxy)-RvE1 1367 was synthesized to resist rapid metabolic inactivation and 1368

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proved to retain biological activity reducing PMN infiltration 1369 and pro-inflammatory cytokine/chemokine production in vivo. 1370 These results established the structure of a novel RvE1 1371 initial metabolite, indicating that conversion of RvE1 to 1372 the oxo product represents a mode of RvE1 inactivation. 1373 Moreover, the designed RvE1 analog, which resisted further 1374 metabolism/inactivation, could be a useful tool to evaluate 1375 the actions of RvE1 in complex disease models (Arita et al., 1376 20061377

These lipids act as paracrine and autocrine mediators of leukocytes to promote resolution of acute injuries, including AKI-initiated inflammation and fibrosis and rescue of kidney functions (Zhao et al., 2016), by shortening PMN life span and promoting macrophage *efferocytosis* of ACs and the subsequent exit of the phagocytes from inflammatory tissue.

1384 RvD1 and RvE1 also switch macrophage to the phenotype produces pro-resolving interleukin-10. RvDs that 1385 or protectin/neuroprotectin D1 (PD1/NPD1) inhibits PMN 1386 infiltration into injured kidney, blocks TLR-mediated 1387 inflammatory activation of macrophage and mitigates renal 1388 dysfunction. RvDs also repress renal interstitial fibrosis, and PD1 1389 promotes reno-protective heme-oxygenase-1 expression. These 1390 findings provide novel approaches for targeting inflammation 1391 resolution and LMs or modulation of LM-associated pathways 1392 for developing better clinical treatments for AKI. Moreover, in 1393 LPS-induced AKI, RvD1 could decrease TNFa level, ameliorate 1394 kidney pathological injury, protect kidney function, and improve 1395 animal survival by down-regulating NFkB inflammatory signal 1396 as well as inhibiting renal cell apoptosis (Zhao et al., 2016). 1397 Intriguingly, RvE1 counter-regulates leukocytes partially via 1398 increased LXA<sub>4</sub> biosynthesis (Levy et al., 2011). Since AKI 1399 1400 is the major complication of renal allograft transplantation 1401 (Bellomo et al., 2012), these results further demonstrate the effectiveness of LXA4 or RvE1 in reducing AKI. LX actions 1402 converge with the pro-resolving characteristics of RvD1, as 1403 LXA4 and RvD1 both activate the same GPCRs ALXR/FPR2 and 1404 GPR32. 1405

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## 1408 SPMs IN CHRONIC DIABETES1409 COMPLICATIONS

Unresolved inflammation drives the development of clinically
relevant chronic diseases. Here, we focus the attention on the role
of SPMs, particularly LXs and Rvs, on DAA, CKD and, briefly, on
DR.

As discussed previously, sustained, non-resolved low-1415 grade inflammation, over decades, promotes formation of 1416 1417 atherosclerotic lesions characterized by large necrotic cores, thin 1418 fibrous caps and thrombosis. In advanced atherosclerosis, there is an imbalance between levels of SPMs and proinflammatory 1419 lipid mediators, which results in sustained leukocyte influx into 1420 lesions, inflammatory macrophage polarization, and impaired 1421 efferocytosis. In animal models of advanced atherosclerosis, 1422 1423 restoration of SPMs limits plaque progression by suppressing inflammation, enhancing efferocytosis, and promoting an 1424 increase in collagen cap thickness (Fredman and Tabas, 2017). 1425

From a CKD-perspective, there is a clear mechanistic 1426 link between non-resolving inflammation and fibrosis. Non-1427 resolving inflammation results in sustained secretion of pro-1428 fibrotic cytokines and other inflammatory mediators from both 1429 resident and infiltrating cells, eliciting fibroblast proliferation 1430 and epithelial cell de-differentiation. Sustained or unresolved 1431 inflammation is recognized to be an underlying component of 1432 many chronic disease states in diverse organ systems, including 1433 CKD (Serhan, 2014; Brennan et al., 2017). 1434 Q14

## The Role of LXs and Rvs in Atherosclerosis and DAA

It is now established that the local LO-induced biosynthesis of lipid mediators, including LXA<sub>4</sub>, RvD1 and PD1, protects against atherosclerosis. These mediators exert potent agonist actions on macrophages and vascular ECs that can control the magnitude of the local inflammatory response (Merched et al., 2008), as depicted in **Figure 1** (left).

Enhanced biosynthesis of LXA<sub>4</sub> in transgenic mice is 1445 associated with decreased lesion formation in models of 1446 atherosclerosis (Merched et al., 2008). Atheroprotective 1447 responses of macrophages and ECs to SPMs include enhanced 1448 efferocytosis of apoptotic debris and modulation of adhesion 1449 molecules expression (VCAM-1, ICAM-1, P-Sel). It has been 1450 shown that LXA<sub>4</sub> increases ABCA1 expression and promotes 1451 cholesterol efflux through LXRa pathway in THP-1 macrophage-1452 derived foam cells (Sha et al., 2015). Moreover, it has been 1453 recently demonstrated that ATL signals through FPR2/ALX in 1454 vascular SMCs and protects against intimal hyperplasia after 1455 carotid ligation (Petri et al., 2015). 1456

Over the past few years, Brennan's work focused on the role 1457 of miR in both DKD (see details below) and DAA. The let-1458 7 miRNA family plays a key role in modulating inflammatory 1459 responses. Vascular SMC proliferation and EC dysfunction are 1460 critical in the pathogenesis of atherosclerosis, including in the 1461 setting of diabetes. The therapeutic potential of LXA4-induced 1462 restoration of let-7 mimic levels was observed in vitro in 1463 SMCs, in vivo via tail vein injection in a 24 h murine model, 1464 and ex vivo, where significant changes to the secretome in 1465 response to let-7 therapy were seen. It has been proposed that 1466 restoration of let-7 expression, a mimic of response to LXA<sub>4</sub>, 1467 could provide a new target for an anti-inflammatory approach 1468 in diabetic vascular disease (Brennan et al., 2017). Very recently, 1469 LXA<sub>4</sub> and the synthetic LX mimic benzo-LXA<sub>4</sub> have also been 1470 shown to be athero-protective in murine model of DAA (STZ-1471 induced diabetic Apo $E^{-/-}$  mouse). Here there was significant 1472 reduction in plaque area. The authors also demonstrated that 1473 these SPMs could attenuate vascular SMCs migration and 1474 proliferation, EC-monocytes interactions, as well as modulate the 1475 pro-inflammatory secretome signature in human carotid plaque 1476 explants. Of particular note was the finding that LX treatment 1477 reduced pre-existing plaque burden in diabetic mice (Brennan 1478 et al., 2018). 1479

Oxidation of native LDLs plays an important role in the 1480 development of atherosclerosis. A very recent work showed 1481 that although ox-LDLs are known to be pro-inflammatory 1482

and deleterious in the context of atherosclerosis, they are also 1483 able to induce a pro-resolution effect by self-induction of 1484 RvD1 from HMEC (Dufour et al., 2018). Moreover, circulating 1485 1486 inflammation-resolving lipid mediators RvD1 and DHA are decreased in patients with acutely symptomatic carotid disease 1487 (Bazan et al., 2017). Similarly, RvE1 and ATL plasma levels were 1488 found to be significantly lower in symptomatic peripheral arteries 1489 disease than in healthy controls (Ho et al., 2010). 1490

#### In addition to lipid agonists, the ALX/FPR2 can also bind 1491 peptides, such as Annexin-1 (Maderna et al., 2010). In an 1492 advanced model of atherosclerosis, the Annexin-1 derivative 1493 acetylated peptide (Ac2-26), was delivered using Collagen IV-1494 targeted nanoparticles and it showed therapeutic effect in fat-fed 1495 LDL- $R^{-/-}$ mice, including an increase in the protective collagen 1496 layer overlying lesions, suppression of oxidative stress and a 1497 decrease in plaque necrosis, thus, suggesting a new form of 1498 therapy (Fredman et al., 2015). 1499

#### <sup>1500</sup> 1501 The Role of LXs in CKD and DKD

Advances in understanding the effects of LXs in the context of 1502 RF arose from investigating their actions on the main cell types 1503 involved in kidney failure (mesangial cells, fibroblasts, epithelia, 1504 adipocytes) (see details below). As outlined in Figure 2 (right), 1505 work from Rodgers, McMahon and Mitchell investigated the 1506 potential of LXA<sub>4</sub> to regulate PDGF-induced gene expression 1507 and the associated autocrine TGF<sup>β1</sup> production in human renal 1508 mesangial cells, and found that LXA4 is a potent modulator of 1509 matrix accumulation and pro-fibrotic change, thus suggesting a 1510 potential protective role in progressive renal disease (McMahon 1511 et al., 2002; Mitchell et al., 2004; Rodgers et al., 2005). In an 1512 experimental model of RF, i.e., unilateral ureteric obstruction 1513 1514 (UUO), LXA<sub>4</sub> and its synthetic benzo-analog attenuated injury 1515 by inhibiting TGFβ1-induced fibroblast activation, proliferation and gene expression (Borgeson et al., 2011). 1516

Aging, defined as a state of chronic, low-grade, sterile 1517 inflammation (inflamm-aging) (Franceschi et al., 2017) and 1518 adiposity, have recently been proposed as one of the major 1519 risk factors underlying the pathophysiological development 1520 of obesity-associated complications, including T2D, and its 1521 complications DAA and DKD (Todd et al., 2015). Therefore, 1522 of particular relevance in the diabetes context is the work that 1523 Borgeson et al. (2011) subsequently carried out, in 2012 and 1524 2015, on the effect of the native LXA4 on obesity-induced adipose 1525 tissue inflammation and related diseases. Firstly, using a model 1526 of age-associated adipose inflammation, inflamm-aging it was 1527 shown that LXA<sub>4</sub> attenuates adipose inflammation, decreasing 1528 1529 IL-6 and increasing IL-10 expression. The altered cytokine milieu correlated with increased the insulin-regulated glucose 1530 1531 transporter-4 and the insulin receptor substrate-1 expression, 1532 suggesting improved insulin sensitivity. Further investigations revealed the ability of LXA4 to rescue macrophage-induced 1533 desensitization to insulin-stimulated signaling and glucose 1534 uptake in cultured adipocytes, thus suggesting that LXA4 may 1535 represent a potentially useful and novel therapeutic strategy 1536 1537 to subvert adipose inflammation and insulin resistance, key components of T2D (Borgeson et al., 2012). Later on, the role 1538 of LXs in obesity-related pathologies was further explored by 1539

investigating their impact on impaired glucose tolerance, adipose 1540 inflammation, fatty liver and CKD. In particular, LXs attenuated 1541 obesity-induced CKD, reducing glomerular expansion, mesangial 1542 matrix and urinary H<sub>2</sub>O<sub>2</sub>. These data suggested a protective role 1543 for LXs against obesity-induced systemic disease, and supported 1544 a novel therapeutic paradigm for treating obesity and associated 1545 pathologies, such as TD2 and its related complications (Borgeson 1546 et al., 2015). A role in the context of aging-related pathologies 1547 (including obesity, atherosclerosis, renal disease and diabetes) 1548 for SPMs has been also recently reviewed by Doyle et al. 1549 (2018).1550

Certain miRs have been implicated in fibrosis (both renal 1551 and cystic). In cultured HK-2 cells, LXA4 suppresses TGF-1552 1-induced RF through a mechanism involving upregulation 1553 of the miR let-7c and downregulation of TGF R1. Expression 1554 of let-7c targets is dysregulated in human RF (Brennan et al., 1555 2013). The effects of let-7 on TGF 1-mediated responses of 1556 renal epithelia have also been shown by others, including 1557 Cooper and Kantharidis, leading to the proposal that let-1558 7b miR represents a potential new target for the treatment 1559 of RF in diabetic and non-diabetic nephropathy (Wang 1560 et al., 2014; Kantharidis et al., 2015; Brennan et al., 2017). 1561 Interestingly, LXA4 demonstrated to attenuate TGF- 1-1562 induced fibrotic responses whereby epithelial cells express 1563 mesenchymal markers (Brennan et al., 2013). In cultured 1564 renal epithelia upregulation of thrombospondin and CTGF is 1565 a well-documented fibrotic response (Crawford et al., 1998; 1566 Liu et al., 2013). While, in cystic fibrosis, miR181b is indeed 1567 downregulated by LXA4 and RvD1, through ALX/FPR2 1568 activation (Pierdomenico et al., 2015). Moreover, very recent 1569 interesting observations showed that LXs can also reverse 1570 established atherosclerosis (Brennan et al., 2018) and DKD 1571 (Brennan et al., 2018). 1572

Very recently, in a DKD murine model, Brennan has 1573 also identified a series of transcripts regulated by LXA4 and 1574 Benzo-LXA<sub>4</sub>, modulating well established (TGF-β1, PDGF, 1575 TNF- $\alpha$ , NF- $\kappa\beta$ ) and novel (early growth response-1) networks 1576 in DKD, demonstrating that LXs can reverse established 1577 diabetic complications and supporting a therapeutic paradigm 1578 to promote the resolution of inflammation (Brennan et al., 1579 2018). Interestingly, a recent study from Goicoechea measured 1580 circulating level of ATL in patients with diabetic and non-1581 diabetic kidney disease and found that diabetes was associated 1582 with lower levels of the SPMs and that this could be restored 1583 by 12-month low dose aspirin treatment (Goicoechea et al., 1584 2017). 1585

## The Role of Rvs in Diabetic Wound Healing

The work from Spite greatly deepened the knowledge around the SPMs properties of re-epithelialization and/or re-vascularization post ischemia, particularly focussing on Rvs bioactions. RvD2 stimulates arteriogenic revascularization in a murine model of hind limb ischemia suggesting that resolvins may be a novel class of mediators that both resolve inflammation and promote arteriogenesis (Zhang et al., 2016), a mechanism

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which can provide protection against nephropathy and 1597 atherosclerosis. 1598

1599 Altered resolution of acute inflammation in the context of obesity and diabetes, in which PMN apoptosis is 1600 delayed and macrophage efferocytosis is defective, cause 1601 persistent leukocyte and AC accumulation and defective 1602 wound closure (Baltzis et al., 2014). Wound healing 1603 in diabetes is enhanced by RvD1 and RvE1 via the 1604 promotion of macrophage-mediated AC clearance and 1605 re-epithelialization (Bannenberg et al., 2005; Spite et al., 1606 2014). Moreover, RvD1 decreases adipose tissue macrophage 1607 accumulation and improves insulin sensitivity in obese-1608 diabetic mice, suggesting that RvD1 could provide a novel 1609 therapeutic strategy for treating obesity-induced diabetes 1610 1611 (Hellmann et al., 2011).

