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Title: Potential Role of Mesenchymal Stem Cells (MSCs) in the Breast Tumor Microenvironment: Stimulation of Epithelial to Mesenchymal Transition (EMT)

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4 **Abstract**
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6 Bone marrow derived Mesenchymal Stem Cells (MSCs) are known to specifically
7 migrate to and engraft at tumor sites. Understanding interactions between cancer cells
8 and MSCs has become fundamental to determining whether MSC-tumour interactions
9 should be harnessed for delivery of therapeutic agents or considered a target for
10 intervention. Breast Cancer Cell lines (MDA-MB-231, T47D & SK-Br3) were cultured
11 alone or on a monolayer of MSCs, and retrieved using epithelial specific magnetic beads.
12 Alterations in expression of 90 genes associated with breast tumorigenicity were analyzed
13 using low density array. Expression of markers of Epithelial-Mesenchymal transition and
14 array results were validated using RQ-PCR. Co-cultured cells were analyzed for changes
15 in protein expression, growth pattern, and morphology. Gene expression and proliferation
16 assays were also performed on indirect co-cultures. Following direct co-culture with
17 MSCs, breast cancer cells expressed elevated levels of oncogenes (NCOA4, FOS), proto-
18 oncogenes (FYN, JUN), genes associated with invasion (MMP11), angiogenesis (VEGF)
19 and anti-apoptosis (IGF1R, BCL2). However, universal downregulation of genes
20 associated with proliferation was observed (Ki67, MYBL2), and reflected in reduced
21 ATP production in response to MSC-secreted factors. Significant upregulation of
22 Epithelial-Mesenchymal Transition specific markers (N-cadherin, Vimentin, Twist and
23 Snail) was also observed following co-culture with MSCs, with a reciprocal
24 downregulation in E-cadherin protein expression. These changes were predominantly cell
25 contact mediated and appeared to be MSC specific. Breast cancer cell morphology and
26 growth pattern also altered in response to MSCs. Mesenchymal Stem Cells may promote
27 breast cancer metastasis through facilitation of Epithelial-Mesenchymal Transition.
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48 **Keywords:** Mesenchymal Stem Cells (MSCs), Breast Cancer, Epithelial-Mesenchymal
49 Transition (EMT), Invasion, Co-culture
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4 **Introduction**
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6 Breast cancer remains the most common malignancy in women, accounting for one
7 quarter of all female cancers [1]. The preferential spread of tumour cells to bone and
8 subsequent development of osteolytic metastatic deposits remains a devastating event in
9 the course of the disease [2, 3]. It is now understood that tumour epithelial cells develop
10 in a symbiotic rather than an independent manner with surrounding stroma. This stromal
11 environment consists of a dynamic network of immune cells, fibroblasts, tumour
12 vasculature and extracellular matrix [4]. Tumours actively recruit cells, including bone
13 marrow derived mesenchymal stem cells (MSCs), into the tumour microenvironment and
14 these cells may play a role in facilitating cancer progression [5]. MSCs are a subset of
15 non-haematopoietic cells found within the bone marrow stroma that have an innate ability
16 both to self renew and to differentiate into cells of multiple lineages, including
17 osteoblasts, chondrocytes and adipocytes [6]. They have also been seen to influence the
18 morphology and proliferation of cells within their vicinity through both cell to cell
19 interactions and the secretion of chemoattractant cytokines and paracrine factors [7-10].
20 Studies assessing systemically delivered MSCs have confirmed that these circulating
21 cells engraft and facilitate healing at sites of inflammation and injury including head
22 trauma, stroke and myocardial infarction [10, 11]. Malignancy may also be considered as
23 a nidus of chronic inflammation or “wound that never heals” [12] and reports have shown
24 a similar pattern of MSC engraftment at these sites [11]. This tumor homing ability has
25 prompted researchers to analyse MSCs as possible vectors for the targeted delivery of
26 anti-cancer agents to tumor microenvironments [13]. However evidence suggests that
27 interactions between MSCs and breast cancer cells may impact upon the phenotype of the
28 cancer cell and promote their metastatic potential [7-9, 14-16]. Understanding these
29 interactions has become fundamental to determining whether the homing ability of MSCs
30 should be harnessed for delivery of therapeutic agents or whether the MSC-tumour
31 interactions should be considered a target for intervention.
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4 cell adhesion molecules E-cadherin and Epithelial Specific Antigen (ESA) [7, 8].
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6 Conflicting reports exist with relation to the effect of MSCs on proliferation of breast
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8 cancer cells, with some studies reporting no change [7] and others suggesting
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10 proliferative changes occurring in an estrogen dependant manner [8, 9].

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12 More recently a pivotal study by Karnoub et al [14] reported that, when mixed with
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14 breast cancer cells prior to implantation, MSCs enhance breast cancer cell motility,
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16 invasion and metastatic potential in vivo. Knockdown of the CCL5–CCR5 loop led to an
17
18 abrogated metastatic response confirming that these paracrine interactions play an
19
20 important role in MSC-mediated metastatic spread [14]. These studies highlight the
21
22 distinct effect that MSCs have on breast cancer cells, and thus understanding the
23
24 pathways governing these effects remains imperative.

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26 Epithelial to Mesenchymal Transition (EMT) is a process essential to organogenesis
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28 during embryonic development [17], however its reactivation during adult life has been
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30 ascribed to certain pathological processes including the facilitation of carcinogenesis
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32 [18]. EMT has been shown to promote the detachment of cancer cells from the primary
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34 tumour and facilitate their subsequent migration through the acquisition of stem like
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36 properties, including a loss of cellular polarity, adhesion and proliferation [18, 19].
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38 Studies have demonstrated evidence of EMT in primary human breast carcinomas
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40 showing a proclivity toward the more invasive basal breast cancer phenotype [20, 21].
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42 Despite recognition of the role EMT plays in the metastatic cascade, stimuli inducing
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44 EMT at the primary tumour site remain largely unknown.

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46 Further understanding of MSC/tumour cell interactions is required to determine their role
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48 in breast cancer progression or therapy. This study aimed to further elucidate the effect
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50 MSCs have on breast cancer cells and to potentially identify pathways mediating these
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52 effects.