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#### The Role of LXs and Rvs in DR 1613

1614 Although the anti-inflammatory (anti-TNF $\alpha$  and anti-VEGF) 1615 approach is still the standard therapy for DR, recent in vitro and 1616 in vivo models are shifting the attention toward a pro-resolving 1617 novel strategy (Das, 2013; Wang and Daggy, 2017).

1618 Since corneal, retinal neuronal degeneration (Srinivasan et al., 1619 2017), conjunctivitis (Stuebiger et al., 2015) and uveitis (Sivaraj 1620 et al., 2009) have been associated with DR, the effects of LXs 1621 (Gronert, 2005; Biteman et al., 2007; He et al., 2011; Hodges et al., 1622 2017) and Rvs (Tian et al., 2009; Settimio et al., 2012; Li et al., 1623 2013: Lee et al., 2015) in dampening DR are of relevance.

1624 In a well established in vivo model of STZ-induced Diabetes, 1625 hyperglycemia induces persistent inflammation and tissue 1626 damage, due to decreased expression of heme-oxygenase (HO) in the ciliar body (Rossi et al., 2006). Recently, the effect of 1627 1628 RvD1on STZ-induced DR has been explored. RvD1 regulates the 1629 NLRP3 inflammasome and NFkB signaling pathway (Yin et al., 1630 2017).

1631 Moreover, by using an in vivo deletion of 12/15-LOX model, 1632 associated with exacerbated inflammation and impaired wound 1633 healing, due to a failure of HO-1 induction, it has been 1634 demonstrated that LXA<sub>4</sub>, restored the HO synthesis and activity, 1635 rescuing the wound healing phenotype (Biteman et al., 2007).

Overall, the therapeutic potential of SPMs in the treatment of 1636 1637 DR are promising.

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#### REALIZING THERAPEUTIC POTENTIAL 1640

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The therapeutic challenges presented by diabetes-associated 1642 complications such as DAA and DKD are well documented, and 1643 experimental evidence, as outlined above, suggests a role for 1644 1645 SPM-based mimetics as adjuvants to current therapies. Clinical 1646 trials specifically investigating the therapeutic potentials of LXs and Rvs have been limited. 1647

In a randomized controlled trial, AT-LXA<sub>4</sub> and a 1648 comparatively stable analog of LXB<sub>4</sub>, 15R/S-methyl-LXB<sub>4</sub>, 1649 reduced the severity of eczema in a study of 60 infants 1650 1651 (Wu et al., 2013).

A synthetic analog of RvE1 is in clinical phase III testing 1652 for the treatment of the inflammation-based dry eye syndrome; 1653

along with this study, other clinical trials using an RvE1 1654 analog to treat various conditions are underway, such as 1655 in a single study where inhaled LXA4 decreased LTC4-1656 initiated bronchoprovocation in patients with asthma (Basil 1657 and Levy, 2016). RvE1, Mar1 and NPD1 are in clinical 1658 development studies for the treatment of neurodegenerative 1659 diseases and hearing loss (Serhan et al., 2015; Basil and Levy, 1660 2016). 1661

A clinical trial phase-I evaluating the effects of n-3 fatty acid 1662 supplementation on plasma SPMs in patients with CKD showed 1663 that endogenous production of SPMs was increased after 8-weeks 1664 n-3 fatty acid supplementation in patients with CKD, potentially 1665 impacting also patient risk of CVD complications (Mas et al., 1666 2016). 1667

More recently, Gilroy introduced the above mentioned 1668 first translational cantharidin-induced skin blister model 1669 in healthy male volunteers, providing insights into the 1670 mechanisms of self-resolving infections in humans, 1671 identifying cells and soluble mediators that may control the 1672 resolution phase. Further use of this model will improve 1673 our understanding of the evolution and resolution of 1674 inflammation in humans, how defects in these over-lapping 1675 pathways may contribute to the variability in disease 1676 longevity/chronicity, and lends itself to the screen of putative 1677 anti-inflammatory or pro-resolution therapies (Motwani et al., 1678 2016). 1679

## SUMMARY, CONCLUSIONS, AND FUTURE PERSPECTIVES

Aging populations, increasing urbanization and widening social 1685 inequalities are all contributing factors to the rapid rise in 1686 diabetes prevalence seen over the past 40 years worldwide. 1687 Reducing premature mortality from non-communicable diseases, 1688 including diabetes, has become a global priority. For people 1689 with either T1D or T2D, advances in clinical care, such as 1690 development of better glucose-lowering drugs and structured 1691 education programs promoting life-style changes, have led 1692 to considerable increases in life expectancy. Effectively, more 1693 people are living with diabetes for longer. Understanding 1694 the disease course, onset of complications, and comorbid 1695 conditions is critical to improving specialized care for people with 1696 diabetes. 1697

The most prevalent complications are affecting the 1698 microvascular (DKD, DR) and the macrovascular (DAA) 1699 systems. As mortality from cardiovascular complications 1700 continues to decline, attention must be turned to 1701 identifying, preventing, and treating other diabetes 1702 complications. In this context, advances in research in 1703 the molecular biology of such complications unveiled 1704 novel players and novel unified mechanisms driving 1705 different diabetes related complications. As highlighted 1706 here, inflammation is central to these processes. Evidence 1707 is accumulating that agonism of resolution of inflammation 1708 is a rational and tractable target that may be an attractive 1709 adjuvant in the context of chronic complications of 1710

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diabetes applying a novel therapeutic paradigm to a vast and growing unmet need.

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## 1715 AUTHOR CONTRIBUTIONS

REFERENCES

MdG, EB, and CG conceived and designed the review article.
MdG prepared the first draft of the manuscript. CM, AC, JH,
and EB contributed to pre-publication data. All authors read and
approved the final manuscript.

#### 1721

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- Abe, J. I., and Berk, B. C. (2013). Athero-prone flow activation of the SREBP2-NLRP3 inflammasome mediates focal atherosclerosis. *Circulation* 128, 579–582.
   doi: 10.1161/CIRCULATIONAHA.113.004390
- Abegunde, D. O., Mathers, C. D., Adam, T., Ortegon, M., and Strong, K. (2007).
  The burden and costs of chronic diseases in low-income and middle-income countries. *Lancet* 370, 1929–1938. doi: 10.1016/S0140-6736(07)61696-1
- Ahlqvist, E., Van Zuydam, N. R., Groop, L. C., and McCarthy, M. I. (2015). The genetics of diabetic complications. *Nat. Rev. Nephrol.* 11:277. doi: 10.1038/ nrneph.2015.37
- Ali, I. H. A., and Brazil, D. P. (2014). Bone morphogenetic proteins and their antagonists: current and emerging clinical uses. *Br. J. Pharmacol.* 171, 3620–3632. doi: 10.1111/bph.12724
- Allahverdian, S., Chehroudi, A. C., McManus, B. M., Abraham, T., and Francis,
   G. A. (2014). Contribution of intimal smooth muscle cells to cholesterol accumulation and macrophage-like cells in human atherosclerosis. *Circulation* 129, 1551–1559. doi: 10.1161/CIRCULATIONAHA.113.005015
- Angiolillo, D. J. (2009). Antiplatelet therapy in diabetes: efficacy and limitations of current treatment strategies and future directions. *Diabetes Care* 32, 531–540. doi: 10.2337/dc08-2064
- Arita, M., Oh, S. F., Chonan, T., Hong, S., Elangovan, S., Sun, Y. P., et al. (2006).
   Metabolic inactivation of resolvin E1 and stabilization of its anti-inflammatory actions. *J. Biol. Chem.* 281, 22847–22854. doi: 10.1074/jbc.M603766200
- 1743 Arita, M., Ohira, T., Sun, Y. P., Elangovan, S., Chiang, N., and Serhan, C. N.
- (2007). Resolvin E1 selectively interacts with leukotriene B4 receptor BLT1 and ChemR23 to regulate inflammation. J. Immunol. 178, 3912–3917. doi: 10.4049/ jimmunol.178.6.3912
- Bajpai, G., Schneider, C., Wong, N., Bredemeyer, A., and Hulsmans, M. (2018). The
  human heart contains distinct macrophage subsets with divergent origins and
  functions. *Nat. Med.* 24, 1234–1245. doi: 10.1038/s41591-018-0059-x
- Baker, N., O'Meara, S. J., Scannell, M., Maderna, P., and Godson, C. (2009).
   Lipoxin A4: anti-inflammatory and anti-angiogenic impact on endothelial cells.
   *J. Immunol.* 182, 3819–3826. doi: 10.4049/jimmunol.0803175
- 1751Baltzis, D., Eleftheriadou, I., and Veves, A. (2014). Pathogenesis and treatment1752of impaired wound healing in diabetes mellitus: new insights. Adv. Ther. 31,1753817–836. doi: 10.1007/s12325-014-0140-x
- Bannenberg, G. L., Chiang, N., Ariel, A., Arita, M., Tjonahen, E., Gotlinger, K. H.,
  et al. (2005). Molecular circuits of resolution: formation and actions of resolvins
  and protectins. *J. Immunol.* 174, 4345–4355. doi: 10.4049/jimmunol.174.7.
  4345
- Bannister, C. A., Holden, S. E., Jenkins-Jones, S., Morgan, C. L., Halcox, J. P.,
  Schernthaner, G., et al. (2014). Can people with type 2 diabetes live longer than
  those without? A comparison of mortality in people initiated with metformin
  or sulphonylurea monotherapy and matched, non-diabetic controls. *Diabetes. Obes. Metab.* 16, 1165–1173. doi: 10.1111/dom.12354
- Barquera, S., Pedroza-Tobías, A., Medina, C., Hernández-Barrera, L., Bibbins-Domingo, K., Lozano, R., et al. (2015). Global overview of the epidemiology of atherosclerotic cardiovascular disease. *Arch. Med. Res.* 46, 328–338. doi: 10.1016/j.arcmed.2015.06.006
- Barter, P. J. (2011). The causes and consequences of low levels of high density
  lipoproteins in patients with diabetes. *Diabetes Metab J.* 35, 101–106. doi: 10.
  4093/dmj.2011.35.2.101
- 1767