53 **Materials and Methods**

54 *Cell Culture*

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56 Breast cancer cell lines included MDA-MB-231 cells cultured in Liebowitz-15 medium
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58 (L-15); T47D cells cultured in RPMI 1640 medium; and SK-Br-3 cells cultured in
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60 McCoys-5a medium. Normal human embryonic lung fibroblasts (WI-38 cells) were
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4 cultured in Eagle's minimal essential medium (EMEM). All media were supplemented
5 with 10% Fetal Bovine Serum (FBS), 100 IU/ml Penicillin /100µg/ml Streptomycin (P/S)
6 and 1% L-glutamine.
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10 Mesenchymal Stem Cell (MSCs) were supplied by the Regenerative Medicine Institute
11 (REMEDI) at NUI Galway. With ethical approval and informed consent, bone marrow
12 was aspirated from the iliac crests of healthy donors following a defined clinical protocol
13 [22]. MSCs were isolated from the marrow aspirates by direct plating and subsequently
14 cultured for 12-15 days to deplete the non-adherent haematopoietic cell fraction. Cells
15 were maintained in Dulbecco's Modified Eagle's Medium (DMEM) supplemented with
16 pre-selected FBS (10%) and P/S. The ability of MSCs to differentiate into chondrocytes,
17 adipocytes and osteoblasts was confirmed prior to use. Characterization of surface
18 receptors was performed targeting the markers CD105, CD73, CD90 (positive) and
19 CD34, CD45 (negative). MSCs derived from three separate donors were utilised for
20 experiments. All cells were maintained at 37°C and 5% CO₂.
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32 *Direct Co-Culture:* Primary MSCs or normal fibroblasts (WI-38 cells) were seeded at a
33 density of 2×10^4 cells/cm² and allowed to adhere overnight. Breast cancer cell lines were
34 then seeded at a density of 1.3×10^4 cells/cm² onto the monolayers of MSCs or normal
35 fibroblasts. All cell types were cultured individually in parallel as controls. Cells were
36 maintained in MSC specific medium and following a 3 or 7 day incubation, media was
37 harvested and epithelial cells retrieved as described below.
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45 *Retrieval of Epithelial Cells:* Following direct co-culture epithelial cells were separated
46 from MSCs in co-culture using an EasySep[®] positive selection kit (Stem Cell
47 Technologies). As per manufacturer's instructions, co-culture populations were
48 trypsinized and dispersed into a single cell suspension, and EasySep[®] positive selection
49 cocktail and magnetic nanoparticles were added during serial incubations on ice. The
50 magnetic nanoparticles bind selectively to viable epithelial cells which are positively
51 selected by placing the tube in a magnet. Retrieved cells were centrifuged and stored at -
52 80°C until required for RNA extraction.
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4 *Gene Expression:* RNA was extracted from both cells cultured alone and epithelial cells
5 retrieved following co-culture with MSCs or WI-38 cells using the RNeasy[®] Mini Kit
6 (QIAGEN Ltd.) following manufacturer's protocol. cDNA was generated using
7 SuperScript III reverse transcription enzyme and analyzed by both Taqman[®] Low-
8 Density array (TLDA) and relative quantitative-PCR (RQ-PCR). The array plate was
9 designed to simultaneously measure expression of 90 genes specifically associated with
10 breast cancer tumorigenicity and 6 endogenous controls. Following identification of target
11 genes of interest, co-culture experiments were repeated in triplicate and results validated
12 by RQ-PCR using the ABI Prism 7000 sequence detector system (Applied Biosystems).
13 PreDeveloped Taqman[®] Assay Reagents (PDARS) specific to genes associated with
14 EMT, including N-cadherin, Vimentin, Twist and Snail, were also used to quantify
15 changes in expression by RQ-PCR. The comparative C_T method was used to quantify
16 expression of genes and this was normalized to the endogenous control, Peptidyl-Prolyl
17 Isomerase A (PPIA). Results from cells retrieved from co-culture were expressed relative
18 to the same cells cultured alone. Changes in gene expression were expressed using the 2⁻
19 $\Delta\Delta C_T$ method [23] and the fold change in triplicate experiments was recorded and
20 presented as Mean \pm SEM.
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37 *Collection of Conditioned Media (CM) for indirect co-culture:* MSCs were seeded at a
38 density of 2 x 10⁴ cells/cm² in DMEM supplemented with pre-selected FBS (10%) and
39 P/S. Media was aspirated at 24hr intervals and transferred to breast cancer cells lines to
40 determine the effect of MSC secreted factors on cell proliferation and gene expression
41 (indirect co-culture) as described below. Breast cancer cells grown in MSC medium that
42 had not been exposed to MSCs served as a control.
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50 *Proliferation Assay:* Breast cancer cell lines were seeded onto 96 well white walled
51 plates at a density of 8 x 10³ cells/well in 100 μ l media and allowed to adhere overnight.
52 Media was aspirated and replaced with MSC-conditioned medium (CM) as described
53 above at 24hr intervals for 72hrs. An Apoglow[®] assay was performed to assess changes
54 in proliferation based on the level of ATP production as quantified by a luminometer.
55 Results presented represent triplicate experiments and are expressed as Mean \pm SEM.
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6 *Indirect co-culture:* Breast cancer cells were cultured in media that had been exposed to
7 MSCs as described above. Following culture in MSC CM, cells were lysed and RNA
8 extracted. Changes in expression of genes associated with EMT were quantified by RQ-
9 PCR as described for the direct co-culture experiments.
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15 *Western Blot Analysis.* Protein was extracted from cells cultured individually and those
16 retrieved following co-culture. Briefly, cells were washed and resuspended in Triton-X
17 lysis buffer [150mM NaCl, 20 mM HEPES, 2 mM EDTA, 1% Triton-X100, 2mM
18 Sodium Orthovanadate, 10mM Sodium Fluoride, 10ul/mL Protease inhibitor cocktail
19 (Fisher Scientific)], frozen at -20°C and then centrifuged at $500 \times g$ for 15 mins at 4°C
20 to remove cellular debris. The protein content was determined using the Micro BCA™
21 Protein Assay Kit (Thermo Scientific). Protein (40 μg) was reduced in DTT (0.5 M) for
22 10 mins at 70°C and samples run on a 4-12% gradient pre-cast NuPAGE Bis-Tris
23 polyacrylamide gel for 1 hr at 200V. Protein molecular weight standards (20-220 kDa)
24 were run simultaneously on each gel. Electroblothing was performed for 1hr at 25V to
25 transfer protein samples to a nitrocellulose membrane. Blots were blocked in 5% milk in
26 TBS-T [20 mM Tris, 137 mM NaCl, 0.1% Tween-20] for 1 hr, and probed with
27 antibodies targeting E-cadherin (1 $\mu\text{g}/\text{mL}$, R & D Systems), Vimentin (1:100, Abcam), or
28 Snail (1 $\mu\text{g}/\text{mL}$, Abcam) for 1.5 hrs and washed in TBS-T. β -actin was used to confirm
29 equal loading in wells. Horseradish peroxidase labelled goat anti-rabbit (1:3,000; Abcam)
30 or rabbit anti-mouse antibody (1:2,000; Abcam) was then added to the membranes for 1.5
31 hrs. Following washing steps, SuperSignal West Pico Chemiluminescent substrate
32 (Thermo Scientific) was applied to the membranes for 5min. Images were captured using
33 a Syngene G-Box and GeneSnap software.
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51 *Immunohistochemistry & Fluorescent Microscopy:* After 72hrs co-culture in chamber
52 slides, cells were fixed in methanol. Immunohistochemical analysis was performed using
53 monoclonal antibodies targeting E-cadherin (R&D Systems), MNF116 pancytokeratin
54 (Dako, Denmark) and CD90 (Dako, Denmark). E-cadherin and CD90 were visualized
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4 using the chromagen 3,3'- Diaminobenzidine (DAB), with Acid fast red (RED) used for
5 detection of pancytokeratin in dual staining experiments.
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8 To assess changes in breast cancer cell morphology in response to MSCs, cells were dual
9 labelled and examined by fluorescence microscopy. Prior to mixing the cell populations,
10 epithelial breast cancer cells were labelled with PKH26 (red fluorescent label, Excitation
11 551nm, Emmission 567nm, Sigma). Following 72hrs co-culture, cells were fixed in 4%
12 paraformaldehyde and the cytoskeleton of the mixed populations was labelled with
13 Alexafluor[®] 488 phalloidin (green fluorescent label, Excitation 495nm, Emmission
14 518nm, Invitrogen, Eugene, OR). Cells were examined using an Olympus IX81-ZDC[®]
15 microscope and Confocal Andor Revolution spinning disc system[®].
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24 **Results**

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26 *Cell Separation:* Cell separation using the EasySep[®] positive selection kit (Stem Cell
27 Technologies) was assessed. Following two washes, a positive retrieval rate of 94.4 ±
28 1.1% was achieved (range 92 - 97.5% retrieval). It has previously been shown in an
29 extensive study by Woelfle et al. [24], that the immunoselection procedure does not alter
30 breast cancer cell gene expression. To further confirm this, expression of Vimentin, E-
31 cadherin, CXCL12 and CXCR4 in breast cancer cells selected with beads, was compared
32 to unselected cells with a <1-fold change in gene expression detected.
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41 *Analysis of Gene Expression:* Low density array analysis of 90 genes associated with
42 breast cancer tumorigenicity was performed on all breast cancer cell lines retrieved
43 following 72hrs co-culture with MSCs, relative to the same cells cultured alone. Any
44 change >2.5 fold is presented (*Table 1*). Upregulation of oncogenes, proto-oncogenes and
45 genes associated with angiogenesis, anti-apoptosis and invasion was observed. A range of
46 genes exhibited greater than 10 fold upregulation (FOS, FYN, MET, VEGF, CD68 and
47 MMP11) while others were upregulated over 1,000 fold (CAV-1, TGFβR2 and
48 CXCL12). However down-regulation of genes associated with proliferation (Ki67,
49 CCNE1 and MYBL2) was recorded across all breast cancer cells following co-culture
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4 Observed changes in specific genes of interest were validated in triplicate experiments
5 using RQ-PCR (Figure 1). Significant upregulation of the chemokine, CXCL12, was
6 observed in SK-Br3 cells ($9,949 \pm 4,787$ fold, $p < 0.05$) following co-culture with a
7 reciprocal downregulation in its cognate receptor CXCR4 (3 ± 1 fold). In MDA-MB-231
8 cells, CXCL12 expression was significantly increased ($17,066 \pm 1,109$ fold) whereas
9 T47D cells exhibited upregulation of its receptor, CXCR4 (6 ± 2 fold, $p < 0.05$). The
10 proliferation marker, Ki-67, was downregulated in all breast cancer cells (Range: 2 – 4
11 fold decrease, T47D $p < 0.05$), while the invasive marker, MMP11, was significantly
12 upregulated.
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22 *Cell Proliferation in response to MSC secreted factors:* Breast cancer cells were cultured
23 in the presence of MSC CM for 72 hours after which ATP levels were quantified using a
24 luminometer based Apoglow[®] Assay (Figure 2). There was a significant reduction in
25 proliferation observed in all three breast cancer cell lines cultured in the presence of
26 factors secreted by MSCs (SK-Br-3 $p < 0.05$; T47D and MDA-MB-231 $p < 0.001$).
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33 *Expression of markers associated with EMT:* Significant upregulation in defined markers
34 of EMT were observed in both SK-Br3 and T47D cells retrieved following 72hrs co-
35 culture with MSCs (Figure 3a). Due to the magnitude of the increases seen, results are
36 expressed as Log_{10} values. Upregulation of most EMT markers in the MDA-MB-231 cell
37 line occurred to a lesser degree: Vimentin (3 fold), Snail (5 fold), N-cadherin (50 fold),
38 while Twist expression increased $>10,000$ fold. To determine whether the effects seen
39 were transient, T47D and Sk-Br3 cells were also retrieved following 7 days direct co-
40 culture with MSCs. In the case of the T47D cells, a significant increase in Vimentin (244-
41 fold) and Snail (5-fold) was still detected, while Twist and N-cadherin had returned to
42 baseline. At Day 7 the SK-Br3 cells retained increased expression of N-cadherin (28-
43 fold), Vimentin (153-fold) and Snail (10-fold).
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53 To determine whether the changes in gene expression were detected at the protein level,
54 protein was extracted from cells cultured individually and those retrieved following co-
55 culture. Lysates were then subjected to western blot using antibodies directed against
56 Vimentin, Snail (Figure 3b) and E-cadherin (Figure 5c). To confirm that differences seen
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4 were not as a result of variation in protein sample, β -actin was also targeted and found to
5 be at similar levels in all samples. Increased expression of Vimentin and Snail protein
6 was detected in both Sk-Br3 and T47D protein lysates harvested from cells retrieved
7 following direct co-culture with MSCs (Figure 3b).
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11 Overall the greatest increase in all EMT markers examined was seen in Sk-Br3 cells. To
12 determine whether this was an MSC specific effect, SK-Br3 cells were cultured directly
13 on a confluent monolayer of normal fibroblasts (WI-38 cells). No significant change in
14 expression of genes associated with EMT was observed following co-culture with WI-38
15 cells. Mesenchymal markers N-cadherin and Vimentin were downregulated 1.5 and 2.1
16 fold respectively, with expression of the transcription factors Twist and Snail both
17 decreased by 1.5 and 1.4 fold respectively (results not shown).
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26 *Expression of EMT markers following indirect co-culture:* To determine whether results
27 observed were due to cell contact mediated effects, breast cancer cells were exposed to
28 MSC conditioned medium and changes in expression of the same EMT markers were
29 analysed (Figure 4). In T47D and SK-Br3 cells, a small increase in expression of Twist
30 and Snail was observed (range 1 - 2 fold and 4 – 7 fold respectively). A greater
31 upregulation was seen in N-cadherin (range 9 – 32 fold) with the most marked increase
32 observed in vimentin expression (range 158 - 276 fold). Although the changes in
33 expression were significant ($p < 0.05$) for Snail, Twist and Vimentin, the increase was
34 considerably lower than that seen in the same cells following direct co-culture with
35 MSCs (Figure 3). When the length of exposure to MSC secreted factors was increased to
36 7 days, levels of target expression had returned to baseline (< 2 -fold change in gene
37 expression compared to cells cultured in standard medium). No change in expression of
38 EMT markers was observed in MDA-MB-231 cells following indirect co-culture (results
39 not shown).
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53 *Immunohistochemistry:* Breast cancer cells and MSCs cultured individually and in co-
54 culture were stained with cell type specific antibodies to distinguish populations and
55 analyse changes in morphology and growth pattern. Changes in E-cadherin protein
56 expression were also examined. E-cadherin has strong membrane targeted expression in
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4 T47D cells (Figure 5a), while MSCs have no detectable expression. When T47D cells
5 were cultured on a monolayer of MSCs a marked decrease was observed in the intensity
6 of E-cadherin staining (Figure 5b). E-cadherin expression was particularly reduced at
7 junctions where T47D cells were in direct contact with the MSCs (indicated by arrows)
8 compared with cells located within a cluster of breast cancer cells. This change in E-
9 cadherin protein expression was confirmed by western blot of T47D protein lysates
10 harvested from cells cultured individually, and those retrieved following co-culture
11 (Figure 5c).
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19 *Dual Staining:* Breast cancer cells (MDA-MB-231) cultured alone stained positive for the
20 epithelial specific cytokeratin, MNF116 (red), with nuclei counterstained with
21 haematoxylin (blue), and grew in a typical random asymmetric pattern (Figure 5d).
22 Stromal cells (MSCs) staining positive for CD90 grew in a symmetrical pattern with a
23 typical parallel alignment of spindle shaped cells when cultured alone. When cultured on
24 a monolayer of MSCs, MDA-MB-231 cells altered their growth pattern from the random
25 cellular distribution observed to align in parallel with adjacent MSCs (Figure 5e)
26 reflecting a change in cellular polarity.
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33 *Confocal Fluorescent Microscopy:* PKH26 labelled (red) T47D cells when cultured alone
34 were seen to grow in a typical clustered growth pattern, with the Alexafluor labelled cell
35 cytoskeleton (green) seen to be non-branching and closely adherent to the nuclei (Figure
36 5f). These same cells, when co-cultured directly on a monolayer of MSCs, appeared to
37 lose cellular adhesion leading to a more dispersed single cell distribution. Furthermore,
38 the breast cancer cell cytoskeleton was more branching and elongated (indicated by
39 arrows), and appeared to polarize in the direction of adjacent Mesenchymal Stem cells
40 (Figure 5g).
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50 **Discussion**