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- 1779 Basil, M. C., and Levy, B. D. (2016). Specialized pro-resolving mediators: 1780 endogenous regulators of infection and inflammation. *Nat. Rev. Immunol.* 16, 1781
- 51-67. doi: 10.1038/nri.2015.41782Bazan, H. A., Lu, Y., Jun, B., Fang, Z., Woods, T. C., and Hong, S. (2017).<br/>Circulating inflammation-resolving lipid mediators RvD1 and DHA are<br/>decreased in patients with acutely symptomatic carotid disease. Prostaglandins<br/>Leukot. Essent. Fatty Acids 125, 43–47. doi: 10.1016/j.plefa.2017.08.0071782
- Bazan, N. G. (2005). Neuroprotectin D1 (n.d.): a DHA-derived mediator that protects brain and retina against cell injury-induced oxidative stress. Brain Pathol. 15, 159–166. doi: 10.1111/j.1750-3639.2005.tb00513.x
- Beckman, J. A., Creager, M. A., and Libby, P. (2002). Diabetes and atherosclerosis:
  epidemiology, pathophysiology, and management. *JAMA* 287, 2570–2581.
  doi: 10.1001/jama.287.19.2570
  1790
- Beckman, J. A., Paneni, F., Cosentino, F., and Creager, M. A. (2013). Diabetes and vascular disease: pathophysiology, clinical consequences, and medical therapy: part II. *Eur. Heart J.* 34, 2444–2452. doi: 10.1093/eurheartj/eht142
- Bellomo, R., Kellum, J. A., and Ronco, C. (2012). Acute kidney injury. *Lancet* 380, 756–766. doi: 10.1016/S0140-6736(11)61454-2 1794
- Bena, S., Brancaleone, V., Wang, J. M., Perretti, M., and Flower, R. J. (2012).
   Annexin A1 interaction with the FPR2/ALX receptor: identification of distinct domains and downstream associated signaling. J. Biol. Chem. 287, 24690–24697.
   doi: 10.1074/jbc.M112.377101
- Berthier, C. C., Zhang, H., Schin, M., Henger, A., Nelson, R. G., Yee, B., et al. (2008).
   Enhanced expression of JAK-STAT pathway members in human diabetic nephropathy. *Diabetes* 58, 469–477. doi: 10.2337/db08-1328
- Biteman, B., Hassan, I. R., Walker, E., Leedom, A. J., Dunn, M., Seta, F., et al. (2007).
   Interdependence of lipoxin A4 and heme-oxygenase in counter-regulating inflammation during corneal wound healing. *FASEB J.* 21, 2257–2266.
   doi: 10.1096/fj.06-7918com
- Bongarzone, S., Savickas, V., Luzi, F., and Gee, A. D. (2017). Targeting the receptor for advanced glycation endproducts (RAGE): a medicinal chemistry perspective. *J. Med. Chem.* 60, 7213–7232. doi: 10.1021/acs. jmedchem.7b00058
- Boor, P., and Floege, J. (2011). Chronic kidney disease growth factors in renal fibrosis. Clin. Exp. Pharmacol. Physiol. 38, 441–450. doi: 10.1111/j.1440-1681.
   2011.05487.x
- Borgeson, E., Docherty, N. G., Murphy, M., Rodgers, K., Ryan, A., O'Sullivan, T. P., et al. (2011). Lipoxin A(4) and benzo-lipoxin A(4) attenuate experimental renal fibrosis. *FASEB J.* 25, 2967–2979. doi: 10.1096/fj.11-185017
- Borgeson, E., Johnson, A. M., Lee, Y. S., Till, A., Syed, G. H., Ali-Shah, S. T., et al. (2015). Lipoxin A4 attenuates obesity-induced adipose inflammation and associated liver and kidney disease. *Cell Metab.* 22, 125–137. doi: 10.1016/j. 1813
   cmet.2015.05.003
- Borgeson, E., McGillicuddy, F. C., Harford, K. A., Corrigan, N., Higgins, D. F., Maderna, P., et al. (2012). Lipoxin A4 attenuates adipose inflammation. *FASEB* J. 26, 4287–4294. doi: 10.1096/fj.12-208249

Bornfeldt, K. E., and Tabas, I. (2011). Insulin resistance, hyperglycemia, and atherosclerosis. *Cell Metab.* 14, 575–585. doi: 10.1016/j.cmet.2011.07.015 1819

- Brazil, D. P., Church, R. H., Surae, S., Godson, C., and Martin, F. (2015). BMP signalling: agony and antagony in the family. *Trends Cell Biol.* 25, 249–264. doi: 10.1016/j.tcb.2014.12.004
- Brennan, E., Wang, B., McClelland, A., Mohan, M., Marai, M., Beuscart, O., et al.
   (2017). Protective effect of let-7 miRNA family in regulating inflammation in

1883

1887

1888

1889

1907

1908

1909

- 1825 diabetes-associated atherosclerosis. Diabetes Metab. Res. Rev. 66, 2266-2277. doi: 10 2337/db16-1405 1826
- Brennan, E. P., Cacace, A., and Godson, C. (2017). Specialized pro-resolving 1827 mediators in renal fibrosis. Mol. Aspects Med. 58, 102-113. doi: 10.1016/j.mam. 1828 2017 05 001
- 1829 Brennan, E. P., Mohan, M., McClelland, A., de Gaetano, M., Tikellis, C., Marai, M., et al. (2018a). Lipoxins protect against inflammation in diabetes-associated 1830 atherosclerosis. Diabetes 67, 2657-2667. doi: 10.2337/db17-1317 1831
- Brennan, E. P., Mohan, M., McClelland, A., Tikellis, C., Ziemann, M., Kaspi, A., 1832 et al. (2018b). Lipoxins regulate the early growth response-1 network and 1833 reverse diabetic kidney disease. J. Am. Soc. Nephrol. 29, 1437-1448. doi: 10. 1834 1681/ASN.2017101112
- Brennan, E. P., Nolan, K. A., Borgeson, E., Gough, O. S., McEvoy, C. M., Docherty, 1835 N. G., et al. (2013). Lipoxins attenuate renal fibrosis by inducing let-7c and 1836 suppressing TGFbetaR1. J. Am. Soc. Nephrol. 24, 627-637. doi: 10.1681/ASN. 1837 2012060550
- 1838 Brodsky, J. L., and Fisher, E. A. (2008). The many intersecting pathways underlying 1839 apolipoprotein B secretion and degradation. Trends Endocrinol. Metab. 19, 1840 254-259. doi: 10.1016/j.tem.2008.07.002
- Brophy, M. L., Dong, Y., Wu, H., Rahman, H. N. A., Song, K., and Chen, H. (2017). 1841 Eating the dead to keep atherosclerosis at bay. Front. Cardiovasc. Med. 4:2. 1842 doi: 10.3389/fcvm.2017.00002
- 1843 Buchanan, T. A., and Xiang, A. H. (2005). Gestational diabetes mellitus. J. Clin. Invest. 115, 485-491. doi: 10.1172/JCI200524531 1844
- Buchtler, S., Grill, A., Hofmarksrichter, S., Stockert, P., Schiechl-Brachner, G., 1845 Rodriguez Gomez, M., et al. (2018). Cellular origin and functional relevance 1846 of collagen I production in the kidney. J. Am. Soc. Nephrol. 29, 1859-1873. 1847 doi: 10.1681/ASN.2018020138
- 1848 Buckley, C. D., Gilroy, D. W., and Serhan, C. N. (2014). Proresolving lipid mediators and mechanisms in the resolution of acute inflammation. Immunity 1849 40, 315-327. doi: 10.1016/i.immuni.2014.02.009 1850
- Chandrasekharan, J. A., and Sharma-Walia, N. (2015). Lipoxins: nature's way to 1851 resolve inflammation. J. Inflamm. Res. 8, 181-192.

1852 Chin, M. P., Bakris, G. L., Block, G. A., Chertow, G. M., Goldsberry, A., Inker, 1853 L. A., et al. (2018). Bardoxolone methyl improves kidney function in patients 1854 with chronic kidney disease stage 4 and type 2 diabetes: post-hoc analyses from bardoxolone methyl evaluation in patients with chronic kidney disease and type 1855 2 diabetes study. Am. J. Nephrol. 47, 40-47. doi: 10.1159/000486398

1856 Chin, M. P., Reisman, S. A., Bakris, G. L., O'Grady, M., Linde, P. G., McCullough, 1857 P. A., et al. (2014). Mechanisms contributing to adverse cardiovascular events in 1858 patients with type 2 diabetes mellitus and stage 4 chronic kidney disease treated with bardoxolone methyl. Am. J. Nephrol. 39, 499-508. doi: 10.1159/000362906 1859

- Chinetti-Gbaguidi, G., Baron, M., Bouhlel, M. A., Vanhoutte, J., Copin, C., Sebti, Y., 1860 et al. (2011). Human atherosclerotic plaque alternative macrophages display 1861 low cholesterol handling but high phagocytosis because of distinct activities 1862 of the PPARG and LXRa pathways. Circ. Res. 108, 985-995. doi: 10.1161/ 1863 CIRCRESAHA.110.233775
- Claria, J., Lee, M. H., and Serhan, C. N. (1996). Aspirin-triggered lipoxins (15-1864 epi-LX) are generated by the human lung adenocarcinoma cell line (A549)-1865 neutrophil interactions and are potent inhibitors of cell proliferation. Mol. Med. 1866 2, 583-596. doi: 10.1007/BF03401642
- 1867 Clish, C. B., Levy, B. D., Chiang, N., Tai, H. H., and Serhan, C. N. (2000). 1868 Oxidoreductases in lipoxin A4 metabolic inactivation: a novel role for 15onoprostaglandin 13-reductase/leukotriene B4 12-hydroxydehydrogenase in 1869 inflammation. J. Biol. Chem. 275, 25372-25380. doi: 10.1074/jbc.M00286 1870 3200
- 1871 Colgan, S. P., Serhan, C. N., Parkos, C. A., Delp-Archer, C., and Madara, J. L. (1993). 1872 Lipoxin A4 modulates transmigration of human neutrophils across intestinal epithelial monolayers. J. Clin. Invest. 92, 75-82. doi: 10.1172/JCI116601 1873
- Colhoun, H. M., Betteridge, D. J., Durrington, P. N., Hitman, G. A., Neil, H. A., 1874 Livingstone, S. J., et al. (2004). Primary prevention of cardiovascular disease 1875 with atorvastatin in type 2 diabetes in the Collaborative Atorvastatin Diabetes 1876 Study (CARDS): multicentre randomised placebo-controlled trial. Lancet 364,
- 1877 685-696. doi: 10.1016/S0140-6736(04)16895-5 Colwell, J. A. (1997). Aspirin therapy in diabetes. Diabetes Care 20:1767. 1878 doi: 10.2337/diacare.20.11.1767
- 1879 Congdon, N., O'Colmain, B., Klaver, C. C., Klein, R., Munoz, B., Friedman, 1880 D. S., et al. (2004). Causes and prevalence of visual impairment 1881

among adults in the United States. Arch. Ophthalmol. 122, 477-485. doi: 10.1001/archopht.122.4.477

- Cosentino, F., and Luscher, T. F. (1998). Endothelial dysfunction in diabetes 1884 mellitus. J. Cardiovasc. Pharmacol. 32(Suppl 3), S54-S61. 1885
- Cunningham, K. S., and Gotlieb, A. I. (2005). The role of shear stress in the 1886 pathogenesis of atherosclerosis. Lab. Invest. 85, 9-23. doi: 10.1038/labinvest. 3700215
- Dalli, J. (2017). Does promoting resolution instead of inhibiting inflammation represent the new paradigm in treating infections? Mol. Aspects Med. 58, 12-20. doi: 10.1016/j.mam.2017.03.007
- 1890 Das, U. N. (2013). Lipoxins, resolvins, and protectins in the prevention and treatment of diabetic macular edema and retinopathy. Nutrition 29, 1-7. 1891 doi: 10.1016/j.nut.2012.02.003 1892
- de Gaetano, M., Crean, D., Barry, M., and Belton, O. (2016), M1- and M2-1893 type macrophage responses are predictive of adverse outcomes in human 1894 atherosclerosis. Front. Immunol. 7:275. doi: 10.3389/fimmu.2016.00275
- 1895 Distel, E., Barrett, T. J., Chung, K., Girgis, N. M., Parathath, S., Essau, C. C., 1896 et al. (2014). mir33 inhibition overcomes deleterious effects of diabetes on atherosclerosis plaque regression in mice. Circ. Res. 115:759. doi: 10.1161/ 1897 CIRCRESAHA,115,304164 1898
- Docherty, N. G., and le Roux, C. W. (2014). Improvements in the metabolic milieu 1899 following Roux-en-Y gastric bypass and the arrest of diabetic kidney disease. 1900 Exp. Physiol. 99, 1146-1153. doi: 10.1113/expphysiol.2014.078790
- Doganay, S., Evereklioglu, C., Er, H., Turkoz, Y., Sevinc, A., Mehmet, N., et al. 1901 (2002). Comparison of serum NO, TNF-alpha, IL-1 $\beta$ , sIL-2R, IL-6 and IL-8 1902 levels with grades of retinopathy in patients with diabetes mellitus. Eye 16, 1903 163-170. doi: 10.1038/sj/eye/6700095 1904
- Doyle, R., Sadlier, D. M., and Godson, C. (2018). Pro-resolving lipid mediators: agents of anti-ageing? Semin Immunol. doi: 10.1016/j.smim.2018.09.002, [Epub 1905 ahead of print]. doi: 10.1016/j.smim.2018.09.002 1906
- Duffield, J. S. (2010). Macrophages and Immunologic Inflammation of the Kidney. Seminars in Nephrology. Amsterdam: Elsevier.
- Duffield, J. S. (2014). Cellular and molecular mechanisms in kidney fibrosis. J. Clin. Invest. 124, 2299-2306. doi: 10.1172/JCI72267
- Duffield, J. S., Hong, S., Vaidya, V. S., Lu, Y., Fredman, G., Serhan, C. N., 1910 et al. (2006). Resolvin D series and protectin D1 mitigate acute kidney injury. 1911 J. Immunol. 177, 5902-5911. doi: 10.4049/jimmunol.177.9.5902 1912
- Dufour, D., Khalil, A., Nuyens, V., Rousseau, A., Delporte, C., Noyon, C., et al. 1913 (2018). Native and myeloperoxidase-oxidized low-density lipoproteins act in 1914 synergy to induce release of resolvin-D1 from endothelial cells. Atherosclerosis 1915 272, 108-117. doi: 10.1016/j.atherosclerosis.2018.03.012
- Duh, E. J., Sun, J. K., and Stitt, A. W. (2017). Diabetic retinopathy: current 1916 understanding, mechanisms, and treatment strategies. JCI Insight 2:e93751. 1917 doi: 10.1172/jci.insight.93751 1918
- Duncan, B. B., Schmidt, M. I., Pankow, I. S., Ballantyne, C. M., Couper, D., Vigo, A., 1919 et al. (2003). Low-grade systemic inflammation and the development of type 2 diabetes: the atherosclerosis risk in communities study. Diabetes Metab. Res. 1920 Rev. 52, 1799-1805. doi: 10.2337/diabetes.52.7.1799 1921
- Dutta, D., and Mukhopadhyay, S. (2018). Novel diabetes subgroups. Lancet 1922 Diabetes Endocrinol. 6:438. doi: 10.1016/S2213-8587(18)30129-3 1923
- Eddy, A. A., and Neilson, E. G. (2006). Chronic kidney disease progression. J. Am. Soc. Nephrol. 17, 2964-2966. doi: 10.1681/ASN.2006070704 1924
- Engelberg, H., Kuhn, R., and Steinman, M. (1956). A controlled study of the 1925 effect of intermittent heparin therapy on the course of human coronary 1926 atherosclerosis. Circulation 13, 489-498. doi: 10.1161/01.CIR.13.4.489 1927
- Epelman, S., Lavine, K. J., and Randolph, G. J. (2014). Origin and functions of tissue 1928 macrophages. Immunity 41, 21-35. doi: 10.1016/j.immuni.2014.06.013
- 1929 Everett, B. M., Donath, M. Y., Pradhan, A. D., Thuren, T., Pais, P., Nicolau, J. C., et al. (2018). Anti-inflammatory therapy with canakinumab for the prevention 1930 and management of diabetes. J. Am. Coll. Cardiol. 71, 2392-2401. doi: 10.1016/ 1931 j.jacc.2018.03.002 1932
- Falke, L. L., Goldschmeding, R., and Nguyen, T. Q. (2014). A perspective on anti-1933 CCN2 therapy for chronic kidney disease. Nephrol. Dial. Transpl. 29(suppl 1), 1934 i30-i37. doi: 10.1093/ndt/gft430
- Falke, L. L., Leeuwis, J. W., Lyons, K. M., Mummery, C. L., Nguyen, T. Q., 1935 and Goldschmeding, R. (2017). CCN2 reduction mediates protective effects of 1936 BMP7 treatment in obstructive nephropathy. J. Cell Commun. Signal. 11, 39-48. 1937 doi: 10.1007/s12079-016-0358-2 1938