51 Mesenchymal Stem Cells have been reported to interact with breast cancer cells that have
52 metastasised to bone marrow [25] as well as being actively recruited to the primary
53 tumour stromal interface [15]. This tumour homing quality has prompted investigators to
54 assess MSCs as possible delivery vectors for anti-cancer therapies [13]. To realise their
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4 therapeutic potential, interactions between MSCs and breast cancer cells must be fully
5 elucidated.
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8 Studies have previously analysed breast cancer cells and MSCs in direct co-culture noting
9 specific morphological and phenotypical alterations in the breast cancer cells [7-9, 14].
10 However isolation of the cells following co-culture and analysis of changes in gene
11 expression has not previously been assessed. Immunomagnetic selection targeting
12 antigens such as EpCAM is used to capture circulating tumor cells or enrich tumor cells
13 from mixed cell samples. The immunomagnetic enrichment technique itself has
14 previously been shown to have no significant effect on the gene expression profile of
15 breast cancer cells [24].
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18 Reports from this laboratory and others have shown a significant increase in migration of
19 breast cancer cells in response to factors secreted by MSCs [25, 26], and this was
20 reflected by increased expression of migratory genes seen here including MMP11 and
21 CXCL12 [27]. Oncogenes and proto-oncogenes were upregulated both in a cell specific
22 manner and, in the case of FOS and JUN, across all breast cancer cells retrieved
23 following co-culture with MSCs. FOS and JUN are both major components of the
24 activator protein-1 (AP-1) transcription factor complex which has been shown to
25 positively regulate cellular motility and migration [28].
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28 CAV-1 is now considered a marker of poor prognosis with up-regulation correlated to
29 increased cellular dissemination and cell survival [29]. CAV-1 has been shown to
30 mediate its anti-apoptotic properties through upregulation of IGF-1R which was also
31 elevated in the cells following co-culture with MSCs. Interestingly a universal
32 downregulation of genes associated with proliferation (Ki-67, MYBL2, CCNE1) was
33 observed in all breast cancer cells retrieved from co-culture. Subsequent analysis of ATP
34 production by breast cancer cells in the presence of MSC secreted factors (indirect co-
35 culture), revealed a significant reduction in proliferation of all cancer cells. These results
36 concur with those of Hombauer and Minguell [7] who noted no increase in proliferative
37 activity when MCF-7 cells were grown alone or on a monolayer of MSCs.
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40 The apparent promotion of oncogenes and genes associated with invasion and migration
41 with an inhibition of proliferation fits a profile seen in EMT [18, 19]. In order to further
42 investigate whether MSCs were exerting their effects through induction of EMT, a
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4 number of specific genetic markers of EMT were examined in breast cancer cells
5 following co-culture. Anti-apoptotic transcription factors, Twist and Snail, and
6 mesenchymal protein markers, Vimentin and N-cadherin have been consistently
7 associated with mesenchymal transition in epithelial cells [21, 30]. Vimentin upregulation
8 is commonly observed in more invasive basal cancer subtypes and has been positively
9 correlated with poor prognosis in breast cancer patients [31]. Interestingly, Vimentin was
10 upregulated in both T47D and SK-Br-3 cells with no significant upregulation in the
11 MDA-MB-231 breast cancer population. This may be due to the relatively high
12 expression of Vimentin already present in the more invasive MDA-MB-231 cells [32].
13 This upregulation in Vimentin was confirmed at the protein level, and also detected
14 following 7days of in vitro co-culture. Significant upregulation of N-cadherin, Twist and
15 Snail was recorded across all breast cancer cells retrieved from co-culture with MSCs
16 although to a lesser extent in the MDA-MB-231 cells. This proportional difference in
17 EMT changes recorded between MDA-MB-231 cells and other less invasive breast
18 cancer subtypes coincides with findings recorded by Karnoub et al [14] who noted that
19 MDA-MB-231 cells exist in a state of “partial EMT” and that, within their study, CCL5
20 secreted by MSCs did not lead to advancement of this EMT phenotype.

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22 Further analysis of array data confirmed upregulation of a number of genes associated
23 with EMT induction including TGF β R2 [33] and ACVR1 [34], both receptors for TGF β ,
24 which is known to stimulate mesenchymal transition in epithelial cells [35]. Research
25 suggests that upregulated expression of TGF β R2 is an absolute requirement for TGF β
26 mediated EMT [36]. Vascular endothelial growth factor (VEGF), typically associated
27 with angiogenesis, was also upregulated. Non-Angiogenic functions of VEGF include
28 anti-apoptotic and pro-migratory properties [37] as well as an important role in the
29 initiation of EMT through upregulation of Snail expression [38]. EMT appears to be at
30 least partly dependant on VEGF signalling as studies that have blocked VEGF noted a
31 proportional decrease in EMT [39].

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33 To investigate whether changes seen were specific to MSCs, breast cancer cells were
34 directly cultured with normal fibroblasts (WI-38 cells), resulting in no significant change
35 in EMT related gene expression. This suggests the effects observed were MSC specific.
36 To assess whether changes in gene expression were mediated solely through cell to cell
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4 contact, breast cancer cells were also cultured in MSC conditioned medium. No change
5 in expression of EMT markers was seen in MDA-MB-231 cells exposed to MSC
6 conditioned medium. Both T47D and SK-Br3 cells exhibited a relatively mild upregulation
7 in expression of Twist, Snail and N-cadherin, with the most marked increase seen in
8 Vimentin expression. Although significant, these changes following indirect co-culture
9 occurred to a much lesser degree than those seen in cells directly cultured with MSCs.
10 Also, the effects were found to be transient in the indirect co-culture model used, with
11 gene expression returning to baseline following 7 days of indirect co-culture. This may be
12 due to cell-contact mediated inhibition of MSC proliferation, and resultant reduction in
13 secretion of mediating factors. Overall the data suggests that changes in gene expression
14 observed were predominantly mediated through direct cell to cell contact.

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24 Decreased expression of the cell adhesion protein E-cadherin and the resultant cellular
25 dissociation is another marker consistent with the process of EMT [18]. Previous studies
26 investigating breast cancer cells directly co-cultured with MSCs have shown a significant
27 downregulation in E-cadherin protein expression in breast cancer cells [7, 8] an
28 observation also noted in the current study. Dual staining to distinguish between cell
29 populations in co-culture also highlighted alterations in morphology and growth patterns
30 of breast cancer cells. T47D cells appeared to lose adhesiveness and separate from their
31 normal clustered growth pattern, with cells adjacent to MSCs branching and polarizing
32 toward the mesenchymal cells. These changes coincide with the loss of apico-basal
33 polarity seen in cells that undergo EMT [18].

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42 Recent literature has significantly advanced our understanding of the pivotal role EMT
43 plays in the metastatic cascade. Initially regarded with a degree of scepticism,
44 mesenchymal transition has been observed at the primary tumour site in a cohort of 479
45 human breast cancer samples and a positive correlation with basal breast cancer
46 phenotype confirmed [20]. Despite these developments the stimulus inducing EMT at the
47 primary tumour site remains unknown. The current study suggests that MSCs that are
48 actively recruited to tumour stromal microenvironments may act as a stimulus to induce
49 EMT in breast cancer cells and actively increase breast cancer metastatic potential.
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9

10 11 12 13 14 15 **References**

- 16 1. Jemal A, Siegel R, Ward E et al (2008) Cancer statistics, 2008. *CA Cancer J Clin*
17 58:71-96.
 - 18 2. Coleman RE, Rubens RD (1987) The clinical course of bone metastases from breast
19 cancer. *Br J Cancer* 55:61-66.
 - 20 3. Espy DK, Wu XC, Swan J et al (2007) Annual report to the nation on the status of
21 cancer, 1975-2004, featuring cancer in American Indians and Alaska Natives. *Cancer*
22 110:2119-2152.
 - 23 4. Bhowmick NA, Moses HL (2005) Tumor-stroma interactions. *Curr Opin Genet Dev*
24 15:97-101.
 - 25 5. Hu M, Polyak K (2008) Molecular characterisation of the tumour microenvironment in
26 breast cancer. *Eur J Cancer* 44:2760-2765.
 - 27 6. Pittenger MF, Mackay AM, Beck SC et al (1999) Multilineage potential of adult
28 human mesenchymal stem cells. *Science* 284:143-147.
 - 29 7. Hombauer H, Minguell JJ (2000) Selective interactions between epithelial tumour cells
30 and bone marrow mesenchymal stem cells. *Br J Cancer* 82:1290-1296.
 - 31 8. Fierro FA, Sierralta WD, Epanan MJ et al (2004) Marrow-derived mesenchymal stem
32 cells: role in epithelial tumor cell determination. *Clin Exp Metastasis* 21:313-319.
 - 33 9. Sasser AK, Mundy BL, Smith KM et al (2007) Human bone marrow stromal cells
34 enhance breast cancer cell growth rates in a cell line-dependent manner when evaluated
35 in 3D tumor environments. *Cancer Lett* 254:255-264.
 - 36 10. Chen J, Zhang ZG, Li Y et al (2003) Intravenous administration of human bone
37 marrow stromal cells induces angiogenesis in the ischemic boundary zone after stroke in
38 rats. *Circ Res* 92:692-699.
 - 39 11. Spaeth E, Klopp A, Dembinski J et al (2008) Inflammation and tumor
40 microenvironments: defining the migratory itinerary of mesenchymal stem cells. *Gene*
41 *Ther* 15:730-738.
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12. Dvorak HF (1986) Tumors: wounds that do not heal. Similarities between tumor stroma generation and wound healing. *N Engl J Med* 315:1650-1659.
13. Kumar S, Chanda D, Ponnazhagan S (2008) Therapeutic potential of genetically modified mesenchymal stem cells. *Gene Ther* 15:711-715.
14. Karnoub AE, Dash AB, Vo AP et al (2007) Mesenchymal stem cells within tumour stroma promote breast cancer metastasis. *Nature* 449:557-563.
15. Dwyer RM, Potter-Beirne SM, Harrington KA et al (2007) Monocyte Chemoattractant Protein-1 (MCP-1) secreted by primary breast tumors stimulates migration of Mesenchymal Stem Cells (MSCs). *Clin Cancer Res* 13:5020-5027.
16. Molloy AP, Martin FT, Dwyer RM et al (2009) Mesenchymal stem cell secretion of chemokines during differentiation into osteoblasts, and their potential role in mediating interactions with breast cancer cells. *Int J Cancer* 124:326-332.
17. Nakaya Y, Sheng G (2008) Epithelial to mesenchymal transition during gastrulation: an embryological view. *Dev Growth Differ* 50:755-766.
18. Thiery JP, Sleeman JP (2006) Complex networks orchestrate epithelial-mesenchymal transitions. *Nat Rev Mol Cell Biol* 7:131-142.
19. Brabletz T, Jung A, Spaderna S et al (2005) Opinion: migrating cancer stem cells - an integrated concept of malignant tumour progression. *Nat Rev Cancer* 5:744-749.
20. Sarrío D, Rodríguez-Pinilla SM, Hardisson D et al (2008) Epithelial-mesenchymal transition in breast cancer relates to the basal-like phenotype. *Cancer Res* 68:989-997.
21. Mani SA, Guo W, Liao MJ et al (2008) The epithelial-mesenchymal transition generates cells with properties of stem cells. *Cell* 133:704-715.
22. Barry FP, Murphy JM (2004) Mesenchymal stem cells: clinical applications and biological characterization. *International Journal of Biochemistry & Cell Biology* 36:568-584.
23. Livak KJ, Schmittgen TD (2001) Analysis of relative gene expression data using real-time quantitative PCR and the $2^{-\Delta\Delta C(T)}$ Method. *Methods* 25:402-408.
24. Woelfle U, Breit E, Pantel K (2005) Influence of immunomagnetic enrichment on gene expression of tumor cells. *J Transl Med* 3:12.
25. Corcoran KE, Trzaska KA, Fernandes H et al (2008) Mesenchymal stem cells in early entry of breast cancer into bone marrow. *PLoS ONE* 3:e2563.

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26. Molloy AP, Martin FT, Dwyer RM et al (2008) Mesenchymal stem cell secretion of chemokines during differentiation into osteoblasts, and their potential role in mediating interactions with breast cancer cells. *Int J Cancer*.
27. Gelmini S, Mangoni M, Serio M et al (2008) The critical role of SDF-1/CXCR4 axis in cancer and cancer stem cells metastasis. *J Endocrinol Invest* 31:809-819.
28. Eferl R, Wagner EF (2003) AP-1: a double-edged sword in tumorigenesis. *Nat Rev Cancer* 3:859-868.
29. Pinilla SM, Honrado E, Hardisson D et al (2006) Caveolin-1 expression is associated with a basal-like phenotype in sporadic and hereditary breast cancer. *Breast Cancer Res Treat* 99:85-90.
30. De Wever O, Pauwels P, De Craene B et al (2008) Molecular and pathological signatures of epithelial-mesenchymal transitions at the cancer invasion front. *Histochem Cell Biol* 130:481-494.
31. Thomas PA, Kirschmann DA, Cerhan JR et al (1999) Association between keratin and vimentin expression, malignant phenotype, and survival in postmenopausal breast cancer patients. *Clin Cancer Res* 5:2698-2703.
32. McInroy L, Maatta A (2007) Down-regulation of vimentin expression inhibits carcinoma cell migration and adhesion. *Biochem Biophys Res Commun* 360:109-114.
33. Galliher AJ, Schiemann WP (2006) Beta3 integrin and Src facilitate transforming growth factor-beta mediated induction of epithelial-mesenchymal transition in mammary epithelial cells. *Breast Cancer Res* 8:R42.
34. Mercado-Pimentel ME, Runyan RB (2007) Multiple transforming growth factor-beta isoforms and receptors function during epithelial-mesenchymal cell transformation in the embryonic heart. *Cells Tissues Organs* 185:146-156.
35. Derynck R, Akhurst RJ, Balmain A (2001) TGF-beta signaling in tumor suppression and cancer progression. *Nat Genet* 29:117-129.
36. Han G, Lu SL, Li AG et al (2005) Distinct mechanisms of TGF-beta1-mediated epithelial-to-mesenchymal transition and metastasis during skin carcinogenesis. *J Clin Invest* 115:1714-1723.
37. Mercurio AM, Lipscomb EA, Bachelder RE (2005) Non-angiogenic functions of VEGF in breast cancer. *J Mammary Gland Biol Neoplasia* 10:283-290.
38. Wanami LS, Chen HY, Peiro S et al (2008) Vascular endothelial growth factor-A stimulates Snail expression in breast tumor cells: implications for tumor progression. *Exp Cell Res* 314:2448-2453.