2000

2001

2002

2003

2021

2022

2023

2045

2046

- Fallon, J. T., DavidGertz, S., Chesebro, J. H., and Badimon, J. J. (1997). Tissue factor
  modulates the thrombogenicity of human atherosclerotic plaques. *Circulation*95, 594–599. doi: 10.1161/01.CIR.95.3.594
- Feig, J. E., Parathath, S., Rong, J. X., Mick, S. L., Vengrenyuk, Y., Grauer, L.,
  et al. (2011). Reversal of hyperlipidemia with a genetic switch favorably affects
  the content and inflammatory state of macrophages in atherosclerotic plaques.
- 1944 Circulation 123, 989–998. doi: 10.1161/CIRCULATIONAHA.110.984146
- Feig, J. E., Quick, J. S., and Fisher, E. A. (2009). The role of a murine transplantation model of atherosclerosis regression in drug discovery. *Curr. Opin. Investig. Drugs* 10, 232–238.
- Ferrante, C. J., and Leibovich, S. J. (2012). Regulation of macrophage polarization
  and wound healing. *Adv. Wound Care* 1, 10–16. doi: 10.1089/wound.2011.0307
- Fierro, I. M., and Serhan, C. N. (2001). Mechanisms in anti-inflammation and resolution: the role of lipoxins and aspirin-triggered lipoxins. *Braz. J. Med. Biol. Res.* 34, 555–566. doi: 10.1590/S0100-879X2001000500002
- Fisher, E. A. (2016). Regression of atherosclerosis: the journey from the liver to the plaque and back. *Arterioscler. Thromb. Vasc. Biol.* 36, 226–235. doi: 10.1161/ ATVBAHA.115.301926
- Fleg, J. L., Mete, M., Howard, B. V., Umans, J. G., Roman, M. J., Ratner, R. E.,
  et al. (2008). Effect of statins alone versus statins plus ezetimibe on carotid atherosclerosis in type 2 diabetes: the SANDS (stop atherosclerosis in native diabetics study) trial. J. Am. Coll. Cardiol. 52, 2198–2205. doi: 10.1016/j.jacc.
  2008.10.031
- Forbes, J. M., and Cooper, M. E. (2013). Mechanisms of diabetic complications.
   *Physiol. Rev.* 93, 137–188. doi: 10.1152/physrev.00045.2011
- Franceschi, C., Garagnani, P., Vitale, G., Capri, M., and Salvioli, S. (2017). Inflammaging and 'Garb-aging'. *Trends Endocrinol. Metab.* 28, 199–212. doi: 10.1016/j.tem.2016.09.005
- Fredman, G., Kamaly, N., Spolitu, S., Milton, J., Ghorpade, D., Chiasson, R.,
  et al. (2015). Targeted nanoparticles containing the proresolving peptide Ac226 protect against advanced atherosclerosis in hypercholesterolemic mice. *Sci. Transl. Med.* 7:275ra220. doi: 10.1126/scitranslmed.aaa1065
- Fredman, G., and Tabas, I. (2017). Boosting inflammation resolution in atherosclerosis: the next frontier for therapy. *Am. J. Pathol.* 187, 1211–1221. doi:10.1016/j.ajpath.2017.01.018
- Fullerton, J. N., and Gilroy, D. W. (2016). Resolution of inflammation:
  a new therapeutic frontier. *Nat. Rev. Drug Discov.* 15, 551–567.
  doi: 10.1038/nrd.2016.39
- 1970 Funaki, M. (2009). Saturated fatty acids and insulin resistance. J. Med. Invest. 56,
   1971 88–92. doi: 10.2152/jmi.56.88
- 1972 Fuster, V., Badimon, J. J., and Chesebro, J. H. (1998). Atherothrombosis:
   1973 mechanisms and clinical therapeutic approaches. *Vasc. Med.* 3, 231–239.
   1974 doi: 10.1177/1358836X9800300310
- Fuster, V., and Kelly, B. (2011). Summary of the institute of medicine report promoting cardiovascular health in the developing world. *Glob. Heart* 6, 133–142. doi: 10.1016/j.gheart.2011.08.002
- Geissmann, F., Auffray, C., Palframan, R., Wirrig, C., Ciocca, A., Campisi, L., et al.
  (2008). Blood monocytes: distinct subsets, how they relate to dendritic cells, and
  their possible roles in the regulation of T-cell responses. *Immunol. Cell Biol.* 86, 398–408. doi: 10.1038/icb.2008.19
- Godson, C., and Brady, H. R. (2000). Lipoxins: novel anti-inflammatory
   therapeutics? Curr. Opin. Invest. Drugs 1, 380–385.
- Godson, C., Mitchell, S., Harvey, K., Petasis, N. A., Hogg, N., and Brady, H. R.
  (2000). Cutting edge: lipoxins rapidly stimulate nonphlogistic phagocytosis of apoptotic neutrophils by monocyte-derived macrophages. *J. Immunol.* 164, 1663–1667. doi: 10.4049/jimmunol.164.4.1663
- Goicoechea, M., Sanchez-Nino, M. D., Ortiz, A., Garcia de Vinuesa, S., Quiroga, B.,
  Bernis, C., et al. (2017). Low dose aspirin increases 15-epi-lipoxin A4 levels in
  diabetic chronic kidney disease patients. *Prostaglandins Leukot. Essent. Fatty*Acids 125, 8–13. doi: 10.1016/j.plefa.2017.08.009
- Goraya, T. Y., Leibson, C. L., Palumbo, P. J., Weston, S. A., Killian, J. M., Pfeifer,
   E. A., et al. (2002). Coronary atherosclerosis in diabetes mellitus. A population-
- <sup>1990</sup>based autopsy study. J. Am. Coll. Cardiol. 40, 946–953. doi: 10.1016/S0735-1097(02)02065-X
- Greenlee-Wacker, M. C. (2016). Clearance of apoptotic neutrophils and resolution
   of inflammation. *Immunol. Rev.* 273, 357–370. doi: 10.1111/imr.12453
- Gregg, E. W., Cheng, Y. J., Srinivasan, M., Lin, J., Geiss, L. S., Albright, A. L.,
   et al. (2018). Trends in cause-specific mortality among adults with and without

diagnosed diabetes in the USA: an epidemiological analysis of linked national survey and vital statistics data. *Lancet* 391, 2430–2440. doi: 10.1016/S0140-6736(18)30314-3

- Gregg, E. W., Li, Y., Wang, J., Rios Burrows, N., Ali, M. K., Rolka, D., et al. (2014). Changes in diabetes-related complications in the United States, 1990–2010. *New Engl. J. Med.* 370, 1514–1523. doi: 10.1056/NEJMoa1310799
- Gresele, P., Guglielmini, G., De Angelis, M., Ciferri, S., Ciofetta, M., Falcinelli, E., et al. (2003). Acute, short-term hyperglycemia enhances shear stress-induced platelet activation in patients with type II diabetes mellitus. *J. Am. Coll. Cardiol.* 41, 1013–1020. doi: 10.1016/S0735-1097(02)02972-8

Gronert, K. (2005). Lipoxins in the eye and their role in wound healing. *Prostaglandins Leukot. Essent. Fatty Acids* 73, 221–229. doi: 10.1016/j.plefa. 2005.05.009

- Guilford, W. J., and Parkinson, J. F. (2005). Second-generation beta-oxidation resistant 3-oxa-lipoxin A4 analogs. *Prostaglandins Leukot. Essent. Fatty Acids* 73, 245–250. doi: 10.1016/j.plefa.2005.05.012
- Guzik, T. J., and Cosentino, F. (2018). Epigenetics and immunometabolism in diabetes and aging. Antioxid. Redox Signal. 29, 257–274. doi: 10.1089/ars.2017.
   2010
   7299
- Haritoglou, C., Kook, D., Neubauer, A., Wolf, A., Priglinger, S., Strauss, R., et al. (2006). Intravitreal bevacizumab (Avastin) therapy for persistent diffuse diabetic macular edema. *Retina* 26, 999–1005. doi: 10.1097/01.iae.0000247165. 38655.bf
- Harjutsalo, V., and Groop, P. H. (2014). Epidemiology and risk factors for diabetic kidney disease. Adv. Chronic Kidney Dis. 21, 260–266. doi: 10.1053/j.ackd.2014.
   03.009
- b).009
  c).009
  c).0187
  c).009
  <lic).009</li>
  c).009
  c).009
  c).00
- Hellmann, J., Sansbury, B. E., Wong, B., Li, X., Singh, M., Nuutila, K., et al. (2018).
  Biosynthesis of D-series resolvins in skin provides insights into their role in tissue repair. *J. Invest. Dermatol.* 138, 2051–2060. doi: 10.1016/j.jid.2018.03.
  1498
- Hellmann, J., Tang, Y., Kosuri, M., Bhatnagar, A., and Spite, M. (2011). Resolvin D1 decreases adipose tissue macrophage accumulation and improves insulin sensitivity in obese-diabetic mice. *FASEB J.* 25, 2399–2407. doi: 10.1096/fj.10-178657
- Herrington, W., Lacey, B., Sherliker, P., Armitage, J., and Lewington, S. (2016).
   2027

   Epidemiology of atherosclerosis and the potential to reduce the global burden of atherothrombotic disease. *Circ. Res.* 118, 535–546. doi: 10.1161/CIRCRESAHA.
   2029

   115.307611
   2030
- Hewing, B., Parathath, S., Barrett, T., Chung, W. K., Astudillo, Y. M., Hamada, T., et al. (2014). Effects of native and myeloperoxidase-modified apolipoprotein a-I on reverse cholesterol transport and atherosclerosis in mice. *Arterioscler. Thromb. Vasc. Biol.* 34, 779–789. doi: 10.1161/ATVBAHA.113.30
  2031 2032 2033 2034
- Ho, K. J., Spite, M., Owens, C. D., Lancero, H., Kroemer, A. H., Pande, R., et al. (2010). Aspirin-triggered lipoxin and resolvin E1 modulate vascular smooth muscle phenotype and correlate with peripheral atherosclerosis. *Am. J. Pathol.* 177, 2116–2123. doi: 10.2353/ajpath.2010.091082
- Hodges, R. R., Li, D., Shatos, M. A., Bair, J. A., Lippestad, M., Serhan, C. N., et al.
   2038