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6 39. Enciso JM, Gratzinger D, Camenisch TD et al (2003) Elevated glucose inhibits
7 VEGF-A-mediated endocardial cushion formation: modulation by PECAM-1 and MMP-
8 2. J Cell Biol 160:605-615.
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12 **Figure legends**
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16 **Table 1:** Results from Low Density Array analysis of breast cancer cells retrieved
17 following co-culture with MSCs. Results presented show genes where at least one cell
18 line had ≥ 2.5 fold increase or decrease in expression following co-culture with MSCs.
19 *Genes which were then validated in triplicate experiments by RQ-PCR.
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25 **Figure 1:** Change in breast cancer cell gene expression following co-culture with MSCs.
26 Results presented as Mean \pm SEM Log₁₀ Relative Quantity in triplicate experiments. The
27 baseline represents the level of expression in breast cancer cell lines cultured
28 individually. *denotes p<0.05
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34 **Figure 2:** Changes in proliferation of breast cancer cells exposed to factors secreted by
35 MSCs for 72hrs. Cell proliferation was measured using an Apoglow® assay in T47D,
36 SK-Br3 & MDA-MB-231 cells cultured alone and in MSC conditioned media. Results
37 presented represent mean of triplicate experiments \pm SEM. RLU = Relative Light Units
38 detected on luminometer. * p<0.05 **p<0.001
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45 **Figure 3: (A)** Changes in breast cancer cell expression of specific EMT genes following
46 co-culture with MSCs. The baseline represents the level of expression in breast cancer
47 cell lines cultured individually. Results presented represent mean of triplicate
48 experiments \pm SEM. * denotes p<0.05 (B) Protein analysis: Western blot of Sk-Br3 and
49 T47D cells alone and following direct co-culture with MSCs, targeting Vimentin and
50 Snail. β -actin was used to confirm uniform sample loading.
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58 **Figure 4:** Changes in expression of EMT genes in breast cancer cells following culture in
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4 MSC conditioned medium (Indirect co-culture). The baseline represents the level of
5 expression in breast cancer cell lines cultured in the same medium that had not been
6 exposed to MSCs. Results presented represent mean of triplicate experiments \pm SEM. No
7 change in expression of EMT markers was observed in MDA-MB-231 cells following
8 indirect co-culture (results not shown).
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15 **Figure 5 (a-g):** Immunostaining of breast cancer cells and MSCs cultured individually
16 and in direct co-culture on chamber slides for 72hrs.
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18 E-cadherin staining: (a) T47D cells, (b) T47D + MSCs, (c) Confirmation of reduced E-
19 cadherin by western blot of protein lysates from T47D cells cultured individually and
20 those retrieved following direct co-culture with MSCs. β -actin was used to confirm
21 uniform sample loading.
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26 Dual staining with cell type specific antibodies; Red – epithelial specific pancytokeratin,
27 Blue – haemotoxylin stained nuclei. (c) MDA-MB-231 cells, (d) MDA-MB-231 +
28 MSCs.
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31 Confocal fluorescent images of cell cytoskeletons stained with Alexafluor (green).
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33 Epithelial cells were also labeled with PKH26 (red) prior to mixing: (e) T47D cells (f)
34 T47D + MSCs. All images presented are at 200x magnification.
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Fig. 1: Change in gene expression in breast cancer cells retrieved following direct co-culture with MSCs.

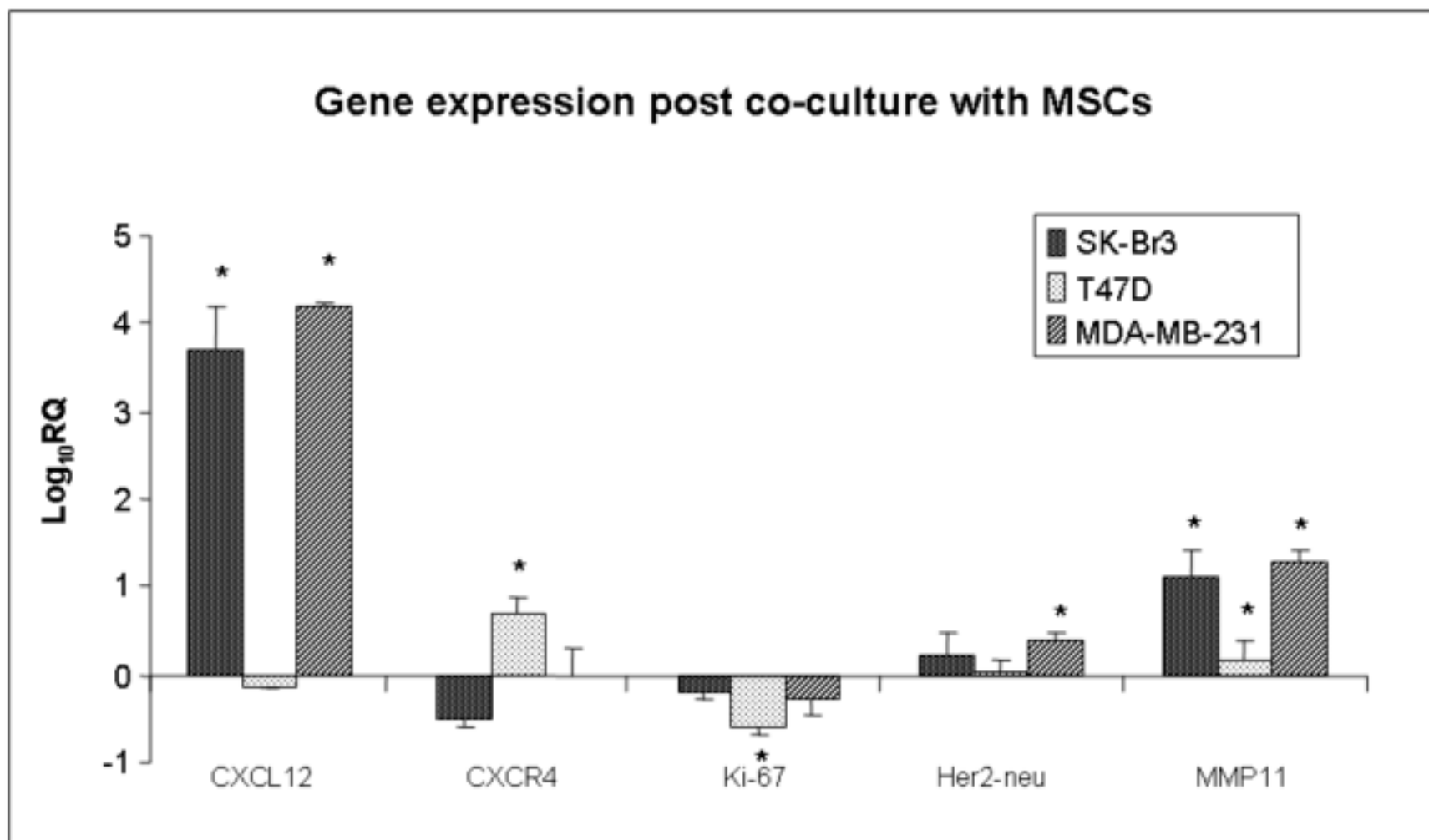


Fig. 2: Changes in proliferation of breast cancer cells exposed to factors secreted by MSCs for 72hrs

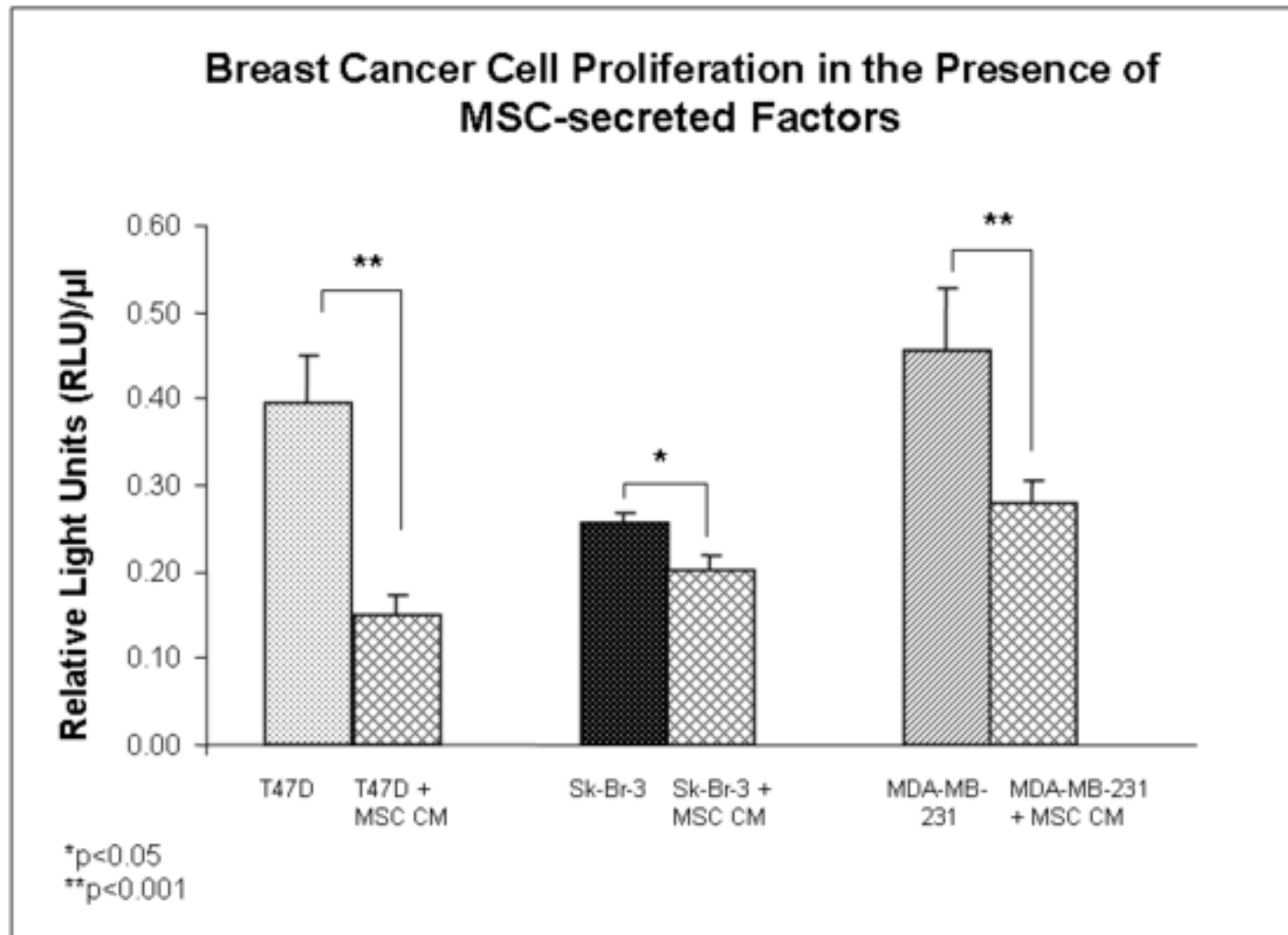


Fig. 3: (A) Changes in expression of EMT specific genes in breast cancer cells retrieved following direct co-culture with MSCs. (B) Representative samples showing increases in expression at the protein level by western blot.

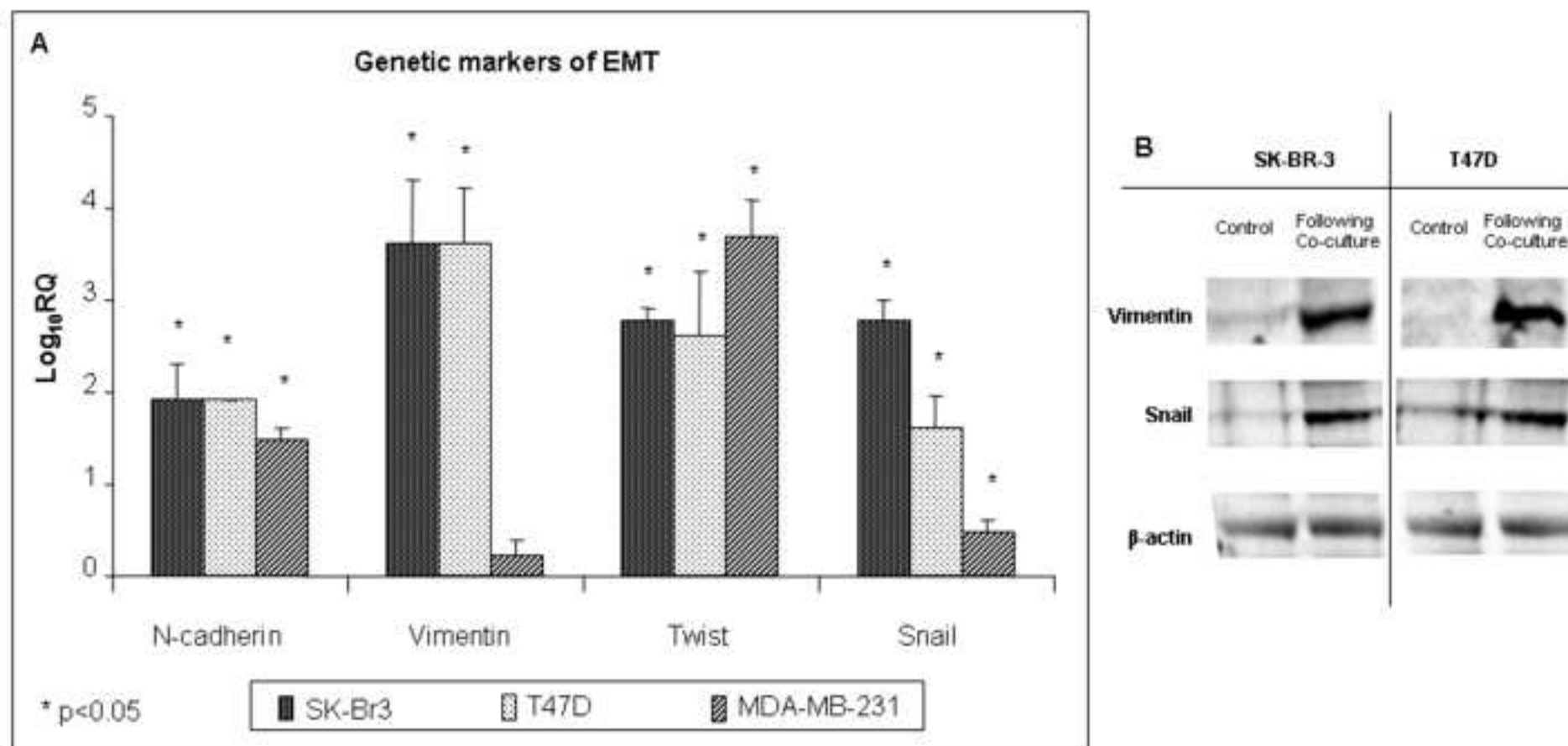


Fig. 4: Changes in expression of EMT genes in breast cancer cells following culture in MSC conditioned medium (indirect co-culture).