   (2017). Lipoxin A4 activates ALX/FPR2 receptor to regulate conjunctival gobbe
   2039

   cell secretion. Mucosal Immunol. 10, 46–57. doi: 10.1038/mi.2016.33
   2040
- Hong, S., Gronert, K., Devchand, P. R., Moussignac, R. L., and Serhan, C. N. (2003). Novel docosatrienes and 17S-resolvins generated from docosahexaenoic acid in murine brain, human blood, and glial cells. Autacoids in anti-inflammation. J. Biol. Chem. 278, 14677–14687. doi: 10.1074/jbc.
   M300218200
- Hong, S., Porter, T. F., Lu, Y., Oh, S. F., Pillai, P. S., and Serhan, C. N. (2008). Resolvin E1 metabolome in local inactivation during inflammation-resolution. *J. Immunol.* 180, 3512–3519. doi: 10.4049/jimmunol.180.5.3512
- Huang, H., Gandhi, J. K., Zhong, X., Wei, Y., Gong, J., Duh, E. J., et al. (2011).2047TNFalpha is required for late BRB breakdown in diabetic retinopathy, and its<br/>inhibition prevents leukostasis and protects vessels and neurons from apoptosis.2048Invest. Ophthalmol. Vis. Sci. 52, 1336–1344. doi: 10.1167/iovs.10-57682050
- Huh, J. H., Kim, H. M., Lee, E. S., Kwon, M. H., Lee, B. R., Ko, H. J., et al. (2018). Dual CCR2/5 antagonist attenuates obesity-induced insulin resistance
   2051 2052

2053	by regulating macrophage recruitment and M1/M2 status. Obesity 26, 378–386.
2054	doi: 10.1002/oby.22103

- Hussein, M. A., Shrestha, E., Ouimet, M., Barrett, T. J., Leone, S., Moore, K. J., et al. (2015). LXR-mediated ABCA1 expression and function are modulated by high glucose and PRMT2. *PLoS One* 10:e0135218. doi: 10.1371/journal.pone.
   0135218
- Ivanov, S., Scallan, J. P., Kim, K. W., Werth, K., Johnson, M. W., Saunders, B. T., et al. (2016). CCR7 and IRF4-dependent dendritic cells regulate lymphatic collecting vessel permeability. *J. Clin. Invest.* 126, 1581–1591. doi: 10.1172/ JCI84518
- Jing, A. M., Tankeu, A. T., Ateba, N. A., and Noubiap, J. J. (2017). Mechanism
  of worsening diabetic retinopathy with rapid lowering of blood glucose: the
  synergistic hypothesis. *BMC Endocrine Disord*. 17:63. doi: 10.1186/s12902-0170213-3
- Kahn, S. E., Hull, R. L., and Utzschneider, K. M. (2006). Mechanisms linking obesity to insulin resistance and type 2 diabetes. *Nature* 444):840.
- Kalluri, R., and Weinberg, R. A. (2009). The basics of epithelial mesenchymal transition. J. Clin. Invest. 119, 1420–1428. doi: 10.1172/JC
   I39104
- Kantharidis, P., Hagiwara, S., Brennan, E., and McClelland, A. D. (2015). Study of microRNA in diabetic nephropathy: isolation, quantification and biological function. *Nephrology* 20, 132–139. doi: 10.1111/nep.12374
- Kay, A. M., Simpson, C. L., and Stewart, J. A. (2016). The role of AGE/RAGE signaling in diabetes-mediated vascular calcification. *J. Diabetes Res.* 2016:6809703. doi: 10.1155/2016/6809703
- Kieran, N. E., Doran, P. P., Connolly, S. B., Greenan, M. C., Higgins, D. F., Leonard, M., et al. (2003). Modification of the transcriptomic response to renal ischemia/reperfusion injury by lipoxin analog. *Kidney Int.* 64, 480–492. doi: 10.1046/j.1523-1755.2003.00106.x
- Klaassen, I., Van Noorden, C. J., and Schlingemann, R. O. (2013). Molecular basis
   of the inner blood-retinal barrier and its breakdown in diabetic macular edema
   and other pathological conditions. *Prog. Retin. Eye Res.* 34, 19–48. doi: 10.1016/
   j.preteyeres.2013.02.001
- Kowluru, R. A., Zhong, Q., and Santos, J. M. (2012). Matrix metalloproteinases in diabetic retinopathy: potential role of MMP-9. *Expert. Opin. Investig. Drugs* 21, 797–805. doi: 10.1517/13543784.2012.681043
- Krishnamoorthy, S., Recchiuti, A., Chiang, N., Yacoubian, S., Lee, C.-H.,
  Yang, R., et al. (2010). Resolvin D1 binds human phagocytes with
  evidence for proresolving receptors. *Proc. Natl. Acad. Sci. U.S.A.* 107,
  1660–1665.
- Krolewski, A. S., Niewczas, M. A., Skupien, J., Gohda, T., Smiles, A., Eckfeldt,
   J. H., et al. (2014). Early progressive renal decline precedes the onset of
   microalbuminuria and its progression to macroalbuminuria. *Diabetes Care* 37,
   226–234. doi: 10.2337/dcl3-0985
- Kunjathoor, V. V., Febbraio, M., Podrez, E. A., Moore, K. J., Andersson, L.,
   Koehn, S., et al. (2002). Scavenger receptors class A-I/II and CD36 are
   the principal receptors responsible for the uptake of modified low density
   lipoprotein leading to lipid loading in macrophages. J. Biol. Chem. 277,
   49982–49988. doi: 10.1074/jbc.M209649200
- Lassila, M., Allen, T. J., Cao, Z., Thallas, V., Jandeleit-Dahm, K. A.,
   Candido, R., et al. (2004). Imatinib attenuates diabetes-associated
   atherosclerosis. Arterioscler. Thromb. Vasc. Biol. 24, 935–942.
   doi: 10.1161/01.ATV.0000124105.39900.db
- Leaf, I. A., and Duffield, J. S. (2017). What can target kidney fibrosis? Nephrol. Dial.
   Transplant. 32(suppl 1), i89–i97. doi: 10.1093/ndt/gfw388
- Lee, I. M., Shiroma, E. J., Lobelo, F., Puska, P., Blair, S. N., Katzmarzyk, P. T.,
  et al. (2012). Effect of physical inactivity on major non-communicable diseases
  worldwide: an analysis of burden of disease and life expectancy. *Lancet* 380,
  210–229. doi: 10.1016/S0140-6736(12)61031-9
- Lee, J., Rhee, M. H., Kim, E., and Cho, J. Y. (2012). BAY 11-7082 is a broad-spectrum inhibitor with anti-inflammatory activity against multiple targets. *Mediat. Inflamm.* 2012:416036. doi: 10.1155/2012/416036
- Lee, J. E., Sun, Y., Gjorstrup, P., and Pearlman, E. (2015). Inhibition of corneal inflammation by the resolvin E1. *Invest. Ophthalmol. Vis. Sci.* 56, 2728–2736. doi: 10.1167/iovs.14-15982
- Lee, S., Huen, S., Nishio, H., Nishio, S., Lee, H. K., Choi, B.-S., et al. (2011). Distinct macrophage phenotypes contribute to kidney injury and repair. *J. Am. Soc. Nephrol.* 22, 317–326. doi: 10.1681/ASN.2009060615

- Leonard, M. O., Hannan, K., Burne, M. J., Lappin, D. W., Doran, P., Coleman, P., et al. (2002). 15-Epi-16-(para-fluorophenoxy)-lipoxin A(4)-methyl ester, a synthetic analogue of 15-epi-lipoxin A(4), is protective in experimental ischemic acute renal failure. J. Am. Soc. Nephrol. 13, 1657–1662. doi: 10.1097/ 01.ASN.0000015795.74094.91 2112
- Levy, B. D., Clish, C. B., Schmidt, B., Gronert, K., and Serhan, C. N. (2001). Lipid mediator class switching during acute inflammation: signals in resolution. *Nat. Immunol.* 2, 612–619. doi: 10.1038/89759
- Levy, B. D., Zhang, Q. Y., Bonnans, C., Primo, V., Reilly, J. J., Perkins, D. L., et al. (2011). The endogenous pro-resolving mediators lipoxin A4 and resolvin E1 preserve organ function in allograft rejection. *Prostaglandins Leukot Essent Fatty Acids* 84, 43–50. doi: 10.1016/j.plefa.2010.09.002
- Li, A. C., and Glass, C. K. (2002). The macrophage foam cell as a target for therapeutic intervention. *Nat. Med.* 8, 1235–1242. doi: 10.1038/nm1102-1235 2121
- Li, D., Hodges, R. R., Jiao, J., Carozza, R. B., Shatos, M. A., Chiang, N., et al. (2013). Resolvin D1 and aspirin-triggered resolvin D1 regulate histaminestimulated conjunctival goblet cell secretion. *Mucosal Immunol.* 6, 1119–1130. doi: 10.1038/mi.2013.7
- Li, S., Sun, Y., Liang, C.-P., Thorp, E. B., Han, S., Jehle, A. W., et al. (2009). Defective phagocytosis of apoptotic cells by macrophages in atherosclerotic lesions of ob/ob mice and reversal by a fish oil diet. *Circ. Res.* 105:1072. doi: 10.1161/CIRCRESAHA.109.199570
- Libby, P., Ridker, P. M., and Maseri, A. (2002). Inflammation and atherosclerosis. 2128 *Circulation* 105, 1135–1143. doi: 10.1161/hc0902.104353 2129
- Lieth, E., Gardner, T. W., Barber, A. J., Antonetti, D. A., et al. (2000). Retinal neurodegeneration: early pathology in diabetes. *Clin. Exp. Ophthalmol.* 28, 3–8. doi: 10.1046/j.1442-9071.2000.00222.x
- Liu, Y. (2011). Cellular and molecular mechanisms of renal fibrosis. *Nat. Rev.* 2132 Nephrol. 7:684. doi: 10.1038/nrneph.2011.149
- Ljubimov, A. V., Burgeson, R. E., Butkowski, R. J., Couchman, J. R., Wu, R. R., Ninomiya, Y., et al. (1996). Extracellular matrix alterations in human corneas with bullous keratopathy. *Invest. Ophthalmol. Vis. Sci.* 37, 997–1007.
   2136
- Llodra, J., Angeli, V., Liu, J., Trogan, E., Fisher, E. A., and Randolph, G. J. (2004).
   Emigration of monocyte-derived cells from atherosclerotic lesions characterizes regressive, but not progressive, plaques. *Proc. Natl. Acad. Sci. U.S.A.* 101, 11779–11784. doi: 10.1073/pnas.0403259101
- Lotfy, M., Adeghate, J., Kalasz, H., Singh, J., and Adeghate, E. (2017). Chronic complications of diabetes mellitus: a mini review. *Curr. Diabetes Rev.* 13, 3–10. doi: 10.2174/1573399812666151016101622

Lusis, A. J. (2000). Atherosclerosis. Nature 407, 233-241. doi: 10.1038/35025203 2142

- Ma, J., Yue, J., Huang, R., Liao, Y., Li, S., and Liu, W. (2018). Reversion of aging-related DHEAS decline in mouse plasma alleviates aging-related glucose tolerance impairment by potentiation of glucose-stimulated insulin secretion of acute phase. *Biochem. Biophys. Res. Commun.* 500, 671–675. doi: 10.1016/j.bbrc. 2018.04.134
- Maderna, P., Cottell, D. C., Toivonen, T., Dufton, N., Dalli, J., Perretti, M., et al. (2010). FPR2/ALX receptor expression and internalization are critical for lipoxin A(4) and annexin-derived peptide-stimulated phagocytosis. *FASEB J.* 24, 4240–4249. doi: 10.1096/fj.10-159913
- Maderna, P., and Godson, C. (2009). Lipoxins: resolutionary road. Br. J. Pharmacol. 158, 947–959. doi: 10.1111/j.1476-5381.2009.00386.x 2151
- Mann, J. F. E., Orsted, D. D., Brown-Frandsen, K., Marso, S. P., Poulter, N. R.,<br/>Rasmussen, S., et al. (2017). Liraglutide and renal outcomes in type 2 diabetes.<br/>N. Engl. J. Med. 377, 839–848. doi: 10.1056/NEJMoa16160112152<br/>2153
- Mantovani, A., Sica, A., Sozzani, S., Allavena, P., Vecchi, A., and Locati, M. (2004). The chemokine system in diverse forms of macrophage activation and polarization. *Trends Immunol.* 25, 677–686. doi: 10.1016/j.it.2004.09.015
- Marcheselli, V. L., Hong, S., Lukiw, W. J., Tian, X. H., Gronert, K., Musto, A., et al. (2003). Novel docosanoids inhibit brain ischemia-reperfusion-mediated leukocyte infiltration and pro-inflammatory gene expression. *J. Biol. Chem.* 278, 43807–43817. doi: 10.1074/jbc.M305841200
   2160
- Martinez, F. O., and Gordon, S. (2014). The M1 and M2 paradigm of macrophage activation: time for reassessment. *F1000prime Rep.* 6, 13–13. doi: 10.12703/P6-13 2162
- Mas, E., Barden, A., Burke, V., Beilin, L. J., Watts, G. F., Huang, R. C., et al. (2016). A randomized controlled trial of the effects of n-3 fatty acids on resolvins in chronic kidney disease. *Clin. Nutr.* 35, 331–336. doi: 10.1016/j. clnu.2015.04.004

2228

2253

2254

2262

- 2167 McGill, H. C. Jr., McMahan, C. A., Malcom, G. T., Oalmann, M. C., and Strong, J. P. (1995). Relation of glycohemoglobin and adiposity to 2168 atherosclerosis in youth. Pathobiological Determinants of Atherosclerosis in 2169 Youth (PDAY) research group. Arterioscler Thromb Vasc. Biol. 15, 431-440. 2170 doi: 10.1161/01.ATV.15.4.431
- 2171 McMahon, B., Mitchell, D., Shattock, R., Martin, F., Brady, H. R., and Godson, C. (2002). Lipoxin, leukotriene, and PDGF receptors cross-talk to regulate 2172 mesangial cell proliferation. FASEB J. 16, 1817-1819. doi: 10.1096/fj.02-0416fje 2173
- Meininger, C. J., Marinos, R. S., Hatakevama, K., Martinez-Zaguilan, R., Rojas, 2174 J. D., Kelly, K. A., et al. (2000). Impaired nitric oxide production in 2175 coronary endothelial cells of the spontaneously diabetic BB rat is due to
- 2176 tetrahydrobiopterin deficiency. Biochem. J. 349(Pt 1): 353-356. doi: 10.1042/ bi3490353 2177
- Merched, A. J., Ko, K., Gotlinger, K. H., Serhan, C. N., and Chan, L. 2178 (2008). Atherosclerosis: evidence for impairment of resolution of vascular 2179 inflammation governed by specific lipid mediators. FASEB J. 22, 3595-3606. 2180 doi: 10.1096/fj.08-112201
- 2181 Merched, A. J., Ko, K., Gotlinger, K. H., Serhan, C. N., and Chan, L. 2182 (2008). Atherosclerosis: evidence for impairment of resolution of vascular inflammation governed by specific lipid mediators. FASEB J. 22, 3595-3606. 2183 doi: 10.1096/fj.08-112201
- 2184 Mitchell, D., Rodgers, K., Hanly, J., McMahon, B., Brady, H. R., Martin, F., 2185 et al. (2004). Lipoxins inhibit Akt/PKB activation and cell cycle progression in human mesangial cells. Am. J. Pathol. 164, 937-946. doi: 10.1016/S0002-2186 9440(10)63181-1 2187
- Mitchell, S., Thomas, G., Harvey, K., Cottell, D., Reville, K., Berlasconi, G., et al. 2188 (2002). Lipoxins, aspirin-triggered epi-lipoxins, lipoxin stable analogues, and 2189 the resolution of inflammation: stimulation of macrophage phagocytosis of 2190 apoptotic neutrophils in vivo. J. Am. Soc. Nephrol. 13, 2497-2507. doi: 10.1097/ 2191 01.ASN.0000032417.73640.72
- Moore, K. J., Sheedy, F. J., and Fisher, E. A. (2013). Macrophages in atherosclerosis: 2192 a dynamic balance. Nat. Rev. Immunol. 13, 709-721. doi: 10.1038/nri3520
- 2193 Moore, K. J., and Tabas, I. (2011). Macrophages in the pathogenesis of 2194 atherosclerosis. Cell 145, 341-355. doi: 10.1016/j.cell.2011.04.005
- 2195 Moore, T. C., Moore, J. E., Kaji, Y., Frizzell, N., Usui, T., Poulaki, V., et al. 2196 (2003). The role of advanced glycation end products in retinal microvascular leukostasis. Invest. Ophthalmol. Vis. Sci. 44, 4457-4464. doi: 10.1167/iovs.02-2197 1063 2198
- Mosser, D. M., and Edwards, J. P. (2008). Exploring the full spectrum of 2199 macrophage activation. Nat. Rev. Immunol. 8, 958-969. doi: 10.1038/nri2448
- 2200 Motwani, M. P., Flint, J. D., De Maeyer, R. P. H., Fullerton, J. N., Smith, A. M., Marks, D. J. B., et al. (2016). Novel translational model 2201 of resolving inflammation triggered by UV-killed E. coli. J. Pathol. 2, 2202 154-165.
- 2203 Murphy, M., Godson, C., Cannon, S., Kato, S., Mackenzie, H. S., Martin, F., et al. 2204 (1999). Suppression subtractive hybridization identifies high glucose levels as a 2205 stimulus for expression of connective tissue growth factor and other genes in human mesangial cells. J. Biol. Chem. 274, 5830-5834. doi: 10.1074/jbc.274.9. 2206 5830 2207
- Nagareddy, P. R., Murphy, A. J., Stirzaker, R. A., Hu, Y., Yu, S., Miller, 2208 R. G., et al. (2013). Hyperglycemia promotes myelopoiesis and impairs the 2209 resolution of atherosclerosis. Cell Metab. 17, 695-708. doi: 10.1016/j.cmet.2013. 04 001 2210
- Nagenborg, J., Goossens, P., Biessen, E. A. L., and Donners, M. M. P. C. (2017). 2211 Heterogeneity of atherosclerotic plaque macrophage origin, phenotype and 2212 functions: implications for treatment. Eur. J. Pharmacol. 816, 14-24. doi: 10. 2213 1016/i.eiphar.2017.10.005
- 2214 National Kidney Foundation (2002). K/DOQI clinical practice guidelines for 2215 chronic kidney disease: evaluation, classification, and stratification. Am. J. Kidnev Dis. 39, S1-S266. 2216
- Nathan, C., and Ding, A. (2010). Nonresolving inflammation. Cell 140, 871-882. 2217 doi: 10.1016/j.cell.2010.02.029
- 2218 O'Sullivan, T. P., Vallin, K. S., Shah, S. T., Fakhry, J., Maderna, P., Scannell, M., et al. 2219 (2007). Aromatic lipoxin A4 and lipoxin B4 analogues display potent biological activities. J. Med. Chem. 50, 5894-5902. doi: 10.1021/jm060270d 2220
- Orban, J.-C., Van Obberghen, E., and Ichai, C. (2018). Acute Complications 2221 of Diabetes. Metabolic Disorders and Critically Ill Patients, Berlin: Springer, 2222 341-363. doi: 10.1007/978-3-319-64010-5\_15 2223

- Paneni, F., Beckman, J. A., Creager, M. A., and Cosentino, F. (2013). Diabetes and 2224 vascular disease: pathophysiology, clinical consequences, and medical therapy: 2225 part I. Eur. Heart J. 34, 2436-2443. doi: 10.1093/eurhearti/eht149 2226
- Papatheodorou, K., Papanas, N., Banach, M., Papazoglou, D., and Edmonds, M. (2016). Complications of diabetes 2016. J. Diabetes Res. 2016:6989453. doi: 10.1155/2016/6989453
- Parkinson, J. F. (2006). Lipoxin and synthetic lipoxin analogs: an overview 2229 of anti-inflammatory functions and new concepts in immunomodulation. 2230 Inflamm. Allergy Drug Targets 5, 91-106. doi: 10.2174/18715280 2231 6776383125
- 2232 Pepine, C. J. (1998). The effects of angiotensin-converting enzyme inhibition on endothelial dysfunction: potential role in myocardial ischemia. Am. J. Cardiol. 2233 82, 23S-27S. doi: 10.1016/S0002-9149(98)90427-1 2234
- Pergola, P. E., Krauth, M., Huff, J. W., Ferguson, D. A., Ruiz, S., Meyer, C. J., et al. 2235 (2011). Effect of bardoxolone methyl on kidney function in patients with T2D 2236 and stage 3b-4 CKD. Am. J. Nephrol. 33, 469-476. doi: 10.1159/000327599
- 2237 Petasis, N. A., Keledjian, R., Sun, Y. P., Nagulapalli, K. C., Tjonahen, E., Yang, R., 2238 et al. (2008). Design and synthesis of benzo-lipoxin A4 analogs with enhanced stability and potent anti-inflammatory properties. Bioorg. Med. Chem. Lett. 18, 2239 1382-1387. doi: 10.1016/j.bmcl.2008.01.013
- 2240 Petri, M. H., Laguna-Fernandez, A., Tseng, C. N., Hedin, U., Perretti, M., 2241 and Back, M. (2015). Aspirin-triggered 15-epi-lipoxin A(4) signals through 2242 FPR2/ALX in vascular smooth muscle cells and protects against intimal hyperplasia after carotid ligation. Int. J. Cardiol. 179, 370-372. doi: 10.1016/j. 2243 ijcard.2014.11.010 2244
- Piccoli, G. B., Grassi, G., Cabiddu, G., Nazha, M., Roggero, S., Capizzi, I., et al. 2245 (2015). Diabetic kidney disease: a syndrome rather than a single disease. Rev. 2246 Diabetic Stud. 12, 87-109. doi: 10.1900/RDS.2015.12.87
- Pierdomenico, A. M., Recchiuti, A., Simiele, F., Codagnone, M., Mari, V. C., Davì, 2247 G., et al. (2015). MicroRNA-181b regulates ALX/FPR2 receptor expression and 2248 proresolution signaling in human macrophages. J. Biol. Chem. 290, 3592-3600. 2249 doi: 10.1074/ibc.M114.592352 2250
- Potteaux, S., Gautier, E. L., Hutchison, S. B., van Rooijen, N., Rader, D. J., 2251 Thomas, M. I., et al. (2011). Suppressed monocyte recruitment drives macrophage removal from atherosclerotic plaques of Apoe-/- mice during 2252 disease regression. J. Clin. Invest. 121, 2025-2036. doi: 10.1172/JCI43802
- Puro, D. G. (2002). Diabetes-induced dysfunction of retinal Muller cells. Trans. Am. Ophthalmol. Soc. 100, 339-352.
- 2255 Qu, X., Zhang, X., Yao, J., Song, J., Nikolic-Paterson, D. J., and Li, J. (2012). 2256 Resolvins E1 and D1 inhibit interstitial fibrosis in the obstructed kidney via 2257 inhibition of local fibroblast proliferation. J. Pathol. 228, 506-519. doi: 10.1002/ path.4050 2258
- Rabbani, N., and Thornalley, P. J. (2018). Advanced glycation end products in the 2259 pathogenesis of chronic kidney disease. Kidney Int. 93, 803-813. doi: 10.1016/j. 2260 kint.2017.11.034 2261
- Randolph, G. J. (2008). Emigration of monocyte-derived cells to lymph nodes during resolution of inflammation and its failure in atherosclerosis. Curr. Opin. Lipidol. 19, 462-468. doi: 10.1097/MOL.0b013e32830d5f09
- 2263 Rashid, S., and Genest, J. (2007). Effect of obesity on high-density lipoprotein 2264 metabolism. Obesity 15, 2875-2888. doi: 10.1038/oby.2007.342
- 2265 Rayner, K. J., Sheedy, F.J., Esau, C. C., Hussain, F. N., Temel, R. E., Parathath, S., et al. (2011). Antagonism of miR-33 in mice promotes reverse cholesterol 2266 transport and regression of atherosclerosis. J. Clin. Invest. 121, 2921-2931. 2267 doi: 10.1172/JCI57275 2268
- Reis, E. D., Li, J., Fayad, Z. A., Rong, J. X., Hansoty, D., Aguinaldo, J.-G., 2269 et al. (2001). Dramatic remodeling of advanced atherosclerotic plaques of the 2270 apolipoprotein E-deficient mouse in a novel transplantation model. J. Vasc. 2271 Surg. 34, 541A-542A. doi: 10.1067/mva.2001.115963 2272
- Ridker, P. M., Everett, B. M., Thuren, T., MacFadyen, J. G., Chang, W. H., Ballantyne, C., et al. (2017). Antiinflammatory therapy with canakinumab for atherosclerotic disease. N. Engl. J. Med. 377, 1119-1131. doi: 10.1056/ NEJMoa1707914
- 2275 Ridker, P. M., MacFadyen, J. G., Glynn, R. J., Koenig, W., Libby, P., Everett, B. M., 2276 et al. (2018). Inhibition of interleukin-1β by canakinumab and cardiovascular outcomes in patients with chronic kidney disease. J. Am. Coll. Cardiol. 71, 2277 2405-2414. doi: 10.1016/j.jacc.2018.03.490 2278
- Ridker, P. M., Thuren, T., Zalewski, A., and Libby, P. (2011). Interleukin-1β inhibition and the prevention of recurrent cardiovascular events: rationale and