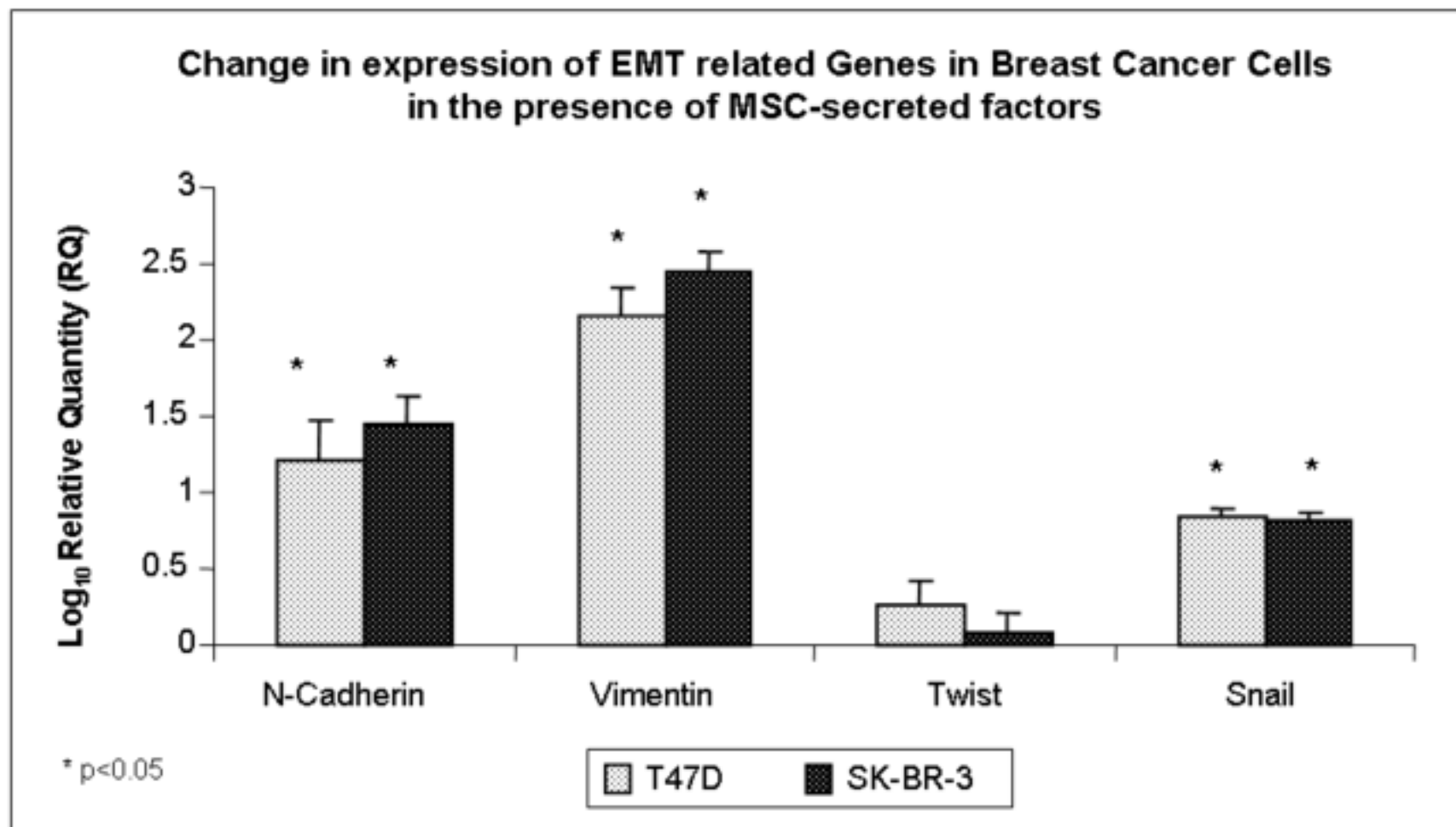


Fig. 5

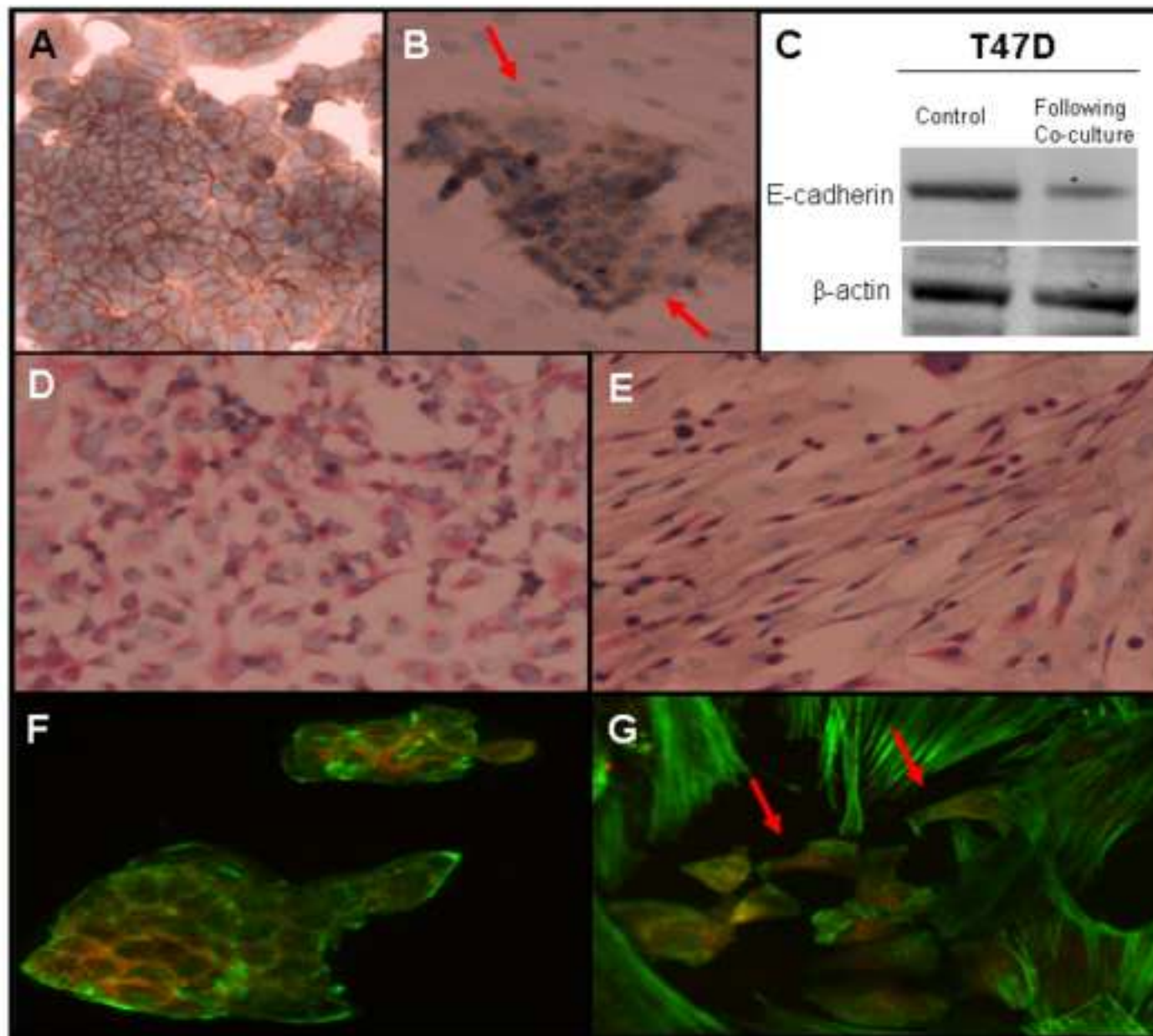


Table 1[Click here to download Table: Table 1.doc](#)

Gene Function	Gene Symbol	Gene Name	Fold Change		
			Sk-Br3	T47D	MDA-MB-231
Oncogene & proto-oncogenes	NCOA4	Nuclear receptor co-activator 4	3.7	3.7	N/A
	FOS	Osteosarcoma oncogene	6.1	38.9	5.5
	MUC1	Mucin 1	6.1	1.6	N/A
	FYN	FYN oncogene	11	17.6	1.9
	JUN	JUN oncogene	3.2	3.4	2.7
	MET	MET proto-oncogene	3.3	15.4	N/A
	EPHA2	Ephrin receptor A2	2.3	5.5	1.5
Macrophage marker	CD68*	CD68 macrophage antigen/microsialin	8.1	35	3.7
Angiogenesis	VEGF	Vascular endothelial growth factor A	7.3	12	N/A
Anti-apoptosis	IGF1R	Insulin-like growth factor 1 receptor	3.5	1.2	2
	BCL2*	B-cell CLL/lymphoma 2	9.4	6.3	N/A
	CAV-1*	Caveolin 1	8.5	3994	1.2
EMT induction	TGFBR2*	Transforming growth factor-beta receptor type II	8.7	2142	1.8
	ACVR1*	Activin A receptor type 1/ TGF beta superfamily receptor 1	6.7	3.3	3.3
Proliferation	CCNE1	Cyclin E1	-2.8	-2.1	N/A
	MKi67*	Antigen identified by monoclonal antibody Ki-67	-3.3	-4.9	-2.4
	MYBL2	Myeoblastosis oncogene	-5.5	-4.4	-2
Invasion & Migration	MMP11*	Matrix metalloproteinase 11 (Stromelysin 3)	15.2	2.9	20
	CXCL12*	Stromal cell-derived factor 1	9,949	-1.2	17,066

Table 1: Low Density Array analysis of breast cancer cells retrieved following co-culture with MSCs. * selected genes validated in triplicate experiments by RQ-PCR