2273

- design of the Canakinumab Anti-inflammatory Thrombosis Outcomes Study 2281 (CANTOS). Am. Heart. J. 162, 597-605. doi: 10.1016/j.ahj.2011.06.012 2282
- Rodgers, K. McMahon, B. Mitchell, D. Sadlier, D. and Godson, C. (2005) 2283 Lipoxin A4 modifies platelet-derived growth factor-induced pro-fibrotic gene 2284 expression in human renal mesangial cells. Am. J. Pathol. 167, 683-694. 2285 doi: 10.1016/S0002-9440(10)62043-3
- Rong, J. X., Li, J., Reis, E. D., Choudhury, R. P., Dansky, H. M., Elmalem, V. I., 2286 et al. (2001). Elevating high-density lipoprotein cholesterol in apolipoprotein 2287 E-deficient mice remodels advanced atherosclerotic lesions by decreasing 2288 macrophage and increasing smooth muscle cell content. Circulation 104, 2289 2447-2452, doi: 10.1161/hc4501.098952
- 2290 Ross, R. (1999). Atherosclerosis-an inflammatory disease. N. Engl. J. Med. 340, 115-126. doi: 10.1056/NEJM199901143400207 2291
- Rossi, S., D'Amico, M., Capuano, A., Romano, M., Petronella, P., and Di Filippo, C. 2292 (2006). Hyperglycemia in streptozotocin-induced diabetes leads to persistent 2293 inflammation and tissue damage following uveitis due to reduced levels of 2294 ciliary body heme oxygenase-1. Mediat. Inflamm. 2006:60285. doi: 10.1155/MI/ 2295 2006/60285
- Rossing, P. (2018). Subclassification of diabetes based on quantitative traits. Nat. 2296 Rev. Nephrol. 14, 355-356. doi: 10.1038/s41581-018-0011-9 2297
- Roy, S., Cagliero, E., and Lorenzi, M. (1996). Fibronectin overexpression in retinal 2298 microvessels of patients with diabetes. Invest. Ophthalmol. Vis. Sci. 37, 258-266. 2299
- Roy, S., Maiello, M., and Lorenzi, M. (1994). Increased expression of basement membrane collagen in human diabetic retinopathy. J. Clin. Invest. 93, 438-442. 2300 doi: 10.1172/ICI116979 2301
- Rubsam, A., Parikh, S., and Fort, P. E. (2018). Role of Inflammation in Diabetic 2302 Retinopathy. Int. J. Mol. Sci. 19: E942. doi: 10.3390/ijms19040942
- 2303 Russell, J. W., and Zilliox, L. A. (2014). Diabetic neuropathies. Continuum 20, 2304 1226-1240. doi: 10.1212/01.CON.0000455884.29545.d2
- Sandholm, N., Forsblom, C., Mäkinen, V.-P., McKnight, A. J., Österholm, A.-2305 M., He, B., et al. (2014). Genome-wide association study of urinary albumin 2306 excretion rate in patients with type 1 diabetes. Diabetologia 57, 1143-1153. 2307 doi: 10.1007/s00125-014-3202-3
- 2308 Sato, T., Kusaka, S., Shimojo, H., and Fujikado, T. (2009). Simultaneous analyses 2309 of vitreous levels of 27 cytokines in eyes with retinopathy of prematurity. 2310 Ophthalmology 116, 2165-2169. doi: 10.1016/j.ophtha.2009.04.026
- Schroder, K., Zhou, R., and Tschopp, J. (2010). The NLRP3 Inflammasome: a sensor 2311 for metabolic danger? Science 327, 296-300. doi: 10.1126/science.1184003
- 2312 Scrivo, R., Vasile, M., Bartosiewicz, I., and Valesini, G. (2011). Inflammation as 2313 common soil of the multifactorial diseases. Autoimmun. Rev. 10, 369-374. 2314 doi: 10.1016/j.autrev.2010.12.006
- Serhan, C. N. (1989). On the relationship between leukotriene and lipoxin 2315 production by human neutrophils: evidence for differential metabolism of 15-2316 HETE and 5-HETE. Biochim. Biophys. Acta 1004, 158-168. doi: 10.1016/0005-2317 2760(89)90264-6
- 2318 Serhan, C. N. (2005). Lipoxins and aspirin-triggered 15-epi-lipoxins are the 2319 first lipid mediators of endogenous anti-inflammation and resolution. Prostaglandins Leukot. Essent. Fatty Acids 73, 141-162. doi: 10.1016/j.plefa. 2320 2005.05.002 2321
- Serhan, C. N. (2014). Novel pro-resolving lipid mediators in inflammation are leads 2322 for resolution physiology. Nature 510, 92-101. doi: 10.1038/nature13479
- 2323 Serhan, C. N., Brain, S. D., Buckley, C. D., Gilroy, D. W., Haslett, C., O'Neill, L. A. J., et al. (2007). Resolution of inflammation: state of the art, definitions and terms. 2324 FASEB J. 21, 325-332. doi: 10.1096/fj.06-7227rev 2325
- Serhan, C. N., Chiang, N., and Dalli, J. (2015). The resolution code of acute 2326 inflammation: novel pro-resolving lipid mediators in resolution. Semin. 2327 Immunol. 27, 200-215. doi: 10.1016/j.smim.2015.03.004
- 2328 Serhan, C. N., Clish, C. B., Brannon, J., Colgan, S. P., Chiang, N., and Gronert, K. 2329 (2000). Novel functional sets of lipid-derived mediators with antiinflammatory actions generated from omega-3 fatty acids via cyclooxygenase 2-nonsteroidal 2330 antiinflammatory drugs and transcellular processing. J. Exp. Med. 192, 2331 1197-1204. doi: 10.1084/jem.192.8.1197
- 2332 Serhan, C. N., Hamberg, M., and Samuelsson, B. (1984). Lipoxins: novel series 2333 of biologically active compounds formed from arachidonic acid in human leukocytes. Proc. Natl. Acad. Sci. U.S.A. 81, 5335-5339. doi: 10.1073/pnas.81. 2334 17.5335 2335
- Serhan, C. N., Hong, S., Gronert, K., Colgan, S. P., Devchand, P. R., and 2336 Mirick, G. (2002). Resolvins: a family of bioactive products of omega-3 2337

fatty acid transformation circuits initiated by aspirin treatment that counter 2338 proinflammation signals. J. Exp. Med. 196, 1025-1037. doi: 10.1084/jem. 2339 20020760 2340

- Serhan, C. N., and Petasis, N. A. (2011). Resolvins and protectins in inflammation 2341 resolution. Chem. Rev. 111, 5922-5943. doi: 10.1021/cr100396c
- 2342 Serhan, C. N., and Sheppard, K. A. (1990). Lipoxin formation during human neutrophil-platelet interactions. Evidence for the transformation of leukotriene 2343 A4 by platelet 12-lipoxygenase in vitro. J. Clin. Invest. 85, 772-780. doi: 10.1172/ 2344 ICI114503 2345
- Serhan, C. N., Yacoubian, S., and Yang, R. (2008). Anti-inflammatory and 2346 proresolving lipid mediators. Annu. Rev. Pathol. 3, 279-312. doi: 10.1146/ annurev.pathmechdis.3.121806.151409 2347
- Serhan, C. N., Yang, R., Martinod, K., Kasuga, K., Pillai, P. S., Porter, T. F., et al. 2348 (2009). Maresins: novel macrophage mediators with potent antiinflammatory 2349 and proresolving actions. J. Exp. Med. 206, 15-23. doi: 10.1084/jem.20081880 2350
- Settimio, R., Clara, D. F., Franca, F., Francesca, S., and Michele, D. (2012). Resolvin 2351 D1 reduces the immunoinflammatory response of the rat eye following uveitis. Mediat. Inflamm. 2012:318621. doi: 10.1155/2012/318621 2352
- Sfikakis, P. P., Markomichelakis, N., Theodossiadis, G. P., Grigoropoulos, V., 2353 Katsilambros, N., and Theodossiadis, P. G. (2005). Regression of sight-2354 threatening macular edema in type 2 diabetes following treatment with the 2355 anti-tumor necrosis factor monoclonal antibody infliximab. Diabetes Care 28, 2356 445-447. doi: 10.2337/diacare.28.2.445
- Sha, Y. H., Hu, Y. W., Gao, J. J., Wang, Y. C., Ma, X., Qiu, Y. R., et al. (2015). Lipoxin 2357 A4 promotes ABCA1 expression and cholesterol efflux through the LXRalpha 2358 signaling pathway in THP-1 macrophage-derived foam cells. Int. J. Clin. Exp. 2359 Pathol. 8, 6708-6715.
- 2360 Sifuentes-Franco, S., Padilla-Tejeda, D. E., and Carrillo-Ibarra, S. (2018). Oxidative stress, apoptosis, and mitochondrial function in diabetic nephropathy. Int. J. 2361 Endocrinol. 2018:1875870. doi: 10.1155/2018/1875870 2362
- Singh, A., Boden, G., Homko, C., Gunawardana, J., and Rao, A. K. (2012). Whole-2363 blood tissue factor procoagulant activity is elevated in type 1 diabetes: effects 2364 of hyperglycemia and hyperinsulinemia. Diabetes Care 35, 1322-1327. doi: 2365 10.2337/dc11-2114
- Sivaraj, R. R., Oswal, K., Stavrou, P., and Murray, P. I. (2009). Diabetes mellitus and 2366 uveitis: a longitudinal study. Invest. Ophthalmol. Vis. Sci. 50, 296-296.

2367

2368

- Spite, M., Clària, J., and Serhan, C. N. (2014). Resolvins, specialized pro-resolving lipid mediators and their potential roles in metabolic diseases. Cell Metab. 19, 2369 21-36. doi: 10.1016/j.cmet.2013.10.006
- 2370 Srinivasan, S., Dehghani, C., Pritchard, N., Edwards, K., Russell, A. W., Malik, 2371 R. A., et al. (2017). Corneal and retinal neuronal degeneration in early stages of diabetic retinopathy. Invest. Ophthalmol. Vis. Sci. 58, 6365-6373. doi: 10.1167/ 2372 iovs.17-22736 2373
- Strutz, F., Zeisberg, M., Hemmerlein, B., Sattler, B., Hummel, K., Becker, V., et al. 2374 2000). Basic fibroblast growth factor expression is increased in human renal 2375 fibrogenesis and may mediate autocrine fibroblast proliferation. Kidney Int. 57, 1521-1538. doi: 10.1046/j.1523-1755.2000.00997.x 2376
- Stuebiger, N., Smiddy, W., Wang, J., Jiang, H., and DeBuc, D. C. (2015). Assessment 2377 of conjunctival microangiopathy in a patient with diabetes mellitus using the 2378 retinal function imager. J. Clin. Exp. Ophthalmol. 6:400. doi: 10.4172/2155-2379 9570.1000400
- Sugimoto, H., LeBleu, V. S., Bosukonda, D., Keck, P., Taduri, G., Bechtel, W., et al. 2380 (2012). Activin-like kinase 3 is important for kidney regeneration and reversal 2381 of fibrosis. Nat. Med. 18:396. doi: 10.1038/nm.2629 2382
- Sun, Y.-P., Tjonahen, E., Keledjian, R., Zhu, M., Yang, R., Recchiuti, A., et al. 2383 (2009). Anti-inflammatory and pro-resolving properties of benzo-lipoxin A(4) 2384 analogs. Prostaglandins Leukot. Essent. Fatty Acids 81, 357-366. doi: 10.1016/j. 2385 plefa.2009.09.004
- Sun, Y. B., Qu, X., Li, X., Nikolic-Paterson, D. J., and Li, J. (2013). Endothelial 2386 dysfunction exacerbates renal interstitial fibrosis through enhancing fibroblast 2387 Smad3 linker phosphorylation in the mouse obstructed kidney. PLoS One 8:e84063. doi: 10.1371/journal.pone.0084063
- 2389 Sun, Y. P., Oh, S. F., Uddin, J., Yang, R., Gotlinger, K., Campbell, E., et al. (2007). 2390 Resolvin D1 and its aspirin-triggered 17R epimer. Stereochemical assignments, anti-inflammatory properties, and enzymatic inactivation. J. Biol. Chem. 282, 2391 9323-9334. doi: 10.1074/jbc.M609212200 2392
- Tampe, D., and Zeisberg, M. (2014). Potential approaches to reverse or repair renal 2393 fibrosis. Nat. Rev. Nephrol. 10:226. doi: 10.1038/nrneph.2014.14 2394

Q16

2395	Tang, H., Liu, Y., Yan, C., Petasis, N. A., Serhan, C. N., and Gao, H. (2014).
2396	Protective actions of aspirin-triggered (17R) resolvin D1 and its analogue,
2397	17R-Hydroxy-19- < em > Para < /em > -fluorophenoxy-resolvin D1 methyl
	ester, in C5a-dependent IgG immune complex-induced inflammation and lung
2398	injury. J. Immunol

- 2399 Tayek, J. A., and Kalantar-Zadeh, K. (2013). The extinguished beacon of 2400 bardoxolone: not a monday morning quarterback story. Am. J. Nephrol. 37, 208-211. doi: 10.1159/000346950 2401
- Teumer, A., Tin, A., Sorice, R., Gorski, M., Yeo, N. C., Chu, A. Y., et al. 2402 (2015). Genome-wide association studies identify genetic loci associated with 2403 albuminuria in diabetes. Diabetes, 65, 803-817. doi: 10.2337/db15-1313
- 2404 The Lancet (2018). Untangling the complications of diabetes. Lancet 391:2389.
- Thiery, J. P., Acloque, H., Huang, R. Y., and Nieto, M. A. (2009). Epithelial-2405 mesenchymal transitions in development and disease. Cell 139, 871-890. doi: 2406 10.1016/j.cell.2009.11.007 2407
- Thorp, E., and Tabas, I. (2009). Mechanisms and consequences of efferocytosis 2408 in advanced atherosclerosis. J. Leukoc. Biol. 86, 1089-1095. doi: 10.1189/jlb. 2409 0209115
- Tian, H., Lu, Y., Sherwood, A. M., Hongqian, D., and Hong, S. (2009). Resolvins 2410 E1 and D1 in choroid-retinal endothelial cells and leukocytes: biosynthesis 2411 and mechanisms of anti-inflammatory actions. Invest. Ophthalmol. Vis. Sci. 50, 2412 3613-3620, doi: 10.1167/jovs.08-3146
- 2413 Tinckam, K. J., Djurdjev, O., and Magil, A. B. (2005). Glomerular monocytes predict worse outcomes after acute renal allograft rejection independent of C4d 2414 status. Kidney Int. 68, 1866-1874. doi: 10.1111/j.1523-1755.2005.00606.x 2415
- Todd, J. N., Dahlstrom, E. H., Salem, R. M., Sandholm, N., Forsblom, C., McKnight, 2416 A. J., et al. (2015). Genetic evidence for a causal role of obesity in diabetic kidney 2417 disease. Diabetes Metab. Res. Rev. 64, 4238-4246. doi: 10.2337/db15-0254
- 2418 Tomkin, G. H. (2014). Treatment of type 2 diabetes, lifestyle, GLP1 agonists and DPP4 inhibitors. World J. Diabetes 5, 636-650. doi: 10.4239/wjd.v5.i5.636 2419
- Tuomi, T. (2005). Type 1 and type 2 diabetes. What Do They Have in Common? 2420 Diabetes 54(Suppl 2), S40-S45. doi: 10.2337/diabetes.54.suppl\_2.S40 2421
- Tuttle, K. R., Brosius, F. C., Adler, S. G., Kretzler, M., Mehta, R. L., Tumlin, J. A., 2422 et al. (2018). JAK1/JAK2 inhibition by baricitinib in diabetic kidney disease: 2423 results from a Phase 2 randomized controlled clinical trial. Nephrol. Dial. 2424 Transpl. 33, 1-10. doi: 10.1093/ndt/gfx377
- Vallejo, S., Palacios, E., Romacho, T., Villalobos, L., Peiro, C., and Sanchez-2425 Ferrer, C. F. (2014). The interleukin-1 receptor antagonist anakinra improves 2426 endothelial dysfunction in streptozotocin-induced diabetic rats. Cardiovasc. 2427 Diabetol. 13:158. doi: 10.1186/s12933-014-0158-z
- 2428 van Zuydam, N. R., Ahlqvist, E., and Sandholm, N. (2018). A genome-wide association study of diabetic kidney disease in subjects with type 2 diabetes. 2429 Diabetes 67, 1414-1427. doi: 10.2337/db17-0914 2430
- vanDam, B., Demirci, C., Reitsma, H. J., van Lambalgen, A. A., van den Bos, G. C., 2431 Tangelder, G. J., et al. (2000). Alterations in nitric oxide activity and sensitivity 2432 in early streptozotocin-induced diabetes depend on arteriolar size. Int. J. Exp. 2433 Diabetes Res. 1, 221-232. doi: 10.1155/EDR.2000.221
- van Dieren S, Beulens JW, van der Schouw YT, Grobbee DE, Neal B. (2010). The 2434 global burden of diabetes and its complications: an emerging pandemic. Eur. J. 2435 Cardiovasc. Prev. Rehabil. 17(1 Suppl.), s3-s8.
- 2436 Vilaysane, A., Chun, J., Seamone, M. E., Wang, W., Chin, R., Hirota, S., et al. (2010). 2437 The NLRP3 inflammasome promotes renal inflammation and contributes to CKD. J. Am. Soc. Nephrol. 21, 1732-44. doi: 10.1681/ASN.2010020143 2438
- Voelker, J., Berg, P. H., Sheetz, M., Duffin, K., Shen, T., Moser, B., et al. (2017). 2439 Anti-TGF-beta1 antibody therapy in patients with diabetic nephropathy. J. Am. 2440 Soc. Nephrol. 28, 953-962. doi: 10.1681/ASN.2015111230
- 2441 Vukicevic, S., Basic, V., Rogic, D., Basic, N., Shih, M. S., Shepard, A., et al. 2442 (1998). Osteogenic protein-1 (bone morphogenetic protein-7) reduces severity 2443 of injury after ischemic acute renal failure in rat. J. Clin. Invest. 102, 202-214. doi: 10.1172/JCI2237 2444
- Wang, B., Jha, J. C., Hagiwara, S., McClelland, A. D., Jandeleit-Dahm, K., Thomas, 2445 M. C., et al. (2014). Transforming growth factor-beta1-mediated renal fibrosis
- 2446 is dependent on the regulation of transforming growth factor receptor 1 2447 expression by let-7b. Kidney Int. 85, 352-361. doi: 10.1038/ki.2013.372
- Wang, H., and Daggy, B. P. (2017). The role of fish oil in inflammatory eye diseases. 2448 Biomed. Hub. 2, 6-6. doi: 10.1159/000455818 2449
- Weber, C., and von Hundelshausen, P. (2017). CANTOS Trial validates the 2450 inflammatory pathogenesis of atherosclerosis: setting the stage for a new 2451

chapter in therapeutic targeting. Circ. Res. 121, 1119-1121. doi: 10.1161/ 2452 CIRCRESAHA 117 311984 2453

- Wijesekara, N.,. Zhang, L.-H., Kang, M. H., Abraham, T., Bhattacharjee, A., 2454 Warnock, G. L., et al. (2012). miR-33a modulates ABCA1 expression, 2455 cholesterol accumulation, and insulin secretion in pancreatic islets. Diabetes 61, 2456 653-658. doi: 10.2337/db11-0944
- Wild, S., Roglic, G., Green, A., Sicree, R., and King, H. (2004). Global prevalence 2457 of diabetes. Estimates for the year 2000 and projections for 2030. Diabetes Care 2458 27, 1047-1053. doi: 10.2337/diacare.27.5.1047 2459
- Wilkinson, C. P., Ferris, F. L. III, Klein, R. E., Lee, P. P., Agardh, C. D., 2460 Davis, M., et al. (2003). Proposed international clinical diabetic retinopathy and diabetic macular edema disease severity scales. Ophthalmology 110, 1677-1682. 2461 doi: 10.1016/S0161-6420(03)00475-5 2462
- Williams, K. J., and Tabas, I. (2005). Lipoprotein retention-and clues for atheroma 2463 regression. Arterioscler. Thromb. Vasc. Biol. 25, 1536-1540. doi: 10.1161/01. 2464 ATV.0000174795.62387.d3
- 2465 Wolfs, I. M., Donners, M. M., and de Winther, M. P. (2011). Differentiation factors 2466 and cytokines in the atherosclerotic plaque micro-environment as a trigger for macrophage polarisation. Thromb. Haemost. 106, 763-771. doi: 10.1160/TH11-2467 05-0320 2468
- Wu, S.-H., Chen, X.-Q., Liu, B., Wu, H.-J., and Dong, L. (2013). Efficacy and safety 2469 of 15(R/S)-methyl-lipoxin A4 in topical treatment of infantile eczema. Br. J. 2470 Dermatol. 168, 172-178. doi: 10.1111/j.1365-2133.2012.11177.x
- Wu, S. H., Liao, P. Y., Dong, L., and Jiang, X. Y. (2006). Protective effects of 15-2471 methyl-lipoxin A4 on mesangioproliferative nephritis in rats. Zhongguo Dang 2472 Dai Er Ke Za Zhi 8, 225-230. 2473
- Wu, S. H., Liao, P. Y., Yin, P. L., Zhang, Y. M., and Dong, L. (2009). 2474 Elevated expressions of 15-lipoxygenase and lipoxin A4 in children with acute poststreptococcal glomerulonephritis. Am. J. Pathol. 174, 115-122. doi: 10. 2475 2353/ajpath.2009.080671 2476
- Wu, S. H., Wu, X. H., Liao, P. Y., and Dong, L. (2007). Signal transduction involved 2477 in protective effects of 15(R/S)-methyl- lipoxin A(4) on mesangioproliferative 2478 nephritis in rats. Prostaglandins Leukot. Essent. Fatty Acids 76, 173-180. 2479 doi: 10.1016/j.plefa.2006.12.006
- Wu, Y., Tang, L., and Chen, B. (2014). Oxidative stress: implications for the 2480 development of diabetic retinopathy and antioxidant therapeutic perspectives. Oxid. Med. Cell. Longev. 2014:752387. doi: 10.1155/2014/752387

2481

2486

2498

2500

2501

2502

2505

- 2482 Wuttke, M., and Köttgen, A. (2016). Insights into kidney diseases 2483 from genome-wide association studies. Nat. Rev. Nephrol. 12:549. 2484 doi: 10.1038/nrneph.2016.107
- Xu, H., Curtis, T., and Stitt, A. (2014). Pathophysiology and Pathogenesis of Diabetic 2485 Retinopathy, Amsterdam: Diapedia, The Living Textbook of Diabetes.
- Yach, D., Stuckler, D., and Brownell, K. D. (2006). Epidemiologic and economic 2487 consequences of the global epidemics of obesity and diabetes. Nat. Med. 12: 62. 2488 doi: 10.1038/nm0106-62
- 2489 Yanagita, M. (2012). Inhibitors/antagonists of TGF-B system in kidney fibrosis. Nephrol. Dial. Transpl. 27, 3686-3691. doi: 10.1093/ndt/gfs381 2490
- Yau, J. W., Rogers, S. L., Kawasaki, R., Lamoureux, E. L., Kowalski, J. W., Bek, T., 2491 et al. (2012). Global prevalence and major risk factors of diabetic retinopathy. 2492 Diabetes Care 35, 556-564. doi: 10.2337/dc11-1909
- 2493 Yin, Y., Chen, F., Wang, W., Wang, H., and Zhang, X. (2017). Resolvin D1 inhibits inflammatory response in STZ-induced diabetic retinopathy rats: possible 2494 involvement of NLRP3 inflammasome and NF-kappaB signaling pathway. Mol. 2495 Vis. 23, 242-250. 2496
- Yoshida, S., Yoshida, A., Ishibashi, T., Elner, S. G., and Elner, V. M. (2003). Role 2497 of MCP-1 and MIP-1alpha in retinal neovascularization during postischemic inflammation in a mouse model of retinal neovascularization. J. Leukoc. Biol. 2499 73, 137-144. doi: 10.1189/jlb.0302117
- You, W.-P., and Henneberg, M. (2016). Type 1 diabetes prevalence increasing globally and regionally: the role of natural selection and life expectancy at birth. BMJ Open Diabetes Res. Care 4:e000161. doi: 10.1136/bmjdrc-2015-000161
- Young, B. A., Burdmann, E. A., Johnson, R. J., Alpers, C. E., Giachelli, C. M., 2503 Eng, E., et al. (1995). Cellular proliferation and macrophage influx precede 2504 interstitial fibrosis in cyclosporine nephrotoxicity. Kidney Int. 48, 439-448. doi: 10.1038/ki.1995.312
- Yurdagul, A., Doran, A. C., Cai, B., Fredman, G., and Tabas, I. A. (2017). 2506 Mechanisms and consequences of defective efferocytosis in atherosclerosis. 2507 Front. Cardiovasc. Med. 4:86. doi: 10.3389/fcvm.2017.00086 2508

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- Zeisberg, M., and Duffield, J. S. (2010). Resolved: EMT produces fibroblasts in the kidney. J. Am. Soc. Nephrol. 21, 1247–1253. doi: 10.1681/ASN.2010060616
- Zhang, J., Pippin, J. W., Krofft, R. D., Naito, S., Liu, Z. H., and Shankland, S. J. (2013). Podocyte repopulation by renal progenitor cells following glucocorticoids treatment in experimental FSGS. *Am. J. Physiol. Renal Physiol.* 304, F1375–F1389. doi: 10.1152/ajprenal.00020.2013
- Zhang, M. J., Sansbury, B. E., Hellmann, J., Baker, J. F., Guo, L., Parmer, C. M., et al.
  (2016). Resolvin D2 enhances post-ischemic revascularization while resolving
  inflammation. *Circulation* 134, 666–680. doi: 10.1161/CIRCULATIONAHA.
  116.021894
- Zhao, Y.-L., Zhang, L., Yang, Y.-Y., Tang, Y., Zhou, J.-J., Feng, Y.-Y., et al. (2016).
  Resolvin D1 protects lipopolysaccharide-induced acute kidney injury by downregulating nuclear factor-kappa b signal and inhibiting Apoptosis. *Chin. Med. J.*129, 1100–1107. doi: 10.4103/0366-6999.180517
- Zimmet, P., Alberti, K. G., and Shaw, J. (2001). Global and societal implications of the diabetes epidemic. *Nature* 414:782. doi: 10.1038/414782a 2567

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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